

ISBN: 978-81-963291-7-4

ISBN: 978-81-963291-7-4



Dr. Bhagwan Singh Patel, M.Sc., M.Phil., Ph.D., Assistant Professor, Maharaja Bhoj Govt. P.G. College (Dev) Ahilya Vishwavidyalaya Indore (M.P.), India. He has an experience of more than 14 years in teaching and research. Contribution in the field of Limnology and Biodiversity, from Zoological Society India. He has received Desh ka Gaurav Award 2023 (Lifetime Environment Society Somnath Nakshatra Vastika Trust Mathura (U.P.)), Member of Zoological Society of India, Gaya (Bihar).



Prof. Shailendra Sharma, MSc, PhD, DSc, Principal, Adarsh Institute of Management and Science, Dhamnod, Dr Sharma has 27 years of academic experience and 15 years of research experience. 98 of your research papers have been published in national and international journals. You have written 5 books and you are a member of many academic institutions as well as a member of the Working Council of the Zoological Council of India. You have been awarded by the Zoological Council of India for Lifetime Achievement. Similarly, you have been awarded by many national and international institutions for your amazing work in research. Under your guidance, 15 students have earned PhD degrees and as well as researchers have earned MPhil degrees. You have completed the research management of three UGCs research projects. You have also been awarded the fellowship of the European Commission. You have worked as a water scientist in IIT Roorkee, as well as a water policy expert and you are also known as an expert in nutrition and yoga.

REFERENCE BOOK

LIMNOLOGY

Author:
BHAGWAN SINGH PATEL

Co-Author:
Dr. SHAILENDRA SHARMA



CRDEEP PUBLICATIONS

☎ 011-43366666
✉ info@crdeeppublications.com
KALSHAN KANAK, SUMERGAU, UTTARAKHAND
www.crdeeppublications.com

CRDEEP PUBLICATIONS

REFERENCE BOOK

**LIMNOLOGICAL STUDIES ON MANSAROVAR
TALAB OF JEERAPURA, DHAR DISTRICT (M.P.)**

Author:

BHAGWAN SINGH PATEL

Co-Author:

Dr. Shailendra Sharma

CRDEEP Publications

Limnological Studies on Mansarovar talab of Jeerapura, Dhar District (M.P.)

No part of this book can be republished, reproduced or reprinted without prior permission of author or publisher

Copyright@CRDEEP Journals
First Ed. 2023

978-81-963291-7-4

Typesetting and Printing at: CRDEEP Publications, Dehradun

**“LIMNOLOGICAL STUDIES ON MANSAROVAR TALAB OF
JEERAPURA, DHAR DISTRICT (M.P.).”**

Author:

BHAGWAN SINGH PATEL

Co-Author:

Dr. Shailendra Sharma

CONTENTS

S. No.	CONTENTS	Page No.
1	INTRODUCTION	1-07
2	REVIEW OF LITERATURE	8-11
3	MATERIAL AND METHODS	12-45
4	OBSERVATIONS AND RESULTS	46-162
5	DISCUSSION	163-176
6	CONCLUSION	177-179
7	SUGGESTIONS FOR CONSERVATION	180-181
8	SUMMARY	182-194
9	REFERENCES	195-214
10	PUBLICATIONS	215-239
11	CONFRENCES/ SEMINARS	140-250

LIST OF TABLES

TABLE NO.	TABLE NAME	PAGE NO.
1.	Monthly Variations in water Temperature ($^{\circ}\text{C}$) in Mansarovar Talab from July 2014 to June 2015.	49
2.	Monthly Variations in water Temperature ($^{\circ}\text{C}$) in Mansarovar Talab from July 2015 to June 2016.	49
3.	Monthly Variations in Transparency (cm.) in Mansarovar Talab from July 2014 to June 2015.	53
4.	Monthly Variations in Transparency (cm.) in Mansarovar Talab from July 2015 to June 2016.	53
5.	Monthly Variations in Turbidity in Mansarovar Talab from July 2014- June 2015.	57
6.	Monthly Variations in Turbidity in Mansarovar Talab from July 2015- June 2016.	57
7.	Monthly Variations in pH in Mansarovar Talab from July 2014 to June 2015.	61
8.	Monthly Variations in pH in Mansarovar Talab from July 2015 to June 2016.	61
9.	Monthly Variations in Dissolved Oxygen (mg/l) in Mansarovar Talab from July 2014 to June 2015.	65
10.	Monthly Variations in Dissolved Oxygen (mg/l) in	65

	Mansarovar Talab from July 2015 to June 2016.	
11.	Monthly Variations in Total Solids (mg/l) in Mansarovar Talab from July 2014 to June 2015.	69
12.	Monthly Variations in Total Solids (mg/l) in Mansarovar Talab from July 2015 to June 2016.	69
13.	Monthly Variations in Free CO ₂ (mg/l) in Mansarovar Talab from July 2014 to June 2015.	73
14.	Monthly Variations in Free CO ₂ (mg/l) in Mansarovar Talab from July 2015 to June 2016.	73
15.	Monthly Variations in Total Alkalinity (mg/l) in Mansarovar Talab from July 2014 to June 2015.	77
16.	Monthly Variations in Total Alkalinity (mg/l) in Mansarovar Talab from July 2015 to June 2016.	77
17.	Monthly Variations in Biochemical Oxygen Demand (mg/l) in Mansarovar Talab from July 2014 to June 2015.	81
18.	Monthly Variations in Biochemical Oxygen Demand (mg/l) in Mansarovar Talab from July 2015 to June 2016.	81
19.	Monthly Variations in Total Hardness (mg/l) in Mansarovar Talab from July 2014 to June 2015.	85
20.	Monthly Variations in Total Hardness (mg/l) in Mansarovar Talab from July 2015 to June 2016.	85
21.	Monthly Variations in Magnesium (mg/l) in Mansarovar Talab from July 2014 to June 2015.	89
22.	Monthly Variations in Magnesium (mg/l) in Mansarovar	89

	Talab from July 2015 to June 2016.	
23.	Monthly Variations in Calcium (mg/l) in Mansarovar Talab from July 2014 to June 2015.	93
24.	Monthly Variations in Calcium (mg/l) in Mansarovar Talab from July 2015 to June 2016.	93
25.	Monthly Variations in Chloride (mg/l) in Mansarovar Talab from July 2014 to June 2015.	97
26.	Monthly Variations in Chloride (mg/l) in Mansarovar Talab from July 2015 to June 2016.	97
27.	Monthly Variations in Nitrate (mg/l) in Mansarovar Talab from July 2014 to June 2015.	101
28.	Monthly Variations in Nitrate (mg/l) in Mansarovar Talab from July 2015 to June 2016.	101
29.	Monthly Variations in Phosphate (mg/l) in Mansarovar Talab from July 2014 to June 2015.	105
30.	Monthly Variations in Phosphate (mg/l) in Mansarovar Talab from July 2015 to June 2016.	105
31.	Monthly Variations in Sulphate (mg/l) in Mansarovar Talab from July 2014 to June 2015.	109
32.	Monthly Variations in Sulphate (mg/l) in Mansarovar Talab from July 2015 to June 2016.	109
33.	Monthly Variations in Total Coli form (MPN/100 ml) in Mansarovar Talab from July 2014 to June 2015.	113
34.	Monthly Variations in Total Coli form (MPN/100 ml) in Mansarovar Talab from July 2015 to June 2016.	113

35.	Monthly Variations in Faecal Coli Form (MPN/100 ml) in Mansarovar Talab from July 2014- June 2015.	117
36.	Monthly Variations in Faecal Coli Form (MPN/100 ml) in Mansarovar Talab from July 2015- June 2016.	117
37.	Mothly variations in Diversity of Macro-invertebrates reported in Mansarovar Talab of Jeerapura (Dhar) Year July 2014 to June 2015.	119
38.	Mothly variations in Diversity of Macro-invertebrates reported in Mansarovar Talab of Jeerapura (Dhar) Year July 2015 to June 2016.	120
39.	List of Benthic Macro-invertebrates species reported in Mansarovar Talab from July 2014 to June 2016.	123
40.	Depthwise Status of Benthic Macro-invertebrates.	124-125
41.	Karl Pearsons coefficient of correlation between Physico-chemical parameters and Macro Invertebrates and Fishes at Station-I	132
42.	Karl Pearsons coefficient of correlation between Physico-chemical parameters and Macro Invertebrates and Fishes at Station-II	132
43.	Karl Pearsons coefficient of correlation between Physico-chemical parameters and Macro Invertebrates and Fishes at Station-III	133
44.	Karl Pearsons coefficient of correlation between Physico-chemical parameters and Macro Invertebrates and Fishes at Station-IV	133
45.	Seasonal Variations in fishes of Mansarovar Talab of	134

	Jeerapura (Dhar) Year July 2014 – June 2015.	
46.	Seasonal Variations in fishes of Mansarovar Talab of Jeerapura (Dhar) Year July 2015 – June 2016.	135
47.	Monthly Variations in Shannon & Weaver index and Simpson's Diversity Index during the study period (July 2014 to June 2016).	136
48.	Fish Species percentage.	138
49.	Seasonal Variations in fish species from July 2014 to June 2015.	139
50.	Seasonal Variations in fish species from July 2015 to June 2016.	140
51.	Water Quality Index-Analysed Status Station I & II (July 2014 to June 2016)	145
52.	Water Quality Index-Analysed Status Station III & IV (July 2014 to June 2016)	154

LIST OF FIGURES

FIGURE NO.	FIGURE NAME	PAGE NO.
01.	Graph showing Monthly Variations in water Temperature ($^{\circ}\text{C}$) in Mansarovar Talab from July 2014 to June 2015.	50
02.	Graph showing Monthly Variations in water Temperature ($^{\circ}\text{C}$) in Mansarovar Talab from July 2015 to June 2016.	50
03.	Graph showing Monthly Variations in Transparency (cm.) in Mansarovar Talab from July 2014 to June 2015.	54
04.	Graph showing Monthly Variations in Transparency (cm.) in Mansarovar Talab from July 2015 to June 2016.	54
05.	Graph showing Monthly Variations in Turbidity in Mansarovar Talab from July 2014- June 2015.	58
06.	Graph showing Monthly Variations in Turbidity in Mansarovar Talab from July 2015- June 2016.	58
07.	Graph showing Monthly Variations in pH in Mansarovar Talab from July 2014 to June 2015.	62
08.	Graph showing Monthly Variations in pH in Mansarovar Talab from July 2015 to June 2016.	62

09.	Graph showing Monthly Variations in Dissolved Oxygen (mg/l) in Mansarovar Talab from July 2014 to June 2015.	66
10.	Graph showing Monthly Variations in Dissolved Oxygen (mg/l) in Mansarovar Talab from July 2015 to June 2016.	66
11.	Graph showing Monthly Variations in Total Solids (mg/l) in Mansarovar Talab from July 2014 to June 2015.	70
12.	Graph showing Monthly Variations in Total Solids (mg/l) in Mansarovar Talab from July 2015 to June 2016.	70
13.	Graph showing Monthly Variations in Free CO ₂ (mg/l) in Mansarovar Talab from July 2014 to June 2015.	74
14.	Graph showing Monthly Variations in Free CO ₂ (mg/l) in Mansarovar Talab from July 2015 to June 2016.	74
15.	Graph showing Monthly Variations in Total Alkalinity (mg/l) in Mansarovar Talab from July 2014 to June 2015.	78
16.	Graph showing Monthly Variations in Total Alkalinity (mg/l) in Mansarovar Talab from July 2015 to June 2016.	78
17.	Graph showing Monthly Variations in Biochemical Oxygen Demand (mg/l) in Mansarovar Talab from	82

	July 2014 to June 2015.	
18.	Graph showing Monthly Variations in Biochemical Oxygen Demand (mg/l) in Mansarovar Talab from July 2015 to June 2016.	82
19.	Graph showing Monthly Variations in Total Hardness (mg/l) in Mansarovar Talab from July 2014 to June 2015.	86
20.	Graph showing Monthly Variations in Total Hardness (mg/l) in Mansarovar Talab from July 2015 to June 2016.	86
21.	Graph showing Monthly Variations in Magnesium (mg/l) in Mansarovar Talab from July 2014 to June 2015.	90
22.	Graph showing Monthly Variations in Magnesium (mg/l) in Mansarovar Talab from July 2015 to June 2016.	90
23.	Graph showing Monthly Variations in Calcium (mg/l) in Mansarovar Talab from July 2014 to June 2015.	94
24.	Graph showing Monthly Variations in Calcium (mg/l) in Mansarovar Talab from July 2015 to June 2016.	94
25.	Graph showing Monthly Variations in Chloride (mg/l) in Mansarovar Talab from July 2014 to June 2015.	98
26.	Graph showing Monthly Variations in Chloride	98

	(mg/l) in Mansarovar Talab from July 2015 to June 2016.	
27.	Graph showing Monthly Variations in Nitrate (mg/l) in Mansarovar Talab from July 2014 to June 2015.	102
28.	Graph showing Monthly Variations in Nitrate (mg/l) in Mansarovar Talab from July 2015 to June 2016.	102
29.	Graph showing Monthly Variations in Phosphate (mg/l) in Mansarovar Talab from July 2014 to June 2015.	106
30.	Graph showing Monthly Variations in Phosphate (mg/l) in Mansarovar Talab from July 2015 to June 2016.	106
31.	Graph showing Monthly Variations in Sulphate (mg/l) in Mansarovar Talab from July 2014 to June 2015.	110
32.	Graph showing Monthly Variations in Sulphate (mg/l) in Mansarovar Talab from July 2015 to June 2016.	110
33.	Graph showing Monthly Variations in Total Coli form (MPN/100 ml) in Mansarovar Talab from July 2014 to June 2015.	114
34.	Graph showing Monthly Variations in Total Coli form (MPN/100 ml) in Mansarovar Talab from July 2015 to June 2016.	114
35.	Graph showing Monthly Variations in Faecal Coli Form (MPN/100 ml) in Mansarovar Talab from July	118

	2014- June 2015.	
36.	Graph showing Monthly Variations in Faecal Coli Form (MPN/100 ml) in Mansarovar Talab from July 2015- June 2016.	118
37.	Graph showing Monthly Variations of Diversity Macro-invertebrates reported in Mansarovar Talab from July 2014 to June 2015.	121
38.	Graph showing Monthly Variations of Diversity Macro-invertebrates reported in Mansarovar Talab from July 2015 to June 2016.	122

LIST OF MAP AND PHOTOGRAPHS

S. No.	Name of Photographs	Page. No.
1.	The map of India	14
2.	The map of Madhya Pradesh	15
3.	The map of Dhar	16
4.	Satellite view of the area of Mansarovar Talab of Jeerapura.	17
5.	The area of Mansarovar Talab of Jeerapura.	17
6.	East direction (Station 01)of Mansarovar Talab of Jeerapura.	18
7.	North direction (Station 02)of Mansarovar Talab of Jeerapura.	18
8.	West direction (Station 03) of Mansarovar Talab of Jeerapura.	19
9.	South direction (Station 04) of Mansarovar Talab of Jeerapura.	19
10.	Kick Net, Hand net, D-shaped net and surber sampler	42
11.	Ekman Grab	43-44
12.	Gill Cost Net	45
13.	Photographs showing the species of Annelids in Mansarovar Talab Jeerapura.	126-127
14.	Photographs showing the species of Molluscans in Mansarovar Talab Jeerapura.	128-129
15.	Photographs showing the species of ArthropodsMansarovar Talab Jeerapura.	130-131
16.	Photographs of Fish species of Mansarovar talab of Jeerapura.	141-144

INTRODUCTION

INTRODUCTION

Organic life, which began about three billion years ago in which water is predominant inorganic constituent of living organic matter. In the creation of the atmosphere, a small portion of the hydrosphere region is actually available as resource. Water is the most important requirement for survival. In generally living cell is made up of three quarters by weight of water. It is one of the five basic elements of the body. Water is available in plenty on the surface of the earth but more than 97% occurs in the form of sea water, whose salinity makes it useless, while fresh water makes up only 2.6%. The 4/5 of this is immobilized as ice and thus practically useless. Only less than 1% water is available for drinking. Both urban and rural people communities are highly dependent on the water available for their daily needs. Little amount really available is that in the form of surface and ground running water.

