

Review Paper

A Review on the Antimicrobial Activity of an Important Medicinal Plant *Inularacemosa* Hook. F.

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Abstract

Microbial infections whether in humans or in plant have posed a great challenge in front of us. The emergence of microbial resistance to the currently available anti-microbial agents necessitates the further research in the discovery of new safe and effective anti-microbial agents. The medicinal plants have shown a promising alternative for the treatment of infectious diseases. Many plant-derived products such as phenols, quinones, terpenes, saponins and alkaloids have been used safely for antimicrobial purposes in the Indian and Chinese traditional medicine with little risk of animal toxicities. For this reason, some phytochemicals are receiving growing attention aiming at developing hopefully new indigenous, nontoxic, biodegradable and renewable pesticides. The root of *Inula racemosa* Hook f. is a traditional medicinal herb frequently used as antimicrobial agents since ancient times. Phytochemical investigation of the plant showed the presence of alantolactone, isoalantolactone, dihydroalantolactone, dihydroisoalantolactone, sitisterol, daucosterol, inunolide, apilotaxene, phenylacetone nitrile and iso-inulin. Recently, the active compound of antimicrobial activity characterized as isoalantolactone, which happens to be the most abundant constituent of the plant. Available literature indicates that the constituents of *Inula racemosa* Hook. F. and the plant extract have shown a great success as an antimicrobial agent both in human and plants.

Key words: Microbial infection, anti-microbial agents, phytochemicals, *Inula racemosa* Hook f.

Introduction

Antimicrobial activity of Inula racemosa Hook. f. in humans

Synthetic pesticides are widely applied to plant protection. Many of such chemicals, not easily degradable in field, are suspected of environmental pollution, leading to residual toxicity such as carcinogenicity and teratogenicity (1). Furthermore, repetitious applications of these agrochemicals make pests develop resistance (2). Accordingly, there is an urgent need to search for new highly effective and safe (or easily biodegradable) chemicals that will be expected to be more advantageous in plant protection. Many plant-derived products such as phenols, quinones, terpenes, saponins and alkaloids have been used safely for antimicrobial purposes in the Indian and Chinese traditional medicine with little risk of animal toxicities (3, 4). For this reason, some phytochemicals are receiving growing attention aiming at developing hopefully new indigenous, nontoxic, biodegradable and renewable pesticides (5-7). The root of *Inula racemosa* Hook f. (Compositae) is a traditional Chinese medicinal herb (called Tumuxiang) frequently used as antimicrobial agents since ancient times. Phytochemical investigation of the plant showed the presence of sesquiterpene lactones like alantolactone, isoalantolactone, dihydroalantolactone, dihydroisoalantolactone, sitisterol, daucosterol, inunolide, apilotaxene, phenylacetone nitrile and iso-inulin which shows high antimicrobial activity whether in plants or in humans (8).

According to the study, isoalantolactone, a major constituent of *Inula racemosa* shows high activity against microbes. It was tested for its antimicrobial activity against five bacteria, six human and six plant pathogenic fungi. The lactone showed absolute toxicities at 500 µg/mL against 3 soil borne phytopathogenic fungi (*Gaeumannomyces graminis* var. *tritici*, *Rhizoctonia cerealis* and *Phytophthora capsici*) with the MICs determined to be 100, 100 and 300 µg/mL, respectively. At the MICs, isoalantolactone exhibited its fungistatic nature of toxicity. The lowest fungicidal concentrations of the lactone to *G. graminis* var. *tritici*, *R. cerealis* and *P. capsici* were shown to be 150, 150 and 350 µg/mL, respectively. Moreover, isoalantolactone displayed weaker antibacterial activities against *Bacillus subtilis*, *Escherichia coli*, *Pseudomonas fluorescense*, *Sarcinulentus* and *Staphylococcus aureus* with MICs of 125, 425, 150, 150 and 100 µg/mL, respectively. The investigation disclosed the strong inhibition of isoalantolactone to the phytopathogenic fungi, raising a possibility that the lactone could be considered as a starting point for the project aiming at the development of new fungicides (9). Some of the sesquiterpene lactones of *Inula racemosa* has the pharmacological activities (10). Alantolactone and isoalantolactone are the major constituent of *Inula racemosa*, they possess anti-fungal and anti-helminthic activities (11). Isoalantolactone isolated from the methanol roots extract of *I. racemosa* was evaluated for antifungal activity against the human pathogenic fungi *Aspergillus flavus*, *Aspergillus niger*, *Geotrichum candidum*,

Candida tropicalis and *Candida albicans*. The tested compound inhibited the growth of *A. niger*, *A. flavus*, *G. candidum*, *C. albicans* and *C. tropicalis* with MICs values 50, 50, 25, 25 and 25 $\mu\text{g mL}^{-1}$ respectively (12).