The need for storing the surface runoff water, by constructing dam on rivers has been felt ever since the human civilization. This hydraulic civilization has caused creation of several reservoirs in many water deficit areas of the world. The ponds, lakes and reservoirs are inland depressions containing fresh water and are used for navigation, aquaculture, fish culture & transport, hydropower generation recreation, irrigation and a host of other purposes. It has great social and economic importance because it affects man's health in turn influence all his other activities in many parts of the world. Women and children still spend as much as 50% of their time hauling water for household and stock water purposes. Over and above man also uses these water bodies as most convenient and the cheapest refuse-disposal system for domestic and industrial wastes. This cultural exploitation of our

precious water resources has put a severe stress with the result, the fresh water is becoming rarer day by day as the civilization progresses.

Freshwater bodies may be classified into two types viz, standing water (lentic) and flowing water (lotic). Ponds, reservoirs, lakes and swamps come under lentic water, whereas rivers, springs, perennial monsoon streams are included under the lotic water. The weightage of freshwater resources, their protection and utilization has attained utmost importance during the current time.

The Limnology word was first introduced by F. A. Forel in his work entitled 'Le Léman: monographie limnologique' on Lake Geneva, Switzerland in 1892, 95 and 1904. Forel then published the first text book on Limnology in 1901 'Handbuch der Seenkunde: allgemeine Limnologie'. Forel was a professor at the University of Lausanne and he has been considered as the Father of Modern Limnology.

The biological studies of fresh water commenced with the invention of microscope by *Leeuwenhock* in 1674. Though Leeuwenhock considered himself a microbiologist, his work on aquatic biology describing the filamentous green algae *Spirogyra* from Borkelse Lake was of limnological type. This was the first available account of seasonal cycle in lakes, hinting about the food dynamics and influence of wind on algae ecology. The physical studies of water bodies began in Switzerland when the engineer F. de Duillier measured the waves in lakes in 1730. Further the influence of light, heat, temperature and wind was observed in lake ecosystems. These earlier workers not only established and laid down firm foundation of limnology but also provided guidelines and inspiration to the present-day

workers. In the second half of this century the limnological work took rapid strides and detailed investigations with a stress on productivity in relation to other ecological parameters were carried out by *Goldman and Wetzel, 1963; Lund, 1972; Edmondson, 1977*; etc.

Limnology of fresh water bodies in the temperate regions has been well studied by *Berg, 1964; Jenkin, 1942; Mortimer, 1942; Lindman, 1942; Ganapati, 1941, 1960; George, 1976; Sreenivasan, 1969 and 1974; Michael, 1968; Vasisht, 1968; Munawar, 1970, 74.*

In India the science of limnology was pioneered by *Ganapati in 1941* who made the physico-chemical investigations in the ponds of Madras city. Ganapati continued his studies and published an extensive literature on Indian Reservoir Ecology (*1956, 64, 66 and 81*). *Govind (1969 and 1978)* reported the plankton of Tungabhadra reservoir.

The study of the fresh water organism was attempted by Michael *1968. Okland(1964)* has described the surface and bottom fauna with special reference to gastropods. The relationship between water quality and organisms has attracted some attention recently. *Brinkhurst, 1965, 1970, 1974* has studied the Oligochaets fauna and its relation with organic pollution. *Mortimer (1942)*, for the first time suggested the role of physical feature of water body in determining the quality of water and described Oligochaete fauna in relation to organic pollution. Sreenivasan gave a comparative account of major reservoirs of Madras state. *Unni(1972 & 1985)* also reported a comparative limnological account of several reservoirs of central India. In *1982 & 1992, Jhingran* reviewed the available broad morphometric features of Indian reservoirs. *Rao(1987)* discussed the

morphometry of Rangasagar in relation to its productivity and community structure. *Choubey in(1987)* recorded and discussed the morphometric details of the Gandhisagar reservoir. *Bhatnagar(1984)* incorporated morphometric features and sewage pollution & eutrophication in his studies on the lower lake of Bhopal. *Kulshrestha (1988)* reviewed the previous limnological work on lakes of Bhopal and also investigated the effects of M.I.C. gas leakage on aquatic population of lower lake of Bhopal.

The physico-chemical characteristics of the water body are essentially considered to assess and to monitor the water quality for various purposes. *Bhowmik (1968)* reported the environmental factors affecting the fish food in freshwater fisheries. *Golterman (1967)* reported the influence of soil and chemistry of water in relation to productivity.

The contribution gave in planktonic studies by *Welch (1948& 1952)*, *Ruttner (1963)*, *Wetzel(1975)* etc. In India *Sreenivasan(1974)* worked on the planktons of Bhavanisagar reservoir.

Macro benthic invertebrates are a ubiquitous and diverse group of long-lived species that react strongly and often predictably to human influences in aquatic ecosystem. In addition, they are sedentary, therefore body burdens reflect local conditions, allowing detection of a variety of perturbations in a range of aquatic habitats (*Rosenberg & Resh, 1993*).

Macro benthic invertebrates are an important and integral part of any aquatic ecosystem as they form the basis of the trophic level and any negative effects caused by pollution in the community structure can in turn affect trophic relationships. These can include those that feed on them directly or indirectly such as fish and bird populations, respectively. In addition, aquatic

invertebrates have the ability to clean rivers as they utilize the organic and detritus matter. According to *Carlisle et al., (2007)* macro-invertebrate populations in streams and rivers can assist in the assessment of the overall health of the stream.

Biological assessment and criteria can be used as the basis for management programs, restoring and maintaining the chemical, physical and biological integrity of freshwater. Live organisms offer valuable information regarding their surrounding conditions and can be used to evaluate the physical, chemical and biological impact and their cumulative *effects (Karr & Chu, 1999)*.

Central India is a water deficit area. Now it should be everybody's concern to take care and conserve our existing water resources. Very little work is done on the benthic biodiversity of shore line of tropical shallow lake of the West Malwa region in M.P. Mansarovar Talab of Jeerapur reservoir is a medium sized reservoir and was originally constructed for drinking water supply along with fish culture. This important water body has been neglected so far and no limnological work has been attempted to understand its characteristics. This is the first attempted to carried out the **“Limnological studies on mansarovar talab of Jeerapura, Dhar district (M.P.).”**

Selection of Mansarovar Talab for the present investigation is evident in the light of its importance and in the event of present fresh water crisis faced by the area.

The present research work aims to investigate physico-chemical characteristics, plankton density & diversity, productivity, seasonal

variations and correlation between biotic & abiotic variables of Mansarovar Talab reservoir. The investigated data would further be utilized for comparative limnological studies and for maintenance & conservation of this precious water resource.

Now we realize that dams are very important for the human welfare but in other hand due to the dam formation, aquatic biodiversity are reduced day by day and some species are endangered in condition. The noteworthy contribution of this study would be developing a technique to conserve the macro-invertebrates and Ichthyofauna diversity of MANSAROVAR TALAB OF JEERAPURA as well as to provide new modern technique of fishing to fishermen. This information will be help in the development of conservation & management plan for the planners and this data will be utilized by different government departments, scientists, environmentalists and environmental managers.

REVIEW
OF
LITERATURE

REVIEW OF LITERATURE

Limnological studies were carried out by many workers like *Brinkhurst (1970), Naik & Purohit (2001), Khanna & Bhatia (2003)*.

In India, few works have been done on the study of benthic fauna of fresh water aquatic bodies and published record are available. Some of them are *Patil S.G. et al. (1984), Singhal (1991), Barbhuyan & Khan (1992), Sinha et al. (1997), Chatterjee (1994), Kumar & Singh (1997), Shah & Pandit (2001), Sukla & Shrivastva (2004), Negi R. K. et al. (2008)*.

Benthic macroinvertebrates fauna are those organisms that live on or inside the deposit at the bottom of a water body (*Idowu & Ugwumba, 2005*). Water quality are those physical, chemical and biological factors that influence species composition, diversity, stability, production and physiological conditions of indigenous population of a water body (*Boyd, 1982*).

Benthic macroinvertebrates have also been identified and the highest species number was recorded near tributaries due to the availability of food while the lowest are in the impacted areas where there are pollution discharges and gravel excavation (*Beqiraj et al., 2006*). The occurrence and distribution of macro-invertebrates are governed mostly by the physical & chemical quality of water and immediate substrate of occupation. Temperature, Dissolved oxygen, pH and nutrients have considerable effects on the life of aquatic organisms. Macro invertebrates play an important role in aquatic community which includes mineralization, mixing of sediments and flux of oxygen into sediment, cycling of organic matter and also in assessing the quality of inland water (*George et al. 2009*).

Water is one of the most important natural resource around the world (*EPA 2001, Voelz et al., 2005*). It is a necessity for life and provides a variety of use from drinking water in cities to the irrigation of crops in agricultural areas. Water also provides recreational use as well as habitat for wildlife. River and streams are very important natural, environment and linked to human lives, animal and vegetation (*Haase& Blodgett 2009*).

According to *Wetzel (1975)*, limnology is the study of functional relationship and productivity of fresh water biotic environmental parameters.

According to *Golterman (1978)*, limnology is interdisciplinary science combining various aspects of hydrobiology, hydrochemistry, hydrophysics and geology.

The distribution of macro-invertebrates fauna is determined by a number of factors such as the physical nature of the substratum, depth and nutritive content, degree of stability and oxygen content of the water body. Macro-invertebrate organisms are threatened by changes in their habitat which are associated with pollution, erosion and siltation (*Lydeard et al., 2004*).

Most of these macro-invertebrates share their biological life in fresh water while their adults fly over for miles in search of suitable habitat for reproduction and laying eggs (*Mustow 1996; Akolkar et al., 1999; Muhammed Zaheer Khan 2006; Khanna & Vats 2006; Prasum et al., 2006; Sharma et al., 2013*).

Macro-invertebrate diversity for bioassessment provides a simpler approach and this is due to the fact that they can be sampled quantitatively as well as the known relatively sensitive or tolerance of some of them to

contamination (*Adakole&Annune, 2003*). Species vary in their degree of tolerance with the result that under polluted conditions, a reduction in species diversity is the most obvious effect (*Rosenberg &Resh, 1993; Edokpayi et al., 2000; Emere, 2000; Olomukoro&Egborge; 2003*). The macro-invertebrates are popular as pollution indicators (*Hellawell, 1986*). Benthic macro-invertebrates are best indicator for bio assessment (*Kumar, 2003*).

Studies on water quality management using macro-invertebrates in evaluating the impacts of specific pollutants in aquatic environments have been reported by *Ogbeibu, (2001); Ogbogu& Olajide, (2002); Hart &Zabbey, (2005); Arimoro&Ikomi, (2007); Strayer, (2008); George et al., (2009); Esenowo&Ugwumba, (2010); Sharma et al., (2010); OgidiakaEfe(2012)* and Bio monitoring of Kukatpally IDL lake using Benthic Macro-invertebrates done by *Sultana & Kala (2012)*.

Fish production in lake, talab & reservoir is directly or indirectly dependent on the abundance of plankton and bottom fauna (*Das and Chand, 2003*).The physico-chemical properties of water determine the quality and quantity of the fauna (*Srivastava, 1980*). According to *S. K. Pathak & L.K. Mudgal (2005)* the most dominant family was cyprinidae followed by Bargridae and Mastacembelidae. This is because these three groups are dominant in lentic water reservoirs of India and Bangladesh and are more tolerant towards pollution.

MATERIAL
&
METHODS

MATERIAL AND MEHTOD

Study Area- Dhar is the city located in the Malwa region of western Madhya Pradesh state in Central India. It is situated between latitude 22.598° N and longitude is 75.304° E. Mansarovar Talab of jeerapura is situated in the Dhar, Mandav road, 23k.m. away from Dhar in Madhya Pradesh. The talab is made by parmars. This is very old talab. The talab used to agriculture, fish culture, drinking etc.

The jeerapura is a very old and a gateway of the Mandav of Dhar (M.P.). The jeerapura town is closely located to the Mandav. This is very rich in water resources having large ponds, rivers, irrigational channels. Along these, the mansarovar talab deserves a leading position as it plays a significant role in daily routine work of local inhabitants. It is large, deep, perennial water body and irregularly rectangular in shape. The mansarovar talab was well infested with aquatic weeds.



Photograph showing the map of India



Photograph showing the map of Madhya Pradesh



Photograph showing Satellite view of the area of Mansarovar Talab of Jeerapura



**Photograph showing the area of Mansarovar Talab of Jeerapur
Sampling stations**

This study was carried out at four selected sampling stations identified in the East (Station 01), North (Station 02), West (Station 03) and South (Station 04).



Photograph showing the East direction (Station 01) of Mansarovar Talab of Jeerapura



Photograph showing the North direction (Station 02) of Mansarovar Talab of Jeerapura



Photograph showing the West direction (Station 03) of Mansarovar Talab of Jeerapura



Photograph showing the South direction (Station 04) of Mansarovar Talab of Jeerapura

METHODOLOGY

The research work of present study was during July 2014 to June 2016 for the period of 02 years. For the assessment of water quality following parameters such as Transparency, Water Temperature, Turbidity, Hydrogen Ion Concentration (pH), Dissolved Oxygen (D.O.), Total Solid, Free CO₂, Total Alkalinity, Biological Oxygen Demand (BOD), Hardness, Magnesium, Calcium, Chloride, Nitrate, Phosphate, Sulphate, Total Coliform & Faecal Coliform were estimated.

Following physico-chemical parameters were estimated as per methods given in *APHA (2002)*, *Welch (1998)*, *Trivedi and Goel (1986)* & *Golterman (1978)*.

1. Water Temperature:

Water temperature as a function of depth is often required. Identification of source of water supply such as deep wells often is possible by temperature measurement alone.

Procedure: - Normally, temperature may be made with any good mercury filled Celsius-thermometer. As a minimum, a thermometer should have a scale marked for every 0.1° C, with marking etched on the capillary glass and a minimal thermal capacity to permit rapid equilibrium.

Depth temperature is required for limnological studies were measured with a reversing thermometer. The temperature of surface and sub surface waters was recorded by drawing water sample with the help of a sample or by dipping the thermo probe to the desired depth. Since only the physico-chemical characterization of water bodies was intended not the detailed

study of thermal regime. Simple mercury thermometer was used to note the water temperature usually at the time of sampling that was morning.

2. Transparency:

Transparency was determined by Metal secchi disc of 20 cm in diameter was prepared with two white and two black equal quadrants alternatively on its upper surface. To eliminate the possibility of reflection of light from the other side it was painted black on the middle of the upper surface. A hook was soldered to tie a long wide plastic string and an opposite surface a heavy iron rod was fixed. This extra weight helped in the immersion of disc in the water. The disc was dipped into water with the help of tagged thread and the point of its disappearance was noted. It was then gradually lifted till also disappeared. The point of its reappearance was recorded. The turbidity was calculated by these two readings.

$$\text{Turbidity (cm.)} = \frac{d_1 + d_2}{2}$$

Where,

d_1 = depth when Secchi disc disappeared

d_2 = depth when Secchi disc reappeared

3. Turbidity:

Turbidity is a characteristic property of water. it measures the depth to which light penetrates into the water body. The turbidity of the water sample was measured by Nephelometer turbidity meter. The method is based on the comparison of the intensity of the light scattered by a sample and a standard reference under same condition. For this 5 ml. of hydrazine sulphate solution 1% mixed with 5 ml of hexa methylene tetramine solution (10%) and diluted

to 1000 ml. 10 ml of this solution is diluted to 400 ml forming turbidity standard. Result is expressed in NTU.

4. Hydrogen ion concentration (pH):

pH was determined by pH meter.

Principle: -The pH of the solution refers to its Hydrogen ion activity and is expressed as logarithm of reciprocal of hydrogen ion concentration in moles per liter at given temperature. pH is “intensity” factor of acidity. pH scale ranges from 0-14 with midpoint 7 as a neutral point below and above acidic or alkaline respectively.

Nearest equation governing the effect of concentration of ions and formation of single electrode potential across the glass membrane is the basic concept of pH measurement. pH is an important factor of water chemistry since it enters into the calculation of acidity & alkalinity and processes such as coagulation, disinfection, softening and corrosion control.

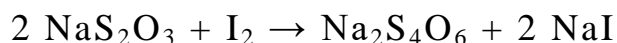
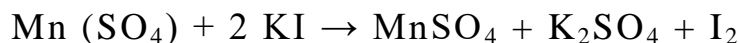
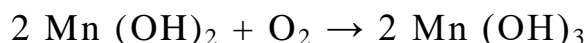
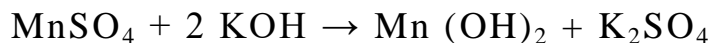
Procedure: - pH measured by systronix battery operated pH meter. The pH meter was earlier calibrated against standard buffer solutions of 7.0 pH and 9.2 pH buffers.

5. Dissolved Oxygen:

The paramount importance to all living organism and is considered to be the lone Factor which to greater extent reveal the nature of whole aquatic system.

Principal: - The magnoussulphate reacts with the alkaline potassium hydroxide to form a white precipitate of magnous hydroxide which in the

presence of oxygen gets oxidized to brown colour compound. In the strong acid medium manganic ions are reduced by iodine which gets converted to iodine equivalent to the original concentration of oxygen in the sample. The liberated iodine can be titrated against sodium thiosulphate using starch as an indicator.



Procedure: - The sample is collected in 300 ml BOD bottle. 2 ml manganous sulphate (36%) and 2 ml alkaline potassium iodine solution (100 g KOH and 50 g KI in 200ml of distilled water) is added to the sample and shake.

The precipitate is allowed to settle. Then 2 ml of concentrate H_2SO_4 is added shaken well till the precipitate dissolved. Titrate the liberated I_2 with 0.25 $\text{Na}_2\text{S}_2\text{O}_3$ (Sodium Thiosulphate) using starch as an indicator.

Calculation: -

$$\text{Dissolved Oxygen in mg/l} = \frac{V1 \times N \times 8 \times 1000}{V2}$$

Where

V1= Volume of $\text{Na}_2\text{S}_2\text{O}_3$ in ml.

N= Normality of $\text{Na}_2\text{S}_2\text{O}_3$.

V2= Volume of sample used.

6. Total Solids (TS):

Procedure: - Take 100 ml beaker and empty weight is taken using weighing balance. Take 100 ml of sample in the place it in the oven at 103⁰c for 24 hrs. (01 day) the water will evaporate and solids will remain as residue at the bottom of the beaker. Put the beaker in the dedicator and allow it to cool for some time. Now the weight of the beaker. The total solids are given the following formula.

$$\text{Total solids} = \frac{\text{Weight of beaker with residue} - \text{Weight of empty beaker}}{\text{ml of sample taken}}$$

7. Free CO₂:

Carbon dioxide concentration influence the acidity of water and thus causes corrosion in the distribution system. The influence of carbon dioxide on water's pH also means that CO₂ concentration can influence the amount of lime which must be added to soften water. The test for CO₂ concentration is very similar to that used to calculate phenolphthalein alkalinity. However, the titrant used in CO₂ testing is N/44 NaOH reacts with CO₂ to form sodium bicarbonate.

Procedure: - free CO₂ was determined by titrating 50 ml of water sample with N/44 NaOH solution using phenolphthalein as indicator.

$$\text{Free CO}_2 \text{ (mg/l)} = \frac{V_1 \times 1000 \times N \times 0.02}{V_2}$$

Where,

V1= Volume of NaOH.

V2= Volume of water sample.

N= Normality of NaOH.

8. Total Alkalinity:

Alkalinity is a measure of the basic constituents of water and is defined as the capacity of a solution to neutralize a standard acid, in natural water. It is usually present as the carbonate & bicarbonatesalts of Calcium, Magnesium, Sodium and Potassium.

Alkalinity is determined by titration with a standard solution of a strong acid to certain end points given by indicator solutions. Phenolphthalein is satisfactory for the first point (pH up to 8.3) contributed by hydroxide & carbonate and methyl orange is used for the second point (pH up to. 4.5) contributed by the bicarbonates.

The phenolphthalein end point of the titration is defined as 'P' alkalinity and the end point observed by continuing the titration with same solution using methyl orange indicator is known as total or T- alkalinity.

Procedure: -

Take 50 or 100 ml of sample in an Erlenmeyer flask, add 2 drops of Phenolphthalein indicator and titrate over a white surface with 0.02N H₂SO₄, until the color change pink coloration just disappears.

Calculation: -

Total Alkalinity = ml. of 0.02N H₂SO₄ X 1000 / ml of sample.

9. Biological Oxygen Demand (BOD):

Biological oxygen demand is the measure of the degraded organic material present in a water sample and can be defined as the amount of Oxygen required by the microorganism in stabilizing the biologically degradable organic matter under aerobic conditions.

Principle: - The principle of the method involves measuring the difference of the oxygen concentration between the sample before and after incubating it for 3 days at 27°C.

Procedure: - Two BOD bottles are taken and filled fully with sample up to the neck. One of the bottles is placed in incubator for 3 days at 27° C and in the second BOD bottle, initial BOD is determined by fixing it with 1 ml of Alkali azide and 1 ml of Magnus sulphate. Then 2 ml. of conc. H₂SO₄ is added so that the precipitate gets settle down. Now 200 ml. of this sample is taken and titrate with Sodium Thiosulphate by adding starch as an indicator, till the sample becomes colourless. BOD bottle is taken out after 3 days from the incubator and the final BOD is determined using the same procedure.

Calculations: -

$$\text{BOD in mg/l} = (D_0 - D_3)$$

Where: D₀ = Initial D₀ in the sample; D₃ = Final D₃ after 3 days

at 27⁰ C.

10. Total Hardness:

Total hardness in water is the sum of the concentration of alkaline earth metal action (Mg^{++} , Ca^{++}) etc.

Principal: -Erichrome black 'T' forms wine red complex compound with metal ion, the di- sodium salt of EDTA (ethylene diamine tetra acetic acid) extracts. The metal ions form the dye metal ion complex as colourless chelate complexes leaving a blue coloured aqueous solution of the dye.

Method: -50 ml. of sample is taken and 2 ml. of ammonia buffer solution and a pinch of Erichrome Black 'T' is added as an indicator. Titrate with EDTA solution until blue colour appears.

Calculation: -

$$\text{Total Hardness in (Mg/l)} = \frac{\text{ml of titrant} \times 1000}{\text{Volume of Sample}}$$

(As $CaCO_3$)

11. Magnesium:

Magnesium in natural water comes mainly from the leaching of igneous and carbonates rocks. In areas where these sources are common, Magnesium concentrations in water often range from 5 to 50mg per litre. Magnesium is

related to water hardness in the same manner as calcium and it also an essential nutrient for plant growth and development.

Reagents:

EDTA solution, 0.01N

Dissolve 3.723gm of EDTA in distilled water to prepare 1000ml solution.

Buffer solution:

(1) Dissolve 16.9gm NH_4Cl in 143ml. of concentrated NH_4OH .

(2) Dissolve 1.179gm of EDTA and 0.78gm $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ in 50ml distilled water.

Mix both (1) and (2) solutions and dilute to 250ml with distilled water.

Eriochrome Black T:

Mix 0.40gm Eriochrome Black T, with 100gm NaCl and grind them.

Procedure: -

(1) Find out the volume of EDTA used in calcium determination.

(2) Also find out the volume of EDTA used in hardness (Mg^{++} , Ca^{++}).

Calculation: -

$$\text{mg/l} = \frac{y - x400.8}{\text{Volume of sample} \times 1.645}$$

Where,

y = EDTA used in hardness determination

x = EDTA used in calcium determination for the same volume of the sample.

12. Calcium:

It is essential for all organisms, being an important cell wall constituent and regulates several physiological processes. It has direct effect on pH and carbonate system.

Principle: - Ammonium purpurate indicator forms pink coloured complexes with Ca^{++} ion with the addition of disodium salts EDTA, the Ca^{++} forms a colourless chelate complex leaving behind a purple solution of the dye.

Procedure: - 50 ml of sample was taken and 1 ml of 0.1 N Sodium Hydroxide Solution and a pinch of ammonium purpurate was added and titrated with EDTA solution untill purple colour appeared.

Calculation: -

$$\text{Ca hardness in mg/l} = \frac{\text{ml of titrant} \times 1000}{\text{Volume of sample}}$$

13. Chloride:

Chloride is usually present in low concentration in natural waters and play metabolically active role in photolysis of water. Free chloride which is commonly used as a disinfectant for drinking and waste soon gets either

converted into chloride or combines with organic matter to form toxic compounds.

Principal: - In potable water the salty taste is produced by chloride ion concentration. The chloride ions are determined by the titration with standard silver nitrate solution in which silver chloride precipitates out. The end point of the titration is indicated by the formation of red silver chromate from excess silver nitrate. The potassium chromate is used as an indicator in neutral to slightly alkaline solution.

Method: - 50 ml sample is taken and 1 to 2 drops of potassium chromate solution is added as an indicator and titrated with silver nitrate solution until colour appears.

Calculations: -

$$\text{Chloride in Mg/l} = \frac{\text{Reading of titrant} \times 500}{\text{Volume of Sample}}$$

14.Nitrate:

Nitrate is the highest oxidation form of nitrogen and in water its most important source is biological oxidation of nitrogenous matter of autochthonous and allochthonous origin. Domestic sewage and agriculture runoff are the chief sources of allochthonous nitrogenous organic matter. Metabolic wastes of aquatic community and dead organisms add to the autochthonous nitrogenous organic matter. Nitrifying bacteria (ammonifying bacteria, Nitrosomonas, Nitrobacter) play significant role in oxidation of such organic matter. Certain nitrogen – fixing bacteria (viz., Azobacter) and algae (viz., blue- greens like Anabena, Nostoc) have

capacity to fix molecule nitrogen in nitrates. In ground water nitrates may through leaching from soil and at times by contamination. The high concentration of nitrate in water is indicative of pollution

Procedure: - Take 10 ml of sample in an Erlenmeyer flask and add 2 ml of sodium chloride solution. Shake the contents and place the flask in a cool water bath. Add slowly 10 ml of sulphuric acid and 0.5 ml of brucine – sulphanilic acid solution. Shake well and put the flask in a hot water bath with boiling water for 20 minutes. Cool the content and record the absorbance on spectrophotometer at 410 nm. Use distilled water as blank.

Calculations:

Run the standard nitrate solutions in similar manner and note the absorbance for each. Plot a standard curve between absorbance and concentrations of various standard solutions. Deduce the value of nitrate – nitrogen in sample by comparing the absorbance of sample (S) with the standard curve and express the result in $\text{mg NO}_3^- \text{-N/l}$.

15. Phosphate:

Phosphate occurs in traces in many natural waters, and often in appreciable amounts during periods of low biologic productivity. Waters receiving raw or treated sewage, agriculture drainage and certain industrial waters normally contain significant concentration of phosphate.

Procedure: - 50 ml of sample was taken. 2 ml of ammonium molybdate solution and 1 ml of stannous chloride solution is added to it. Blue colour appeared for some time and then the reading is taken by spectrophotometre at 690 nm and compared against the calibration curve drawn for various known concentration.

Calculation:

$$\text{Phosphate in mg/l} = \frac{\text{Graph reading} \times 1000 \times \text{dil factor}}{\text{Volume of Sample}}$$

16.Sulphate:

Sulphates are generally present in appreciable concentration and impart hardness to water. Mostly they are present in amounts more than adequate for fresh water productivity.

Principle: - Sulphate gets precipitated with Barium ions in acid solution to form BaSO_4 crystals of uniform size which are estimated spectrophotometrically at 420 nm.

Method: - 100 ml of sample taken and 20 ml buffer solution is then added. Solution is thoroughly stirred and while stirring BaCl_2 crystals are added and kept for 10 minutes for developing the colour. Then it is measured 420 nm using spectrophotometer and compared against the calibration curve drawn for various known concentration.

17.Total Coliform:

Principle: -The coli form group of bacteria is the most widely used indicator organism. It meets many requirements of an ideal indicator.

The coli forms are present in the digestive tract of humans, both diseased and healthy and warm-blooded animals. They find out the number of pathogens in contaminated water and therefore can be detected in a limited volume of water sample. Test for coli forms is relatively simple and inexpensive. It can be performed in ordinary water quality laboratories. The organisms can be reliably identified and enumerated.

They do not multiply in natural aquatic environment. Their die-off rate in the environment is slower compared to most of the pathogenic organisms. Further, they respond to the water treatment processes in the same manner as the pathogen. The coli forms are non-pathogenic, aerobic and facultative, rod shaped bacteria. The coli form group or total coli forms, comprise organisms both of faecal as well as non-faecal origin.

Procedure: -Arrange the fermentation tubes in three rows of five test tubes each.

1. Fill 10 ml of the Tryptose Broth in each test tube by pipette.
2. Place inverted tube in each test tube.
3. Place the cotton plug on each test tube.
4. Shift the filled test tubes and pipettes in the autoclave for sterilization.
5. After the sterilization let the test tubes to cool.
6. Now inoculate the sample with the help of the sterilized pipettes in the volume of 10 ml, 1.0 ml and 0.1 ml using the Spirit lamp as a Sterilizer.
7. Incubate the inoculated tubes at $35 \pm 0.5^\circ\text{C}$ after 48 ± 3 hrs swirl each test tube gently and examine it for growth gas and acidic reactions (Shades of yellow color). The growth with the acidity signifies a positive reaction.

Calculation: -Count the number of Positive tubes in each row and calculate the MPN for total coli form in the sample using the following Table or formula:

$$\text{MPN/100 ml} = \frac{\text{No. of positive tubes} \times 100}{\sqrt{\text{ml. sample in negative tubes} \times \text{ml. sample in all tubes}}}$$

18. Faecal Coliform:

Faecal coliform such as *Escherichia coli* possess an enzyme β -Glucuronidase and are capable of cleaving the fluorogenic substrate MUG (4-methylumbelliferyl- β -D-Glucuronide) with corresponding release of fluorogen when growth in EC-MUG medium at 44.5°C within 24±2 hours, if the test tubes are examined in long wavelength UV lamp, there will be bright blue fluorescence.

Method: 5 tubes in each three row of 10 ml EC-MUG medium were prepared. The fermentation tubes showing growth of gas and acidity for total coli form were rotated gently. Through wire loop the growth was transferred in each EC-MUG test tube. All the inoculated test tubes were kept in incubator at 44.5°C for 24 hours. After incubation all the test tubes were examined for the fluorescence using UV light. The presence of bright blue fluorescence was considered as a positive test for faecal coli form. The numbers of total positive tubes in each row were counted and the MPN number was calculated using MaCardy's table.

Biological samples: -

Monthly samplings of these four selected sampling sites were made (July 2014 to June 2016).

By Different method benthic macro-invertebrates were collected. From the target habitat, samples were collected from the deeper profundal zone by using EKman grab and from shallow profundal zone by using Kick net following *Wetzel (2001)*. Quantitative sampling was done by Kick net and Ekman grab.

Kick Net

Macro-invertebrate samples were collected by using Kick net (20*20 cm) into a single sample following the semi quantitative procedure of *Stark et al., (2001)*. Organisms were collected by stirring and disturbing the substance for about 5 minutes to the depth of several inches to dislodge the borrowing macro-invertebrates ahead of the net per square meter (*Hoffsten&Malmqvist 2000*); (*Llmonen&Paasivira 2005*). Samples were obtained from the same location by brushing the organism of the cobbles and rocks, following standard method of *Borrer et al., (1976)* and *APHA (2002)*.

Ekman grab sampler

This sampler is designed to collect an accurate representative sample of the sediment bottom. Ekman grab equipped with drop-weight system so that sampling can be carried out at any depth. The bite of the sampler should be deep enough so all depths are sampled equally. The sampler should be designed to minimize disturbance of the topmost sediment by the pressure wave as it is lowered to the bottom.

As the sampler is lowered two hinged upper lids swing open to let water pass through and close upon retrieval preventing sample washout. When the sampler reaches the bottom, a messenger is sent down the line tripping the overlapping spring-loaded scoops. Each sampler is constructed of 316 stainless steel including the springs, cables and fasteners.

The number of Macro-invertebrates per unit area was calculated as follows: -

$$\text{Macro-invertebrates No. /cm}^2 = \frac{N \times 10^4}{A}$$

Were,

N = No. of organisms per sample

A = Area of the sampler (20*20cm)

The sample were preserved in 4% formalin solution and transported to the laboratory for further investigation. In the laboratory, samples were washed thoroughly with distilled water to remove preservative through a service with 5mm and 1mm grids. Samples were then poured in a white bottomed tray of the appropriate size for good visualization and the Macro-invertebrates fauna were then identified.