Antibacterial activity of the ethanol and aqueous roots extract of *I. racemosa* was evaluated by disc diffusion method against *E. coli* and *S. aureus*. The aqueous extract of the plant exhibited significant antimicrobial activity for these two microorganisms tested, with MIC values of 6.25 mg mL^{-1} and 12.5 mg mL^{-1} respectively, whereas ethanol extract also had potent activity against microorganisms, with MIC of 15.625 mg mL^{-1} (13). The resistance of different bacteria to the current antibacterial agents, toxicity or the antibacterial agents and the cost of the treatment have led to the development of new active molecules against the bacteria. Since, ancient time's medicinal plants have been used for the treatment of bacterial infections. The roots of the plant *Innula racemosa* has been used as folk medicine in East Asia and Europe. However, no systematic data is available on the antibacterial activity profile of different constituents of *Innula racemosa*. In the present studies, attempt has been made for the isolation of root constituents of *Innula racemosa* and evaluation of its antibacterial activity. The constituents were isolated and purified by column chromatography. The structure of the isolated constituents was confirmed by the spectral analysis and was used for the determination of the antibacterial activity of *Innula racemosa* against various microorganisms. The constituent's alantolactone showed maximum antibacterial activity as compared to the other constituents and ethyl acetate extract of roots (14).

Antimicrobial activity of *Inula racemosa* Hook. f. in Plants:

Plant pathogens cause great losses to agricultural crops and thus threaten food resources all over the world (15). A significant portion of the agricultural produce in the country and the world over become unfit for human consumption due to mycotoxins contamination produced by species of *Aspergillus* (16, 17, 18), *Aspergillus niger* as a strong pathogen can cause the rotting of numerous fruits and vegetable (19, 20) (21 22) such as black rot of onion and rot of tomato (23, 24). *Aspergillus flavus* is a weak opportunistic and a mycotoxigenic fungus and infects several agricultural crops (25). These pathogenic fungi not only involve in food spoilage by retarding its nutritive value (26, 27) but also involve in many fungal infections. The toxin produced by them shows the effect of carcinogenicity, genotoxicity, teratogenicity, nephrotoxicity, hepatotoxicity, reproductive disorders (28, 29, 30) on consumption of spoiled food. The disease in crop caused by phytopathogenic fungi is mainly controlled by synthetic fungicides. However, the application of this agrochemical is increasingly restricted due to the harmful effects of pesticides on human health and the environment (31). In many countries the use of synthetic fungicide has been banned due to the non-biodegradability, residual toxicity, pollutive nature. In order to have safe methods for plant disease control in sustainable agriculture there is a need for reducing the use of synthetics chemicals fungicides there by replacing it with biocides with plant origin. Plants are known to produce a variety of compounds to protect themselves against a variety of pathogens. The plants are rich sources of numerous bioactive secondary metabolites such as alkaloid, flavonoids, terpenoids, saponins, tannins and phenolic compounds which are the important sources of microbiocides, pesticides, antifungal activity and many pharmaceutical compounds (32, 33). Thus plant extracts which is a source of natural pesticides can be developed into new biocidal pesticides (34, 35). Therefore the present study investigate; the efficacy of various high altitude medicinal plant extracts from *Codonopsis clematidea*, *Hippo pharhamnoides*, *Artemisia dracunculus*, *Galium aparine*, *Mentha longifolia*, *Foeniculum vulgare*, *Rubia cordifolia*, *Saussurea lappa*, *Inula racemosa*, *Rheum webbianum*, *Arnebia euchroma*, *Rhodia laheterodanta*, *Rhodia laimbericate*, *Achelia millefolium* and *Hypericum perforatum*, for the thirst of antifungal property against the two *Aspergillus spp.*, through *in vitro* analysis. Recent efforts have focused on development of environmentally safe, long lasting and effective biocontrol methods for the management of plant diseases.