Collected samples were examined under a standard microscope with proper resolution and the fauna was identification using cited taxonomic literature. Samples were assigned to a family/species using taxonomic keys; *APHA (2005)*, *Pennak (2004)*, *Welch (1998)*, *William&Feltmate (1992)*, *Tonapi (1980)* and *Needham & Needham (1969)*.

Collection of Fishes: -

The Collection of fishes will be done from selected station every month during study- period (July 2014 to June 2016). The gill cost net will be used for the collection. After collection of fishes they are preserved in 5% formalin and glycerin. The identification of fishes would be the help of *Shrivastav (1982)*, *Jhingran (1982)* and *Jayaram (1976)*.

Correlation Coefficient between Abiotic (physico-chemical parameters) and Biotic (benthic macro-invertebrates) factors: -

To study the relationship between the physico-chemical parameters and benthic macro-invertebrates during study period (July 2014 to June 2016) Karl Pearson's correlation coefficient (r) method was used.

Correlation analysis provides us information about the relationship between the two variables but it does not indicate us about the causes and effect of relationship.

If both the variables are changing in the same direction that means both are increasing or both are decreasing, then there is a positive correlation between the two variables. If the two variables change in opposite direction, then they possess negative correlation.

The Karl Pearson's correlation is calculated by following formula-

$$r_{xy} = \frac{N \sum XY - \sum X \sum Y}{\sqrt{[N \sum X^2 - (\sum X)^2] [N \sum Y^2 - (\sum Y)^2]}}$$

Where

r = coefficient of correlation.

N = no. of months.

X and Y = variables.

For interpretation purpose

$r = 1$ is considered to be perfect positive correlation.

$0 < r < 0.39$ is considered to be low positive correlation.

$0.40 < r < 0.69$ is considered to be moderate positive correlation.

$0.70 < r < 0.99$ is considered to be high positive correlation.

$-0.39 < r < -0.1$ is considered to be low negative correlation.

$-0.69 < r < -0.40$ is considered to be moderate negative correlation.

$-0.99 < r < -0.70$ is considered to be high negative correlation.

Benthometric Diversity: -

The calculation of diversity index is generally simple as it only requires information of the species number (richness) and the number of individuals of the species (abundance).

Shannon and Weaver Diversity Index

The numerical relationship between the species population and whole communities often provides better reliable indications of pollution than single species (*Dutta & Dutta 1995*). These relationships are represented by “Diversity Indices.” Several types of indices are used. In the present study Shannon and Weaver diversity index (H) (1963) was used.

Shannon and Weaver diversity index: It has been calculated as

$$H = \sum_{i=1}^S (p_i \ln p_i)$$

Where as

H = Shannon and Weaver Index.

$P_i = n_i / N$ (n_i = number of individuals of the species).

N = Total number of individuals in the sample.

The value of Shannon and Weaver Index theoretically range from 0.00 to 4.00. Value less than 1.00 indicates poor water quality, value from 1.00 to 3.00 indicates moderate water quality and value above 3.00 indicates good water quality.

Water Quality Index: -

For calculation of WQI, selection of parameters has great importance. Since selection of too many parameters might widen the water quality index and the importance of various parameters depends on the intended use of water, nine physico-chemical parameters, pH, Temperature, BOD, DO, Phosphate, Faecal Coliform, TDS, Nitrate, and Turbidity were used to calculate the WQI. The calculation of WQI was made using a weighted arithmetic index method given below (*Brown et al., 1970*) in the following steps:

Calculation of sub index of quality rating (q_n)

Let there be n water quality parameters where the quality rating or sub index (q_n) corresponding to the n^{th} parameter is a number reflecting the relative value of this parameter in the polluted water with respect to its standard permissible value. The value of q_n is calculated using the following expression-

$$q_n = 100[(V_n - V_{io}) / (S_n - V_{io})]$$

Where,

q_n = quality rating for the n^{th} water quality parameter.

V_n = estimated value of the n^{th} parameter at a given sampling station.

S_n = standard permissible value of n^{th} parameter

V_{io} = ideal value of n^{th} parameter in pure water.

All the ideal values (V_{io}) are taken as zero for drinking water except for pH=7.0 and dissolved oxygen = 14.6mg/l.

Calculation of quality rating for pH

For pH the ideal value is 7.0 (for natural water) and a permissible value is 8.5 (for polluted water). Therefore, the quality rating for pH is calculated from the following relation:

$$qpH = 100 [(VpH - 7.0) / (8.5 - 7.0)] \quad (2)$$

Where

VpH= observed value of pH during the study period.

Calculation of quality rating for dissolved oxygen-

The ideal value (VDO) for dissolved oxygen is 14.6 mg/l and standard permitted value for drinking water is 5 mg/L. Therefore, quality rating is calculated from following relation:

$$qDO = 100 [(VDO - 14.6) / (5 - 14.6)] \quad (3)$$

where

VDO= measured value of dissolved oxygen

Calculation of unit weight (W_n): -

Calculation of unit weight (W_n) for various water quality parameters are inversely proportional to the recommended standards for the corresponding parameters.

$$W_n = K / S_n \quad (4)$$

where

W_n = unit weight for n^{th} parameters

S_n = standard value for n^{th} parameters

K = constant for proportionality

Calculation of WQI: -

WQI is calculated from the following equation.

$$WQI = \frac{\sum_{n=1}^n q_n W_n}{\sum_{n=1}^n W_n}$$

NSFWQI (National Sanitation Foundation Water Quality Index): -

NSF international is a not-for-profit, non-governmental organization that provides standards development, product certification, auditing, education and risk management for public health and safety. A commonly used water quality index (WQI) was developed by the National Sanitation Foundation (NSF) in 1970 (Brown and others 1970). The NSF WQI was developed to provide a standardized method for comparing the water quality of various bodies of water. The water quality ranges from 0 to 100. The WQI ranges have been defined as 91 to 100 (Excellent), 71 to 90 (Good), 51 to 70 (Medium), 26 to 50 (Poor), 0 to 25 (Bad).

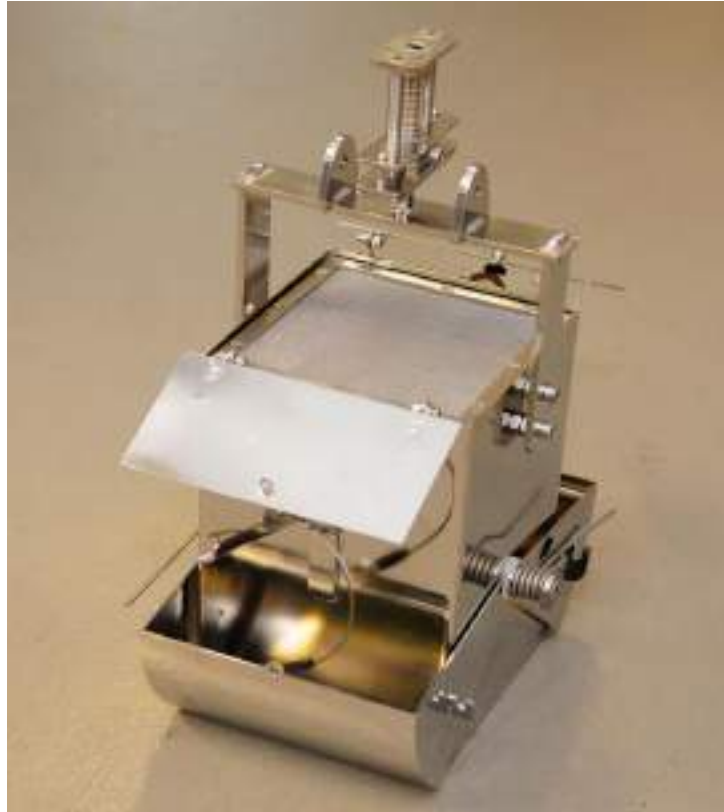
Water quality	NSFWQI	WQ class
Excellent	91 to 100	I
Good	71 to 90	II
Medium or average	51 to 70	III
Poor	26 to 50	IV
Bad	0 to 25	V



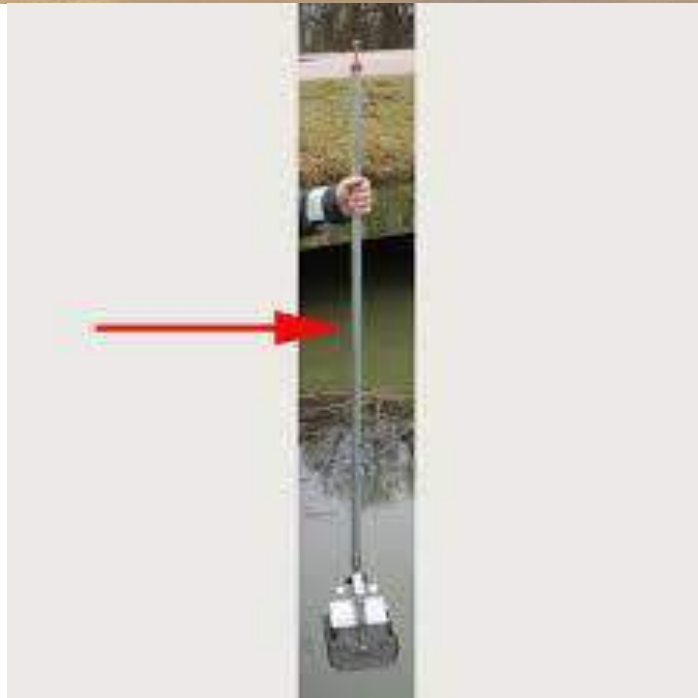
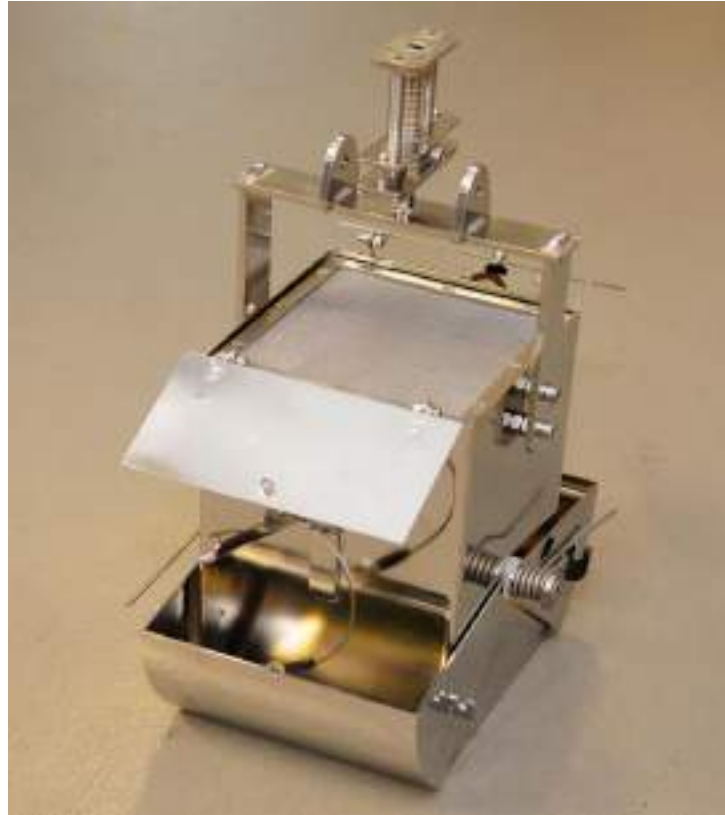
Photograph showing the Kick Net Photograph showing the Hand Net



D-shaped net Surber sampler



Photograph showing the Ekman Grab



Photograph showing the Ekman Grab Sampler



Gill Cost Net



Gill Cost Net

OBSERVATIONS & RESULTS

OBSERVATION AND RESULTS

1. Temperature: -

July 2014 – June 2015

Sampling Site 01. The water temperature varied from 24.9⁰C to 36.2⁰C. The lowest water temperature was observed in December where as highest was in June (Table 1 & fig.1).

Sampling Site 02. At this station water temperature fluctuated between 25.1⁰C to 36.8⁰C. The lowest water temperature was observed in December where as highest was in June (Table 1 & fig.1).

Sampling Site 03. The water temperature oscillated between 25.2⁰C to 36.8⁰C. The lowest water temperature was observed in December where as highest was in June (Table 1 & fig.1).

Sampling Site 04. At this station the water temperature ranged between 24.8⁰C to 37.1⁰C. The lowest water temperature was observed in December where as highest was in June (Table 1 & fig.1).

July 2015 – June 2016

Sampling Site 01. The water temperature varied from 25.5⁰C to 37.0⁰C. The lowest water temperature was recorded in December where as highest was in June (Table 2 & fig.2).

Sampling Site 02. At this station water temperature fluctuated between 26.2⁰C to 37.5⁰C. The lowest water temperature was recorded in December where as highest was in June (Table 2 & fig.2).

SamplingSite 03. The water temperature oscillated between 26.5°C to 37.2°C. The lowest water temperature was recorded in December where the highest was in June (Table 2 & fig.2).

SamplingSite 04. At this station the water temperature ranged between 27.3°C to 37.9°C. The lowest water temperature was recorded in December where the highest was in June (Table 2 & fig.2).

It was fluctuated between 24.8°C to 37.9°C. The lowest water temperature was observed at sampling site 04 in December in the year 2014-2015 and highest value was observed at sampling site 04 in June in the year 2015-2016 (Table 1 & fig.1 and Table 2 & fig.2).

Table- 1. Monthly Variations in Water Temperature (°C) in Mansarovar

Month	Site 01	Site 02	Site 03	Site 04
Jul.	26.5	26.4	25.5	26.9
Aug.	28.8	29.5	27.5	28.8
Sep.	30.5	31.0	30.0	30.5
Oct.	28.8	29.2	28.2	29.0
Nov.	26.5	25.5	26.9	26.5
Dec.	24.9	25.1	25.2	24.8
Jan.	27.0	26.5	27.5	26.9
Feb.	29.5	28.9	30.2	29.5
Mar.	31.2	32.5	30.8	31.9
Apr.	33.9	34.2	33.5	35.0
May	35.8	35.9	35.2	36.2
Jun.	36.2	36.8	36.8	37.1

Talab of Jeerapura (Dhar) Year July 2014- June 2015.

Table- 2. Monthly Variations in Water Temperature (°C) in Mansarovar Talab of Jeerapura (Dhar) Year July 2015- June 2016.

Month	Site 01	Site 02	Site 03	Site 04
Jul.	27.5	28.5	28.2	29.0
Aug.	29.2	29.9	27.9	28.5
Sep.	30.8	31.2	29.5	30.5
Oct.	29.1	29.5	30.0	28.5
Nov.	26.5	27.0	27.5	28.3
Dec.	25.5	26.2	26.5	27.3
Jan.	28.0	27.5	28.8	28.4
Feb.	28.5	28.0	29.5	29.0
Mar.	32.2	32.7	31.9	32.5
Apr.	34.0	34.6	35.0	35.5
May	35.9	36.2	36.0	36.5
Jun.	37.0	37.5	37.2	37.9

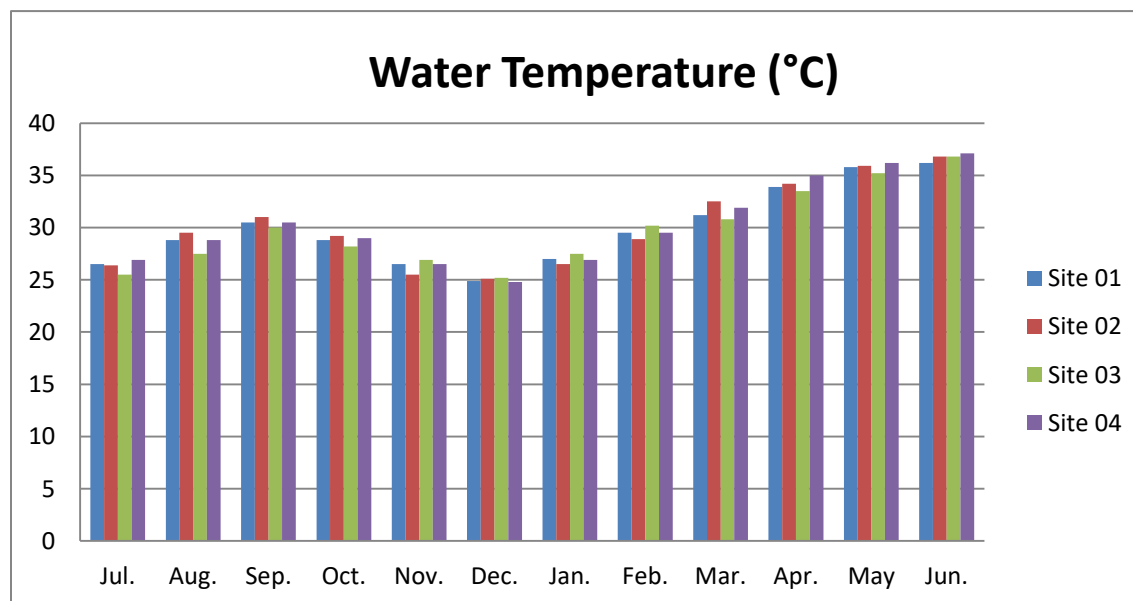


Fig.-1. Monthly Variations in Water Temperature (°C) in Mansarovar Talab of Jeerapura (Dhar) Year July 2014- June 2015.

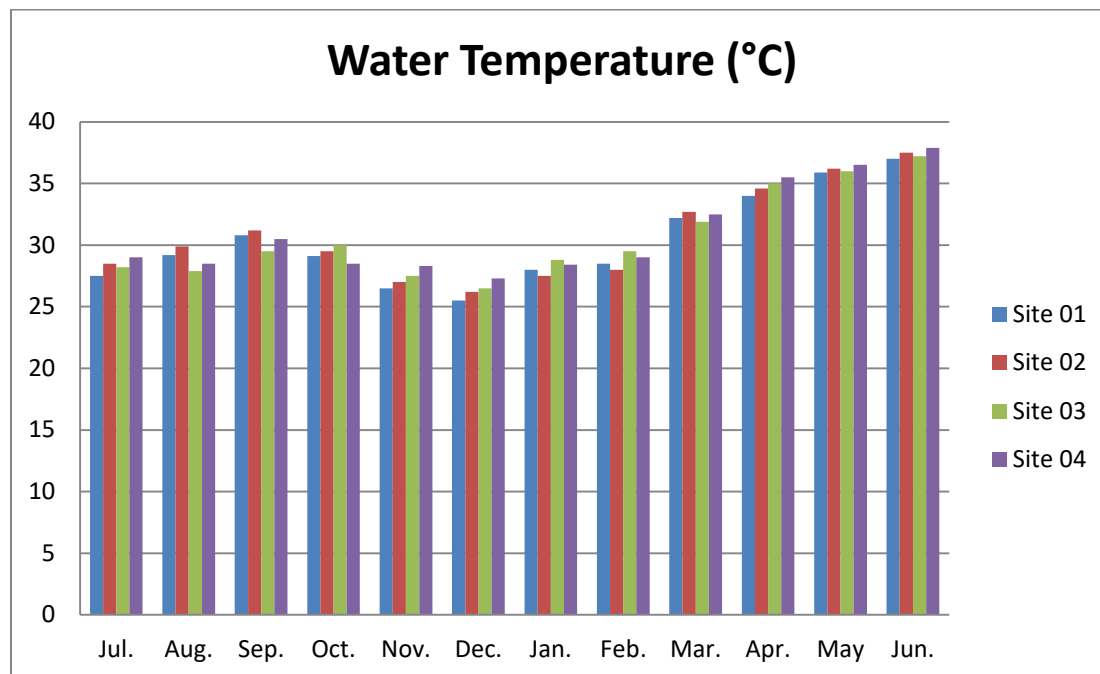


Fig. - 2. Monthly Variations in Water Temperature (°C) in Mansarovar Talab of Jeerapura (Dhar) Year July 2015- June 2016.

2 Light Transparency: -

July 2014 – June 2015

SamplingSite 01.The water light transparency varied from 21.6 cm to 44.2 cm. The lowest water Light Transparency was recorded in May where ashighest was in November (Table 3 & fig.3).

SamplingSite 02.At this station light transparency fluctuated between 20.2 cm. to 49.2 cm. Thelowest waterLight Transparency was recorded in Julywhere ashighest was in October (Table 3& fig.3).

SamplingSite 03.The water light transparency oscillated between 22.0 cm. to 42.2 cm. The lowest water Light Transparency was recorded in July where ashighest was in November (Table 3 & fig.3).

SamplingSite 04.At this station the water light transparency ranged between 21.2 cm. to 44.2 cm. Thelowest Light Transparency was observed in July where ashighest was in October and November (Table 3 & fig.3).

July 2015 – June 2016

SamplingSite 01.The water light transparency varied from 20.0 cm to 48.5 cm. The lowest water light transparency was recorded in Maywhere ashighest was in December (Table 4 & fig.4).

SamplingSite 02.At this station water light transparency fluctuated between 19.8 cm to 47.2 cm. The lowest water light transparency was recorded in Julywhere ashighest was in November (Table 4 & fig.4).

SamplingSite 03.The water light transparencyoscillated between 23.0 cm to 49.4 cm. The lowest water light transparency was recorded in May where ashighest was in December (Table 4 & fig.4).

SamplingSite 04.At this station the water light transparency ranged between 20.5 cm to 50.0 cm. The lowest water light transparency was recorded in July where ashighest was in November (Table 4 & fig.4).

In general transparency fluctuated between 19.8 cm to 50.0 cm. The lowest transparency was observed at sampling site 02 in July in the year of 2015-16. The highest transparency was at samplingsite 04 in November in the year 2015-2016 (Table 4 & fig.4).

Table- 3. Monthly Variations in Light Transparency (cm.) in Mansarovar Talab of Jeerapura (Dhar) Year July 2014 - June2015.

Month	Site 01	Site 02	Site 03	Site 04
Jul.	23.0	20.2	22.0	21.2
Aug.	24.5	21.5	24.9	22.5
Sep.	28.6	24.5	29.2	23.5
Oct.	43.2	49.2	40.2	44.2
Nov.	44.2	46.2	42.2	44.2
Dec.	41.5	40.0	40.5	38.8
Jan.	38.2	37.0	39.1	38.2
Feb.	39.8	37.5	40.2	38.5
Mar.	30.7	31.8	31.6	32.3
Apr.	28.0	31.2	29.0	32.0
May	21.6	38.0	22.5	39.0
Jun.	26.5	35.1	27.4	36.5

Table- 4. Monthly Variations in Light Transparency (cm.) in Mansarovar Talab of Jeerapura (Dhar) Year July 2015- June 2016.

Month	Site 01	Site 02	Site 03	Site 04
Jul.	22.0	19.8	23.2	20.5
Aug.	23.9	20.5	25.0	21.5
Sep.	26.5	24.9	28.5	26.5
Oct.	40.2	44.2	42.5	45.5
Nov.	46.2	47.2	48.0	50.0
Dec.	48.5	42.0	49.4	44.2
Jan.	36.5	35.0	37.5	36.2
Feb.	38.2	35.9	39.5	38.2
Mar.	30.0	29.8	32.0	33.5
Apr.	27.0	30.0	29.0	32.2
May	20.0	36.0	23.0	38.4
Jun.	24.2	32.0	27.5	35.0

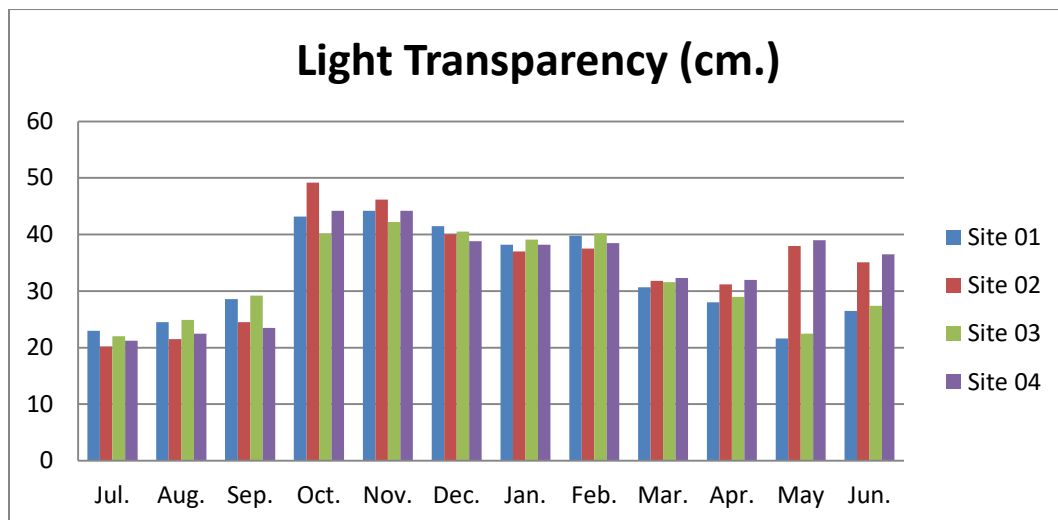


Fig.-3. Monthly Variations in Light Transparency (cm.) in Mansarovar Talab of Jeerapura (Dhar) Year July 2014 - June 2015.

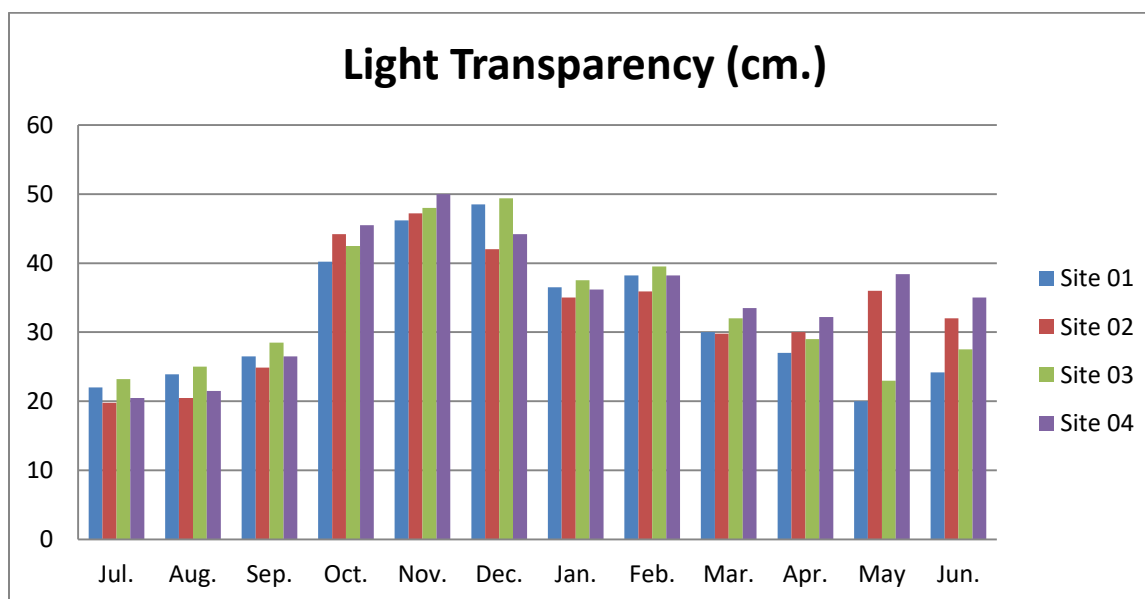


Fig. - 4. Monthly Variations in Light Transparency (cm.) in Mansarovar Talab of Jeerapura (Dhar) Year July 2015- June 2016.

3. Turbidity: -

July 2014 – June 2015

SamplingSite 01.The water turbidity varied from 12 to 55. The lowest water turbidity was recorded in December where ashighest was in October (Table 5 & fig.5).

SamplingSite 02.At this station water turbidity fluctuated between 14 to 62. Thelowest waterturbidity was recorded in Decemberwhere ashighest was in October (Table 5& fig.5).

SamplingSite 03.The water turbidity oscillated between 14 to 50. Thelowest water turbiditywas recorded in December where ashighest was in October (Table 5 & fig.5).

SamplingSite 04.At this station the water turbidity ranged between 11 to 60. Thelowest water turbiditywas recorded in Decemberwhere as highest was in October (Table 5 & fig.5).

July 2015 – June 2016

SamplingSite01.The water turbidity varied from 18 to 55. The lowest water turbidity was recorded in December, March and Aprilwhere ashighest was in October (Table 6 & fig.6).

SamplingSite 02.At this station water turbidity fluctuated from 19 to 65. The lowest water turbidity was recorded in Aprilwhere ashighest was in October (Table 6 & fig.6).

SamplingSite 03.The water turbidity oscillated between 17 to 53. The lowest water turbidity was recorded in April where a highest was in August (Table 6 & fig.6).

SamplingSite 04.At this station the water turbidity ranged between 20 to 68. The lowest water turbidity was recorded in April where a highest was in October (Table 6& fig.6).

In the present study turbidity fluctuated between 11 to 68. The lowest turbidity was observed at site 04 in December in the year 2014-2015 and highest value was observed at site 04 in October in the year 2015-2016 (Table 5 & fig.5 and Table 6 & fig.6).

Table 5. Monthly Variations in Turbidity (NTU) in Mansarovar Talab of Jeerapura (Dhar) Year July 2014- June 2015.

Month	Site 01	Site 02	Site 03	Site 04
Jul.	38	35	36	34
Aug.	30	36	31	33
Sep.	45	48	40	45
Oct.	55	62	50	60
Nov.	30	16	29	18
Dec.	12	14	14	11
Jan.	20	22	23	20
Feb.	28	25	30	28
Mar.	15	18	18	20
Apr.	19	20	17	19
May	30	26	32	29
Jun.	32	34	33	35

Table- 6. Monthly Variations in Turbidity (NTU)in Mansarovar Talab of Jeerapura (Dhar) Year July 2015- June 2016.

Month	Site 01	Site 02	Site 03	Site 04
Jul.	45	40	47	42
Aug.	50	55	53	55
Sep.	46	44	48	49
Oct.	55	65	52	68
Nov.	38	25	42	30
Dec.	18	20	20	23
Jan.	25	28	27	29
Feb.	32	35	34	37
Mar.	18	22	19	23
Apr.	18	19	17	20
May	20	21	21	24
Jun.	35	38	38	39

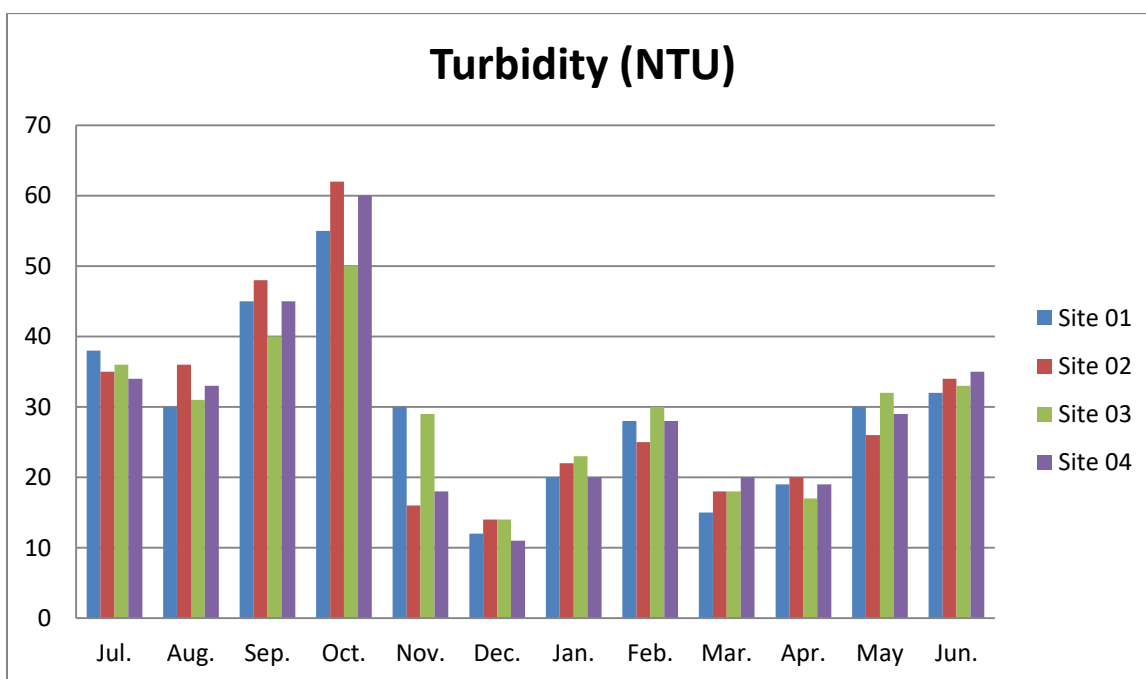


Fig. - 5. Monthly Variations in Turbidity (NTU)in Mansarovar Talab of Jeerapura (Dhar) Year July 2014- June 2015.