Alternaria solani causes early blight in plants belonging to family Solanaceae. It severely infests agriculturally important crops like tomato and potato at different developmental stages. We have screened eighty one extracts of twenty seven medicinally important plants for their antifungal activity against *A. solani*. Twelve extracts of eight plants (*Cinnamomum zeylanicum*, *Syzygium aromaticum*, *Ferula foetida*, *Inula racemosa*, *Hemides musindicus*, *Rubia cordifolia*, *Glycyrrhizaglabra* and *Saussurea lappa*) possessed marginal to excellent antifungal activity. Hexane (Hx) and methanol (MeOH) extracts of *C. zeylanicum* showed complete inhibition of *A. solani in vitro* at a dose of 3 ml/lit water. Formulation trials conducted in shade house using MeOH extract of *C. zeylanicum* at a dose of 2 ml/lit water were highly effective against *A. solani* infesting tomato plants. TLC, HPTLC and GC-MS analysis confirmed the presence of Eugenol, Cinnamaldehyde and 2H -1- Benzopyran -2- one in MeOH extract of *C. zeylanicum*. These secondary metabolites were isolated by preparative TLC to further confirm antifungal activity and probable structures by GC-MS. 2H-Benzopyran-2-one, a coumarin has been reported for the first time as an antifungal against *A. solani* in present study. An effective lab-scale formulation was developed and tested against *A. solani* (36).

The crude oils from roots of *Inula racemosa* Hook.f. and *Saussurea lappa* Clarke at four concentrations (from 0.625 to 5%) significantly reduced the leaf damage done by *Spodoptera litura* (Fabricius) larvae. The rate of feeding deterrence measured after treatment with oils of *I. racemosa* and *S. Lappa* at these concentrations ranged from 20 to 72% and 18 to 62%, respectively. The rate of mortality during larval and prepupal stages was greater than 50% at the 5% concentration. Pupation was adversely affected only at the highest concentration of both oils. Pupal weight was normal, while adult emergence was marginally affected by both oil treatments (37).

Mosquito Larvicidal Activity of *Inula racemosa* Hook. f.:

Qing et al isolated 11,13-dihydroisoalantolactone, macrophyllilactone E, 5- α -epoxyalantolactone and epoxyisoalantolactone from the ethanol roots extract of *I. racemosa*. Mosquito larvicidal activity of all these isolated compounds was evaluated against the

larvae of *Aedes albopictus* and *Asian tiger* mosquitoes. The tested compound 11,13-dihydroisoalantolactone and macrophyllilactone E exhibited strong larvicidal activity against the early fourth-instar larvae of *A. albopictus* with LC₅₀ values of 21.86 µg mL⁻¹ and 18.65 µg mL⁻¹ respectively, whereas 5-α-epoxyalantolactone and epoxyisoalantolactone also possessed larvicidal activity against the *Asian tiger* mosquitoes with LC₅₀ values of 29.37 µg mL⁻¹ and 35.13 µg mL⁻¹ respectively (38).

Conclusion

Microbial infections whether in plants or in humans have posed a great challenge in front of us. The emergence of microbial resistance to the currently available agents necessitates the further research in the discovery of new safe and effective antimicrobial agents. The medicinal plants have shown a promising alternative for the treatment of infectious diseases. In the antimicrobial research, the vast majority of the new chemical entities are natural or natural product-derived molecules. *Inula racemosa* has been used as anti-microbial agents since ages as it has several phytochemicals which shows high anti-microbial activity both in humans and plants. Phytochemicals using as pesticides is the most safest and environment friendly method because they are non-toxic and biodegradable in nature.