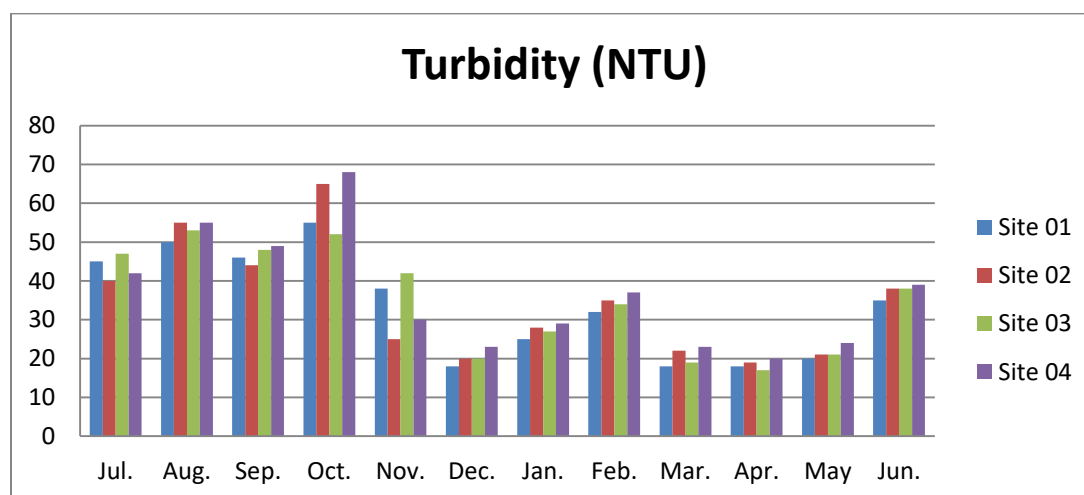


Fig. - 6. Monthly Variations in Turbidity (NTU)in Mansarovar Talab of Jeerapura (Dhar) Year July 2015- June 2016.

4. Hydrogen Ion Concentration (pH): -

July 2014 – June 2015

SamplingSite 01.The water Hydrogen Ion Concentration (pH)varied from 7.4 to 8.5. The lowest water Hydrogen Ion Concentration (pH) was recorded in Januarywhere ashighest was in October (Table 7 & fig.7).

SamplingSite 02.At this station water Hydrogen Ion Concentration (pH) fluctuated between 7.6 to 8.5. The lowest water Hydrogen Ion Concentration (pH) was recorded in Januarywhere ashighest was in June (Table 7& fig.7).

SamplingSite 03.The water Hydrogen Ion Concentration (pH)oscillatedbetween 7.7 to 8.8. The lowest water Hydrogen Ion Concentration (pH) was recorded in August and Januarywhere ashighest was in April (Table 7 & fig.7).

SamplingSite 04.At this station the water Hydrogen Ion Concentration (pH) ranged between 7.8 to 8.9. The lowest water Hydrogen Ion Concentration (pH) was recorded in Januarywhere ashighest was in April and June (Table 7 & fig.7).

July 2015 – June 2016

SamplingSite 01.The water Hydrogen Ion Concentration (pH)varied from 7.6 to 8.9. The lowest water Hydrogen Ion Concentration (pH) was recorded in Januarywhere ashighest was in April (Table 8 & fig.8).

SamplingSite 02.At this station water Hydrogen Ion Concentration (pH) fluctuated between 7.8 to 9.0. The lowest water Hydrogen Ion Concentration (pH) was recorded in July and Januarywhere ashighest was in June (Table 8 & fig.8).

SamplingSite 03.The water Hydrogen Ion Concentration (pH)ocillated between 7.8 to 8.9. The lowest water Hydrogen Ion Concentration (pH) was recorded in Augustwhere ashighest was in June (Table 8 & fig.8).

SamplingSite 04.At this station the water Hydrogen Ion Concentration (pH) ranged between 7.9 to 9.1. The lowest water Hydrogen Ion Concentration (pH) was recorded in Julywhere ashighest was in June (Table 8 & fig.8).

pH fluctuated between 7.4 to 9.1. The lowest pH was observed at samplingsite 01 in January in the year 2014-15. The highest pH was obderved at samplingsite 04 in June in 2015-16 (Table 7 & fig.7 and Table 8 & fig.8).

Table 7. Monthly Variations in Hydrogen Ion Concentration (pH) in Mansarovar Talab of Jeerapura (Dhar) Year July 2014- June 2015.

Month	Site 01	Site 02	Site 03	Site 04
Jul.	7.7	7.9	7.8	8.0
Aug.	7.9	8.1	7.7	8.2
Sep.	8.2	8.4	8.1	8.5
Oct.	8.5	8.2	8.6	8.4
Nov.	7.6	7.8	7.8	7.9
Dec.	7.9	8.1	8.1	8.3
Jan.	7.4	7.6	7.7	7.8
Feb.	7.9	8.2	7.8	8.4
Mar.	8.4	8.3	8.6	8.5
Apr.	8.4	7.9	8.8	8.9
May	8.0	7.8	8.3	8.2
Jun.	8.2	8.5	8.6	8.9

Table 8. Monthly Variations in Hydrogen Ion Concentration (pH) in Mansarovar Talab of Jeerapura (Dhar) Year July 2015- June 2016.

Month	Site 01	Site 02	Site 03	Site 04
Jul.	7.9	7.8	8.0	7.9
Aug.	7.7	8.3	7.8	8.4
Sep.	8.4	8.5	8.6	8.8
Oct.	8.6	8.2	8.9	8.7
Nov.	7.9	8.1	8.5	8.7
Dec.	8.2	8.4	8.5	8.6
Jan.	7.6	7.8	7.9	8.0
Feb.	8.0	8.3	8.2	8.4
Mar.	8.6	8.7	8.8	8.9
Apr.	8.9	8.8	8.7	8.6
May	8.8	8.9	8.4	9.0
Jun.	8.6	9.0	8.9	9.1

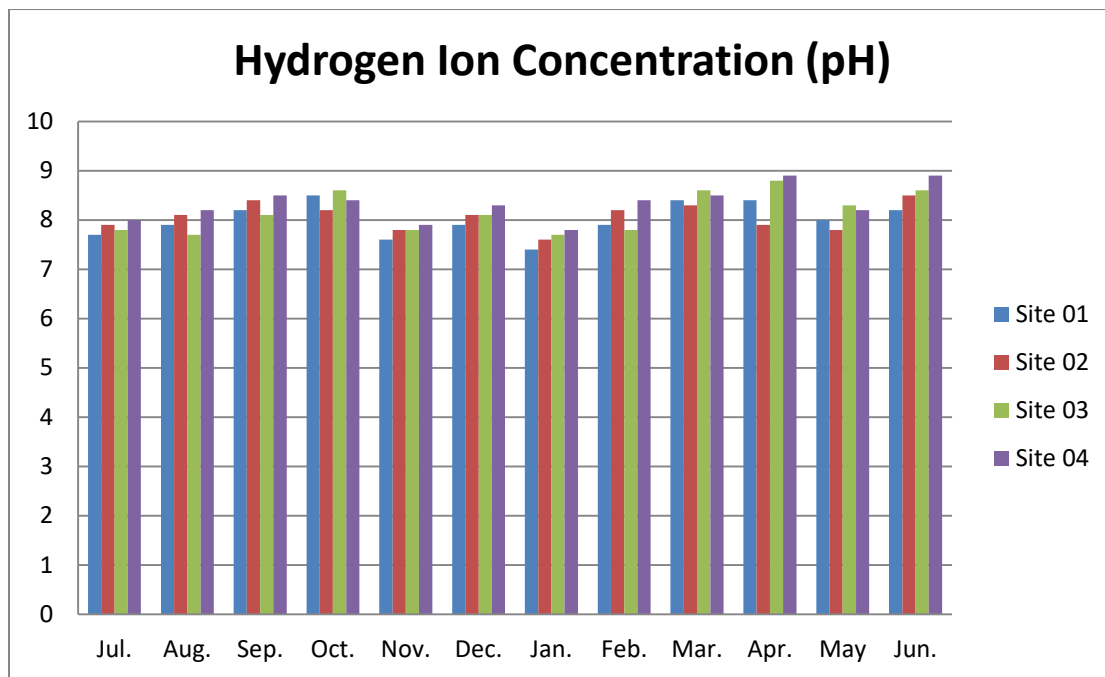


Fig.-7. Monthly Variations in Hydrogen Ion Concentration (pH) in Mansarovar Talab of Jeerapura (Dhar) Year July 2014- June 2015.

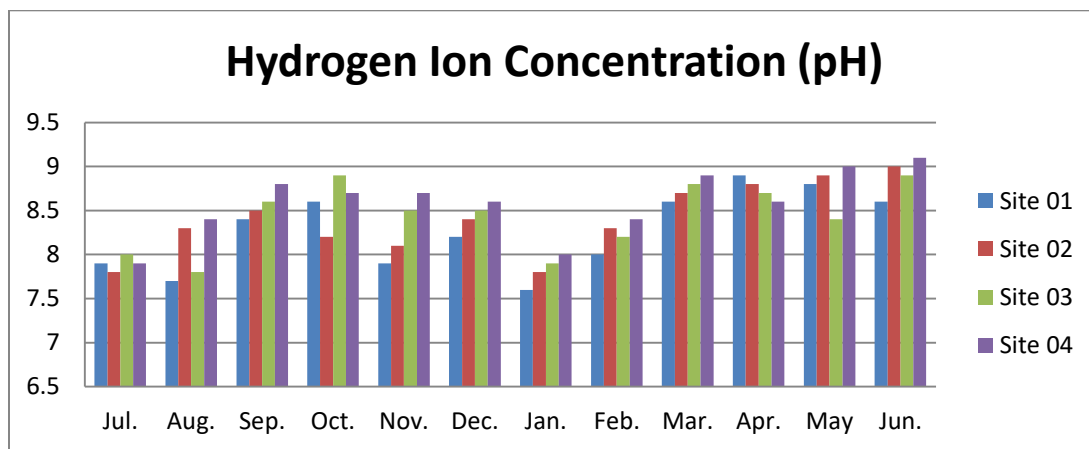


Fig.-8. Monthly Variations in Hydrogen Ion Concentration (pH) in Mansarovar Talab of Jeerapura (Dhar) Year July 2015- June 2016.

5. Dissolved Oxygen (D.O.): -

July 2014 – June 2015

SamplingSite 01.The water Dissolved Oxygen (D.O.) varied from 5.6 mg/l to 12.3 mg/l. The lowest water Dissolved Oxygen (D.O.) was recorded in September where ashighest was in June (Table 9 & fig.9).

SamplingSite 02.At this station water Dissolved Oxygen (D.O.) fluctuated between 6.4 mg/l to 12.0 mg/l. The lowest water Dissolved Oxygen (D.O.) was recorded in Augustwhere ashighest was in November (Table 9 & fig.9).

SamplingSite 03.The water Dissolved Oxygen (D.O.) oscillatedbetween 5.2 mg/l to 10.2 mg/l. The lowest water Dissolved Oxygen (D.O.) was recorded in September where ashighest was in June (Table 9 & fig.9).

SamplingSite 04.At this station the water Dissolved Oxygen (D.O.) ranged between 6.2 mg/l to10.0 mg/l. The lowest water Dissolved Oxygen (D.O.) was recorded in August and September where ashighest was in November (Table 9 & fig.9).

July 2015 – June 2016

SamplingSite 01.The water Dissolved Oxygen (D.O.) varied from 6.2mg/l to 9.8 mg/l. The lowest water Dissolved Oxygen (D.O.) was recorded in Julywhere ashighest was in June (Table 10& fig.10).

SamplingSite 02.At this station water Dissolved Oxygen (D.O.) fluctuated between 6.0 mg/l to 9.8 mg/l. The lowest water Dissolved Oxygen (D.O.)

was recorded in July and September where the highest was in December (Table 10 & fig.10).

Sampling Site 03. The water Dissolved Oxygen (D.O.) oscillated between 5.9 mg/l to 9.0 mg/l. The lowest water Dissolved Oxygen (D.O.) was recorded in August where the highest was in May (Table 10 & fig.10).

Sampling Site 04. At this station the water Dissolved Oxygen (D.O.) ranged between 5.8 mg/l to 8.7 mg/l. The lowest water Dissolved Oxygen (D.O.) was recorded in July and September where the highest was in June (Table 10 & fig.10).

In general, dissolved oxygen varied between 5.2 to 12.3 mg/l. The lowest value was observed at sampling site 03 in September in the year of 2014-15. The highest dissolved oxygen value was recorded at sampling site 01 in June in the year of 2014-15 (Table 10 & fig.10).

Table 9. Monthly Variations in Dissolved Oxygen (mg/l) in Mansarovar Talab of Jeerapura (Dhar) Year July 2014- June 2015.

Month	Site 01	Site 02	Site 03	Site 04
Jul.	6.7	6.9	6.2	6.5
Aug.	6.6	6.4	6.0	6.2
Sep.	5.6	6.5	5.2	6.2
Oct.	8.8	8.5	8.0	7.8
Nov.	10.8	12.0	9.8	10.0
Dec.	9.2	10.0	8.8	9.0
Jan.	9.5	9.2	8.5	8.2
Feb.	7.5	7.9	7.2	7.0
Mar.	8.5	8.9	8.0	8.2
Apr.	9.2	8.8	8.8	8.3
May	10.5	11.5	9.6	9.1
Jun.	12.3	11.2	10.2	9.8

Table 10. Monthly Variations in Dissolved Oxygen (mg/l) in Mansarovar Talab of Jeerapura (Dhar) Year July 2015- June 2016.

Month	Site 01	Site 02	Site 03	Site 04
Jul.	6.2	6.0	6.0	5.8
Aug.	6.9	7.2	5.9	6.9
Sep.	6.6	6.0	7.0	5.8
Oct.	7.5	7.9	6.2	6.8
Nov.	8.8	9.2	7.5	8.0
Dec.	8.2	9.5	7.9	8.2
Jan.	8.8	8.9	8.4	8.2
Feb.	7.9	7.0	7.4	7.6
Mar.	8.0	7.7	7.8	7.5
Apr.	8.5	8.0	8.0	8.3
May	9.5	8.8	9.0	8.3
Jun.	9.8	9.2	8.9	8.7

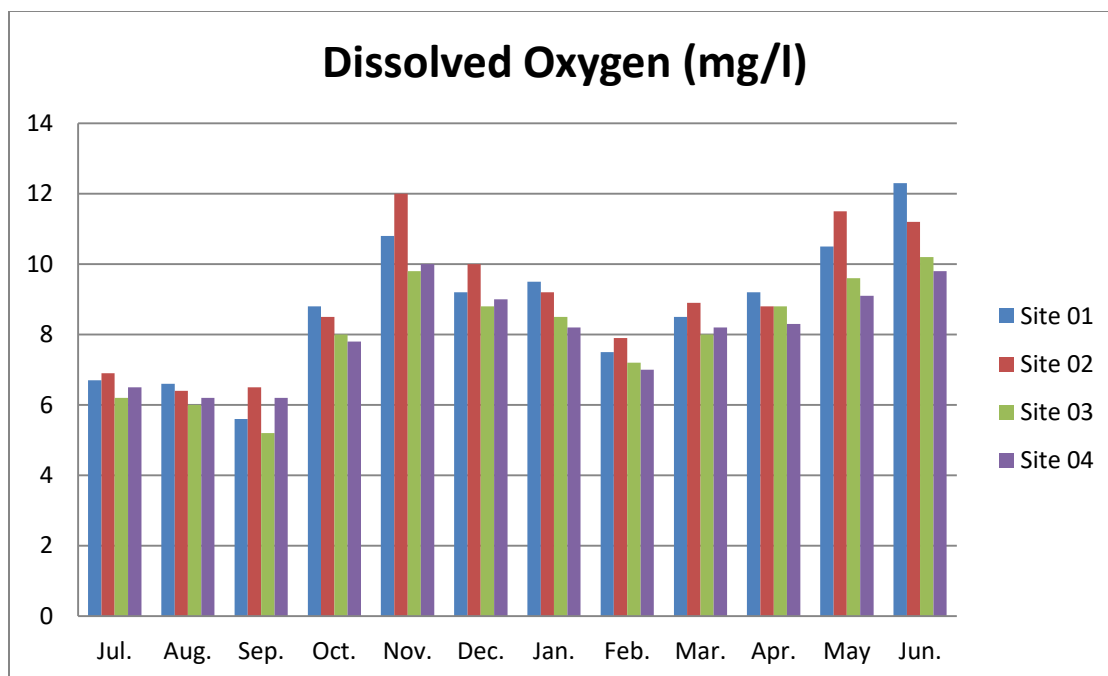


Fig. - 9. Monthly Variations in Dissolved Oxygen (mg/l) in Mansarovar Talab of Jeerapura (Dhar) Year July 2014- June 2015.

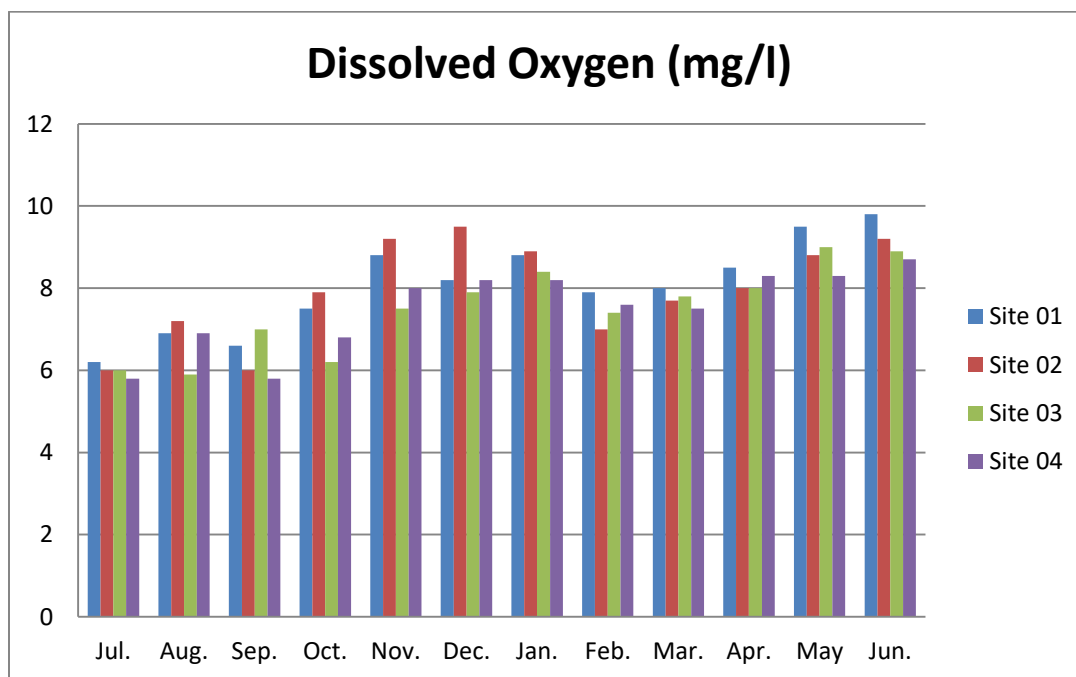


Fig. - 10. Monthly Variations in Dissolved Oxygen (mg/l) in Mansarovar Talab of Jeerapura (Dhar) Year July 2015- June 2016.

6. Total Solid: -

July 2014 – June 2015

SamplingSite 01.The Total Solid varied from 188 mg/l to 380 mg/l. The lowest Total Solid was recorded in November where as highest was in December (Table 11 & fig.11).

SamplingSite 02.At this station Total Solid fluctuated between 170 mg/l to 365 mg/l. The lowest Total Solid was recorded in November where as highest was in December (Table 11 & fig.11).

SamplingSite 03.The Total Solid oscillated between 210 mg/l to 390 mg/l. The lowest Total Solid was recorded in November where as highest was in December (Table 11 & fig.11).

SamplingSite 04.At this station the Total Solid ranged between 235 mg/l to 375 mg/l. The lowest Total Solid was recorded in October where as highest was in December (Table 11 & fig.11).

July 2015 – June 2016

SamplingSite 01.The Total Solid ranged from 190 mg/l to 295 mg/l. The lowest Total Solid was recorded in October and April where as highest was in January (Table 12 & fig.12).

SamplingSite 02.At this station Total Solid fluctuated between 205 mg/l to 285 mg/l. The lowest Total Solid was recorded in October and April where as highest was in July (Table 12 & fig.12).

SamplingSite 03. The Total Solid oscillated between 200 mg/l to 290 mg/l. The lowest Total Solid was recorded in April where as highest was in July and August (Table 12 & fig.12).

SamplingSite 04. At this station the Total Solid ranged between 225mg/l to 305mg/l. The lowest Total Solid was recorded in April where as highest was in July (Table 12 & fig.12).

It was fluctuated between 170 to 390 mg/l. The lowest Total Solid was observed at sampling site 02 in November in the year 2014-2015 and highest value was observed at sampling site 03 in December in the year 2014-2015 (Table 11 & fig.11).

Table 11. Monthly Variations in Total Solid (mg/l) in Mansarovar Talab of Jeerapura (Dhar) Year July 2014- June 2015.

Month	Site 01	Site 02	Site 03	Site 04
Jul.	350	315	370	340
Aug.	300	280	315	260
Sep.	278	225	290	250
Oct.	235	210	225	235
Nov.	188	170	210	240
Dec.	380	365	390	375
Jan.	320	295	333	295
Feb.	255	268	266	280
Mar.	280	270	290	300
Apr.	290	302	280	312
May	325	298	335	358
Jun.	320	335	340	365

Table 12. Monthly Variations in Total Solid (mg /l) in Mansarovar Talab of Jeerapura (Dhar) Year July 2015- June 2016.

Month	Site 01	Site 02	Site 03	Site 04
Jul.	280	285	290	305
Aug.	270	255	290	268
Sep.	215	225	235	255
Oct.	190	205	210	235
Nov.	228	235	252	247
Dec.	245	200	265	230
Jan.	295	278	285	298
Feb.	260	240	277	250
Mar.	205	220	225	240
Apr.	190	205	200	225
May	225	210	245	250
Jun.	255	272	278	288

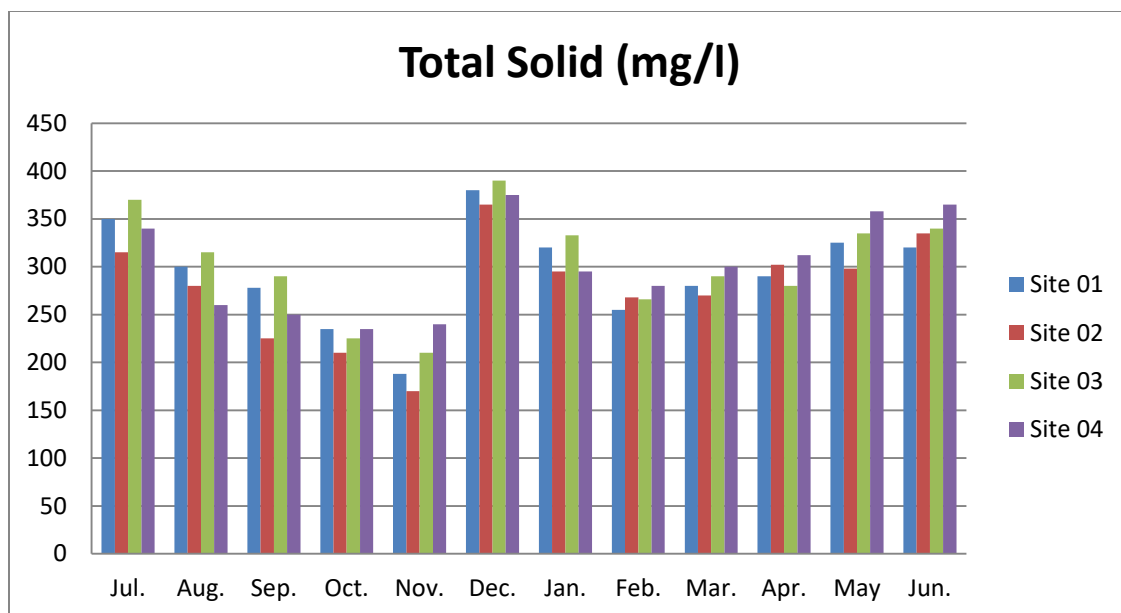


Fig. - 11. Monthly Variations in Total Solid (mg /l) in Mansarovar Talab of Jeerapura (Dhar) Year July 2014- June 2015.

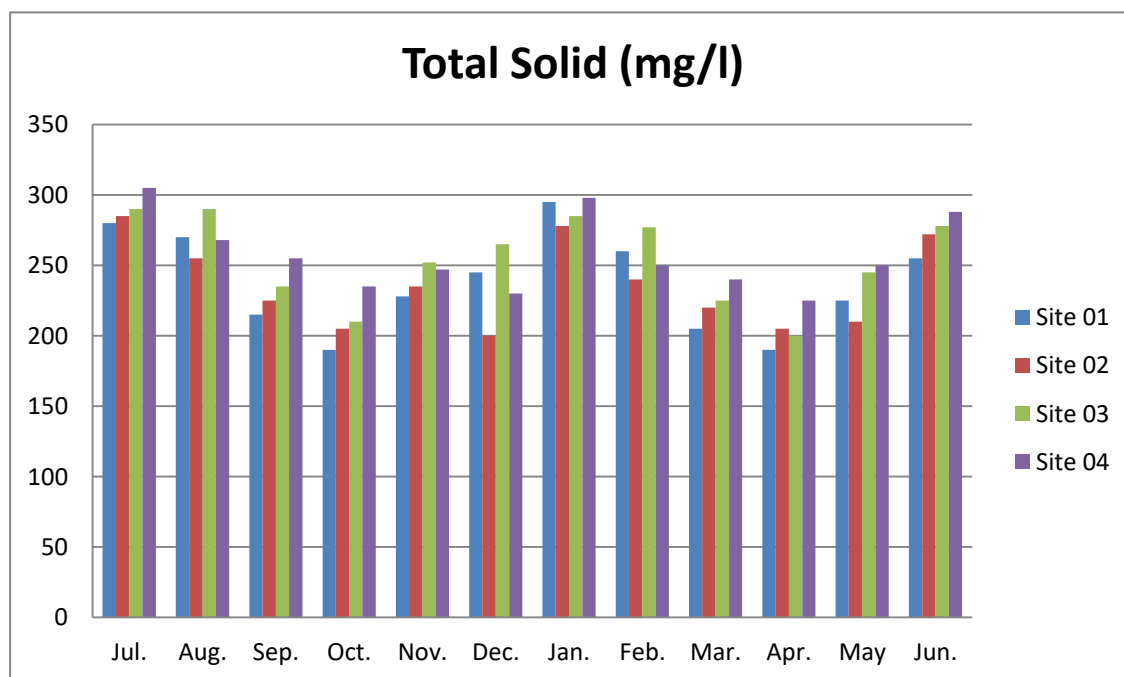


Fig. – 12. Monthly Variationss in Total Solid (mg/l) in Mansarovar Talab of Jeerapura (Dhar) Year July 2015- June 2016.

7.Free CO₂: -

July 2014 – June 2015

SamplingSite 01.The Free CO₂ varied from 1.2mg/l to 6.4mg/l. The lowest Free CO₂ was observed in April where a highest was in October (Table 13& fig.13).

SamplingSite 02.At this station Free CO₂ fluctuated between 1.5mg/l to 6.9mg/l. The lowest Free CO₂ was observed in March where a highest was in November (Table 13& fig.13).

SamplingSite 03.The Free CO₂ oscillated between 1.6mg/l to 6.8mg/l. The lowest Free CO₂ was recorded in April where a highest was in November (Table 13& fig.13).

SamplingSite 04.At this station the Free CO₂ ranged between 1.9 mg/l to 6.5mg/l. The lowest Free CO₂ was recorded in January and March where a highest was in October (Table 13& fig.13).

July 2015 – June 2016

SamplingSite 01.The Free CO₂ varied from 1.9mg/l to 7.0mg/l. The lowest Free CO₂ was observed in April where a highest was in November (Table 14& fig.14).

SamplingSite 02.At this station Free CO₂ fluctuated between 2.2mg/l to 7.5mg/l. The lowest Free CO₂ was observed in January where a highest was in November (Table 14& fig.14).

SamplingSite 03.TheFree CO₂ ocillated between 2.8mg/l to 8.0mg/l. The lowest Free CO₂was recorded in Aprilwhere ashighest was in November (Table 14& fig.14).

SamplingSite 04.At this station the Free CO₂ ranged between 2.6mg/l to 8.5mg/l. The lowest Free CO₂ was recorded in Januarywhere ashighest was in November (Table 14& fig.14).

It was fluctuated between1.2 to 8.5 mg/l. The lowest Free CO₂was observed at samplingsite 01 in April in the year 2014-2015 and highest value was observed at samplingsite 04 in November in the year 2015-2016 (Table 13& fig.13 and Table 14 & fig.14).

Table 13. Monthly Variations in Free CO₂(mg/l) in Mansarovar Talab of Jeerapura (Dhar) Year July 2014- June 2015.

Month	Site 01	Site 02	Site 03	Site 04
Jul.	3.6	3.5	3.0	3.8
Aug.	4.2	4.5	3.9	4.0
Sep.	4.3	4.8	4.1	4.2
Oct.	6.4	6.2	6.0	6.5
Nov.	6.0	6.9	6.8	6.2
Dec.	3.8	3.9	3.2	3.0
Jan.	1.9	1.7	2.0	1.9
Feb.	2.8	2.5	2.9	2.8
Mar.	1.7	1.5	3.0	1.9
Apr.	1.2	1.8	1.6	2.0
May	2.5	2.9	2.9	3.0
Jun.	4.5	4.9	5.0	5.2

Table 14. Monthly Variations in Free CO₂ (mg/l) in Mansarovar Talab of Jeerapura (Dhar) Year July 2015- June 2016.

Month	Site 01	Site 02	Site 03	Site 04
Jul.	3.9	4.0	4.1	4.5
Aug.	4.4	4.8	4.8	4.6
Sep.	4.9	5.2	4.4	5.5
Oct.	6.9	7.2	6.2	7.0
Nov.	7.0	7.5	8.0	8.5
Dec.	4.8	4.9	6.2	5.8
Jan.	2.8	2.2	2.9	2.6
Feb.	3.0	3.5	3.2	3.8
Mar.	2.9	2.5	3.2	3.5
Apr.	1.9	2.5	2.8	2.7
May	2.5	3.5	3.5	4.0
Jun.	4.9	4.5	5.2	5.9

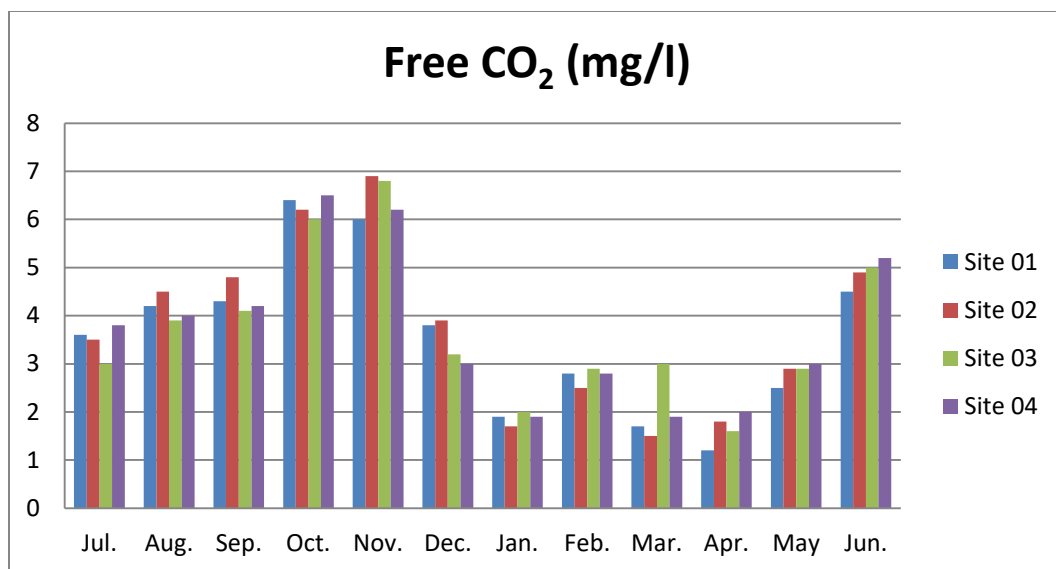


Fig. - 13. Monthly Variations in Free CO₂(mg/l) in Mansarovar Talab of Jeerapura (Dhar) Year July 2014- June 2015.