References

1. Shashikant, V. S., Sahai, Y. N., Trends in Environmental Pollution and Pesticide Toxicology, New Delhi: Jagmander Book Agency, 1989, 375.
2. Spotts, R. A., Cervantes, L. A., Population pathogenicity and benomyl resistance of *Botrytis* spp. *Penicillium* spp. and *Mucor piriformis* in packing house, Plant Disease, 1986, 70: 106.
3. Jiangsu College of New Medicine, A Comprehensive Dictionary of the Traditional Chinese Medicines (Zhong Yao Da Ci Dian), Shanghai: Shanghai People's Press, 1977, 81.
4. Kirtikar, K. R., Basu, B. D., Indian Medicinal Plants, Dehradun: M/S Bisan Singh Mahendra Pal Sirgh, 1975.
5. Mahadevan, A., Biochemical Aspects of Plant Disease Resistance, Part I: Preformed Inhibitory Substances Prohibition, New Delhi: Today and Tomorrow's Printers and Publishers, 1982.
6. Mishra, A. K., Dubey, N. K., Evaluation of some essential oils for their toxicity against fungi causing deterioration of stored food commodities, Applied and Environmental Microbiology, 1994, 60 (3): 1001.
7. Wain, R. L., Chemical aspects of plant disease resistance, in Natural Products and the Protection of Plants (ed. Marini-Bettalo, G. B.), New York: Elsevier Scientific Publishing Co., 1977, 483.
8. Wang K., Liu H, Zhao Y, Chen X, Song Y and Ma X, 2000. Separation and determination of alantolactone and isoalantolactone in traditional Chinese herbs by capillary Electrophoresis Talanta, 52 :1001-1005.
9. Mishra A. K., Changhong, L., Bing H.E. and Renxiang T. (2001). Antimicrobial activities of isoalantolactone, a major sesquiterpene lactone of *Inularacemosa*. *Chinese Science Bulletin*, 46(6).
10. Tripathi, Y.B., Tripathi, P. and Upadhyay, B.N., 1988. Assessment of the adrenergic beta-blocking activity of *Inularacemosa*. *J. Ethnopharmacol.*, 23:3-10.
11. Satyavati, G.V., Gupta, A.K. and Tandon, N. 1987. Medicinal plants of India. Indian Council of Medical Res., New Delhi, Vol. II.
12. Tan, R.X., Tang, H.Q., Hu, J. and Shuai, B. (1998). Lignans and sesquiterpene lactones from *Artemisia -sieversiana* and *Inularacemosa*, *Phytochem.* 49(1):157-161.
13. Y.Q. Yang, H.L. Ma, Z.X. Zhang, Study on medicinal constituents of cultivated *Inularacemosa*, Master's thesis, Henan University of Science and Technology, China, 2009.
14. Lokhade P.D., Gawai K.R., Chabuskwar A.R. Jgadali S.C. and Kuchekar B.S. (2007). Antibacterial activity of isolated constituents and extract of roots of *Inularacemosa* Hook. F. *Research Journal of Medicinal Plant*, 7(1):7-1.
15. Singh R.P., Singh R., Ram, P., Batliwala, P.G., (1993). Use of Pushkarguggal, an indigenous, antischaemic combination in the management of ischaemic heart disease. *International Journal of Pharmacology*, 31:147-160
16. Baniyadi F, Bonjar GHS, Karimi NA, Jorjandi M, Aghighi S, Farokhi PR (2009): Biological control of *Sclerotinia sclerotiorum*, causal agent of sunflower head and stem rot disease, by use of soil borne actinomycetes isolates. *AJABS* 4: 146–151
17. Janardhana, G.R., Raveesha, K.A. and Shetty, H.S. (1999). Mycotoxin contamination of maize grains grown in Karnataka (India). *Food Chemical Toxicology* 37: 863 – 868 3.
18. Chandra, R. and Sarbhoy, A.K. (1997) Production of Aflatoxins and Zearalenone by the toxigenic fungal isolates obtained from stored food grains of commercial crops. *Indian Phytopathology* 50: 458-68.
19. Devi, K.T., Mayo, M.A., Reddy, G., Emmanuel, K.E., Larondelle, Y. and Reddy, D.V.R. (2001). Occurrence of Ochratoxin A in black pepper, coriander, ginger and turmeric in India. *Food Additives Contamination* 18: 830 – 835.
20. Leong S, Hocking AD, Pitt JI (2004): Occurrence of fruit rot fungi (*Aspergillus section Nigri*) on some drying varieties of irrigated grapes. *Austral J Grape Wine Res* 10: 83–88.
21. Diedhiou, P.M., Mbaye, N., Dramé, A., Samb, P.I., (2007): Alteration of post harvest diseases of mango *Mangifera indica* through production practices and climatic factors. *Afr J Biotechnol* 6: 1087–1094
22. Fatima N, Batool H, Sultana V, Ara J, Ehteshamul- Haque S (2009): Prevalence of post-harvest rot of vegetables and fruits in Karachi, Pakistan. *Pakist J Bot* 41: 3185–3190.
23. Mathew S (2010): The prevalence of fungi on the post-harvested guava *Psidium guajava*. *Int J PharmacSci Res* 1: 145–149.
24. Narayana, K.J.P., M. Srikanth, M. Vijayalakshmi and N. Lakshmi, 2007. Toxic spectrum of *Aspergillus niger* causing black mold rot of onion. *Res. J. Microbiol.*, 2:881-884

25. Sinha P and SK Saxena, 1987. Effect of treating tomatoes with leaf extract of *Lantana camara* on development of fruit rot caused by *A. niger* in presence of *Drosophila busckii*. *Indian Journal of Experimental Biology* 25 143-144.
26. Bankole, S.A., B.M. Ogunsanwo, O.O. Mabekoje, 2004. Natural occurrence of moulds and aflatoxin B1 in melon seeds from markets in Nigeria. *Food and chemical toxicology* 42(8):1309-14.
27. Marin. S., Homedes, V., Sanchis, V., Ramos, A.J. and Magan, N. (1999). Impact of *Fusarium moniliforme* and *F. proliferatum* colonisation of maize on calorific losses and fumonisin production under different environmental conditions. *Journal of Stored Product Research* 35: 15 – 26.
28. Janardhana, G.R., Raveesha, K.A. and Shetty, H.S. (1998). Modified atmosphere storage to prevent mould induced nutritional loss in maize. *Journal of Science Food and Agriculture* 76: 573 – 578.
29. Lacey, J. (1988) The microbiology of cereal grains from areas of Iran with a high incidence of oesophageal cancer. *Journal of Stored Product Research* 24: 39-50.
30. Desjardins, A.E., Manandhar, G., Plattner, R.D., Maragos, C.M., Shrestha, K. and McCormick, S.P. (2000) Occurrence of *Fusarium* species and mycotoxins in Nepalese Maize and Wheat and the effect traditional processing method on mycotoxin levels. *Journal of Agricultural and Food Chemistry* 48: 1377-1383
31. Satish S, Mohana DC, Raghavendra MP, Raveesha KA. Antifungal activity of some plant extracts against important seed borne pathogens of *Aspergillus* sp. *J of Agric Tech*, 2007; 3(1): 109-119.
32. Harris, C.A., M.J. Renfrew, and M.W. Woolridge. 2001. Assessing the risk of pesticide residues to consumers: Recent and future developments. *Food Additives and Contamination* 18:1124-1129.
33. Mahesh B. and Satish S. (2008). Antimicrobial activity of some important medicinal plant against plant and human pathogens. *World Journal of Agricultural Sciences*, 4: 839-843.
34. Arif T, Bhosale JD, Kumar N, Mandal TK, Bendre RS, Lavekar GS, Dabur R. Natural products- antifungal agents derived from plants. *J Asian Natural Products Res*, 2009; 11(7): 621-63820.
35. Gangadevi, V., Yogeswari, S., Kamalraj, S., Rani, G and Muthumary, J. (2008). The antibacterial activity of *Acalypha indica* L. *Indian journal of Science and Technology*. 1(6).
36. Brindha, V., Saravanan, A and Manimekalai, R. (2009). Drug designing for ring finger protein 110 involved in adenocarcinoma (human breast cancer) using casuarinin extracted from *Terminalia arjuna*. *Indian Journal of Science and Technology*, 2(2): 22-26.
37. Mendki P. S., Teli, N. P., Kotkar, H. M. And Yeole, G. J. (2014). *Cinnamomum zeylanicum* extracts and their formulations control early blight of tomato. *JBiopest* 7(2):110-123.
38. Qing, H.E., Xin, L.C., Rui, S.Q., Deng, Z.W., Du, S.S., Liu, Z.L. (2014). Mosquito larvicidal constituents from the ethanol extract of *Inularacemosa* Hook. f. Roots against *Aedes albopictus*, *Journal of Chemistry* 1-6.