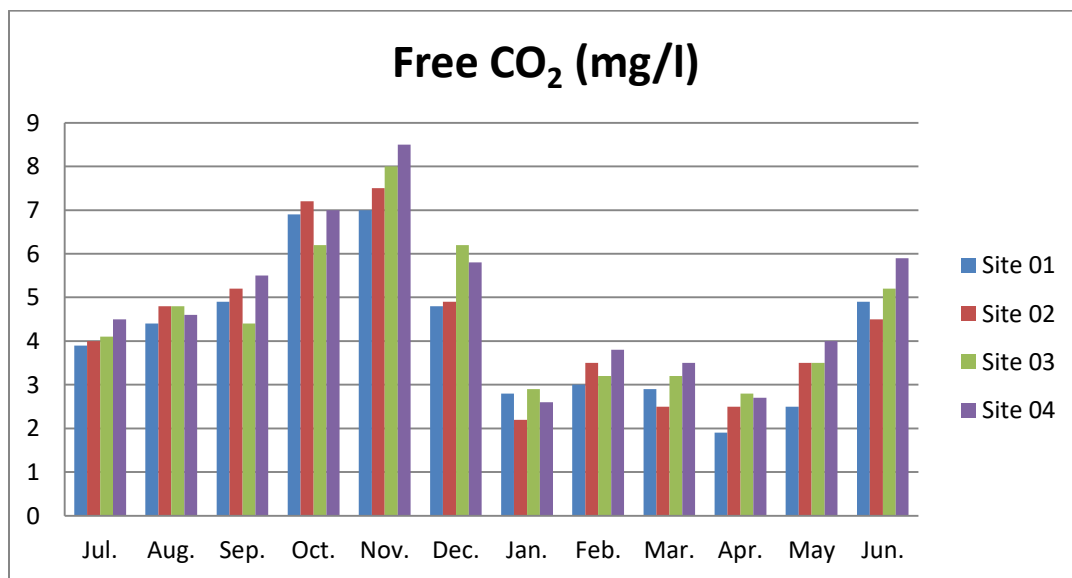


Fig. - 14. Monthly Variations in Free CO₂(mg/l) in Mansarovar Talab of Jeerapura (Dhar) Year July 2015- June 2016.

8. Total Alkalinity: -

July 2014 – June 2015

SamplingSite 01.The Total Alkalinity oscillated from 235.2mg/l to 475.0mg/l. The lowest Total Alkalinity was recorded in March where as highest was in November (Table 15& fig.15).

SamplingSite 02.At this station Total Alkalinity fluctuated between 225.4mg/l to 490.3mg/l. The lowest Total Alkalinity was recorded in March where as highest was in November (Table 15& fig.15).

SamplingSite 03.The Total Alkalinity oscillated between 255.0mg/l to 420.0mg/l. The lowest Total Alkalinity was recorded in March where as highest was in November (Table 15& fig.15).

SamplingSite 04.At this station the Total Alkalinity ranged between 245.9mg/l to 480.0/l. The lowest Total Alkalinity was recorded in March where as highest was in November (Table 15& fig.15).

July 2015 – June 2016

SamplingSite 01.The Total Alkalinity oscillated from 255.9mg/l to 490.0mg/l. The lowest Total Alkalinity was observed in March where as highest was in November (Table 16& fig.16).

SamplingSite 02.At this station Total Alkalinity fluctuated between 245.8mg/l to 499.0mg/l. The lowest Total Alkalinity was observed in March where as highest was in November (Table 16& fig.16).

SamplingSite 03.TheTotal Alkalinityocillated between 265.5mg/l to 478.0mg/l. The lowest Total Alkalinity was recorded in Marchwhere ashighest was in November (Table 16& fig.16).

SamplingSite 04.At this station the Total Alkalinity ocillated between 259.2mg/l to 460.0mg/l. The lowest Total Alkalinity was observed in Marchwhere ashighest was in November (Table 16& fig.16).

The value of total alkalinity is fluctuated from 225.4to 499.0 mg/l. The lowest value was observed at samplingsite 02 in March in the year of 2014-15. The highest value was observed at samplingsite 02 in November in the year of 2015-16 (Table 15 & fig.15 and Table 16 & fig.16).

**Table 15. Monthly Variations in Total Alkalinity (mg/l) in Mansarovar
Talab of Jeerapura (Dhar) Year July 2014 – June 2015.**

Month	Site 01	Site 02	Site 03	Site 04
Jul.	390.0	325.0	398.0	335.0
Aug.	365.5	375.8	355.0	360.2
Sep.	410.5	420.5	400.0	410.0
Oct.	435.5	385.8	405.0	375.2
Nov.	475.0	490.3	420.0	480.0
Dec.	300.5	290.0	315.5	300.0
Jan.	265.0	258.5	285.0	295.0
Feb.	272.5	265.9	290.0	285.5
Mar.	235.2	225.4	255.0	245.9
Apr.	280.5	270.2	290.8	290.0
May	255.0	269.2	275.6	288.4
Jun.	280.5	272.5	299.5	305.5

**Table 16. Monthly Variations in Total Alkalinity (mg/l) in Mansarovar
Talab of Jeerapura (Dhar) Year July 2015- June 2016.**

Month	Site 01	Site 02	Site 03	Site 04
Jul.	390.0	360.0	370.0	385.0
Aug.	385.0	395.0	395.3	405.2
Sep.	425.0	435.1	438.2	442.0
Oct.	455.0	435.0	465.0	455.0
Nov.	490.0	499.0	478.0	460.0
Dec.	405.0	435.0	425.0	448.0
Jan.	385.0	368.5	392.0	374.2
Feb.	292.5	285.2	302.5	325.5
Mar.	255.9	245.8	265.5	259.2
Apr.	290.5	305.2	299.2	315.7
May	325.0	340.2	345.0	360.0
Jun.	286.5	275.5	300.5	322.5

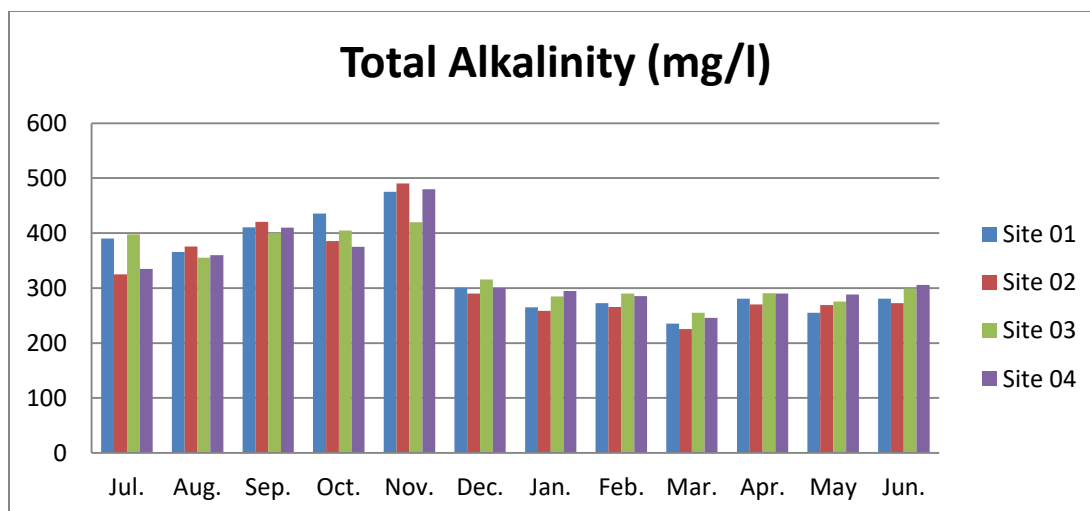


Fig. - 15. Monthly Variations in Total Alkalinity (mg/l) in Mansarovar Talab of Jeerapura (Dhar) Year July 2014 – June 2015.

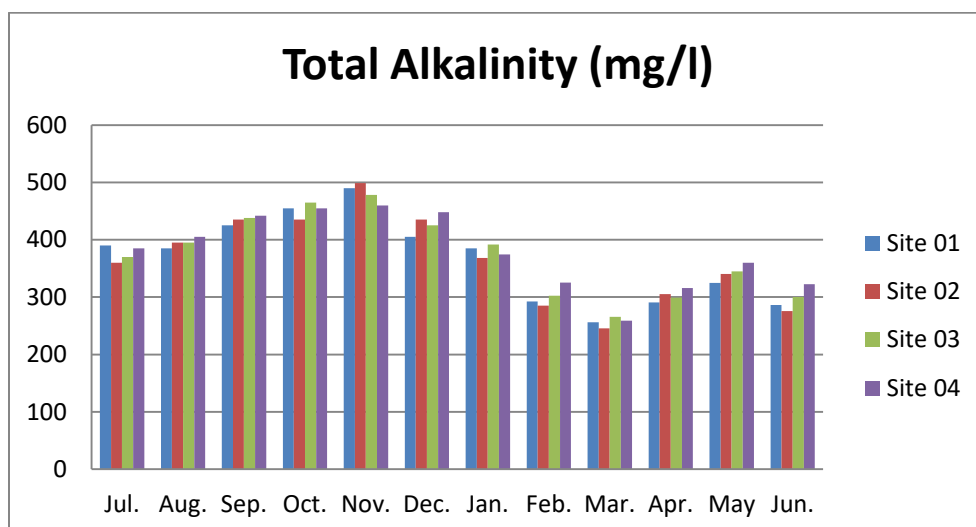


Fig. - 16. Monthly Variationss in Total Alkalinity (mg/l) in Mansarovar Talab of Jeerapura (Dhar) Year July 2015- June 2016.

9. Biochemical Oxygen Demand (B.O.D.): -

July 2014 – June 2015

SamplingSite 01.The B.O.D. varied from 2.0mg/l to 6.3mg/l. The lowest B.O.D. was recorded in August where as highest was in January (Table 17& fig.17).

SamplingSite 02.At this station B.O.D. fluctuated between 2.2mg/l to 6.7mg/l. The lowest B.O.D. was recorded in August where as highest was in January (Table 17& fig.17).

SamplingSite 03.The B.O.D. oscillated between 2.2mg/l to 5.9mg/l. The lowest B.O.D. was recorded in July and March where as highest was in January (Table 17& fig.17).

SamplingSite 04.At this station the B.O.D. ranged between 2.5mg/l to 6.0mg/l. The lowest B.O.D. was recorded in September where as highest was in January (Table 17& fig.17).

July 2015 – June 2016

SamplingSite 01.The B.O.D. varied from 3.5mg/l to 6.8mg/l. The lowest B.O.D. was recorded in March where as highest was in January (Table 18& fig.18).

SamplingSite 02.At this station B.O.D. fluctuated between 3.0mg/l to 6.9mg/l. The lowest B.O.D. was recorded in February where as highest was in January (Table 18& fig.18).

SamplingSite 03.The B.O.D. Oscillated between 2.9mg/l to 6.2mg/l. The lowest B.O.D. was recorded in March where as highest was in January (Table 18& fig.18).

SamplingSite 04.At this station the B.O.D. ranged between 3.5mg/l to 6.0mg/l. The lowest B.O.D. was recorded in February where as highest was in January (Table 18& fig.18).

The biochemical oxygen demand value fluctuated between 2.0 to 6.9 mg/l. The lowest biochemical oxygen demand value was observed at site 01 in August in the year 2014-15. The highest value was observed at site 02 in January in the year of 2015-16 (Table 17 & fig.17 and Table 18 & fig.18).

Table 17. Monthly Variations in Biochemical Oxygen Demand (mg/l) in Mansarovar Talab of Jeerapura (Dhar) Year July 2014- June 2015.

Month	Site 01	Site 02	Site 03	Site 04
Jul.	2.9	2.5	2.2	2.6
Aug.	2.0	2.2	2.8	2.6
Sep.	2.5	2.6	2.4	2.5
Oct.	2.8	3.0	2.8	3.8
Nov.	3.2	3.3	3.4	3.5
Dec.	2.7	2.4	3.8	3.2
Jan.	6.3	6.7	5.9	6.0
Feb.	5.0	3.0	5.2	3.8
Mar.	2.8	3.1	2.2	3.2
Apr.	3.8	4.3	4.8	4.0
May	4.7	5.1	3.9	3.0
Jun.	5.5	5.7	4.5	4.1

Table 18. Monthly Variations in Biochemical Oxygen Demand (mg/l) in Mansarovar Talab of Jeerapura (Dhar) Year July 2015- June 2016.

Month	Site 01	Site 02	Site 03	Site 04
Jul.	4.6	4.5	4.8	4.1
Aug.	4.8	4.2	4.5	4.0
Sep.	3.7	4.2	3.9	4.6
Oct.	4.9	4.6	5.2	5.6
Nov.	4.5	4.8	5.5	5.8
Dec.	5.0	5.9	5.6	5.9
Jan.	6.8	6.9	6.2	6.0
Feb.	5.0	3.0	5.8	3.5
Mar.	3.5	3.9	2.9	4.5
Apr.	4.2	4.8	4.9	5.2
May	4.9	5.5	5.8	5.0
Jun.	5.8	5.9	4.8	4.2

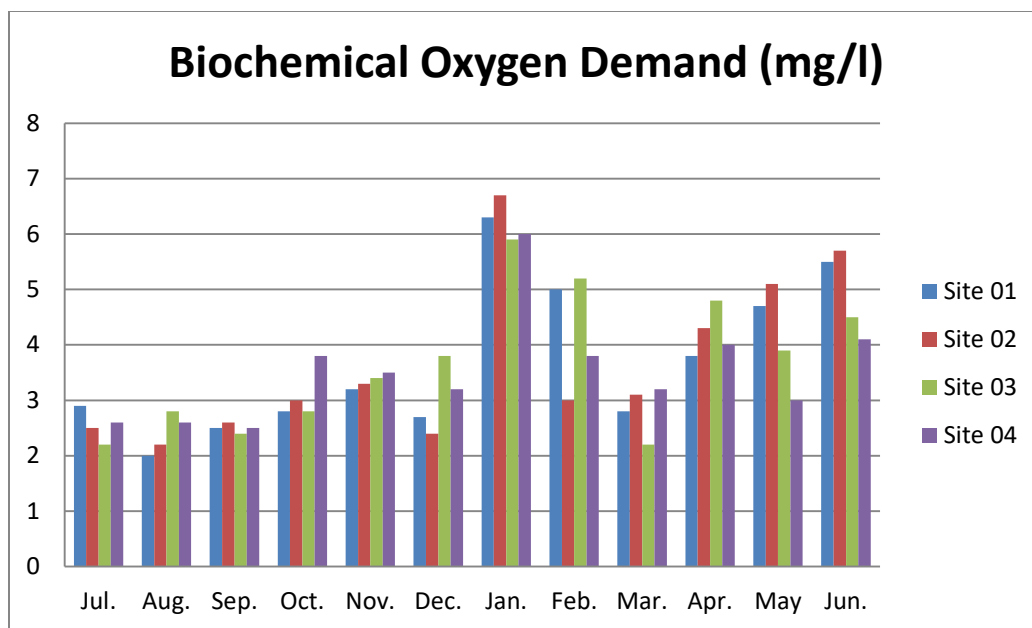


Fig. - 17. Monthly Variations in Biochemical Oxygen Demand (mg/l) in Mansarovar Talab of Jeerapura (Dhar) Year July 2014- June 2015.

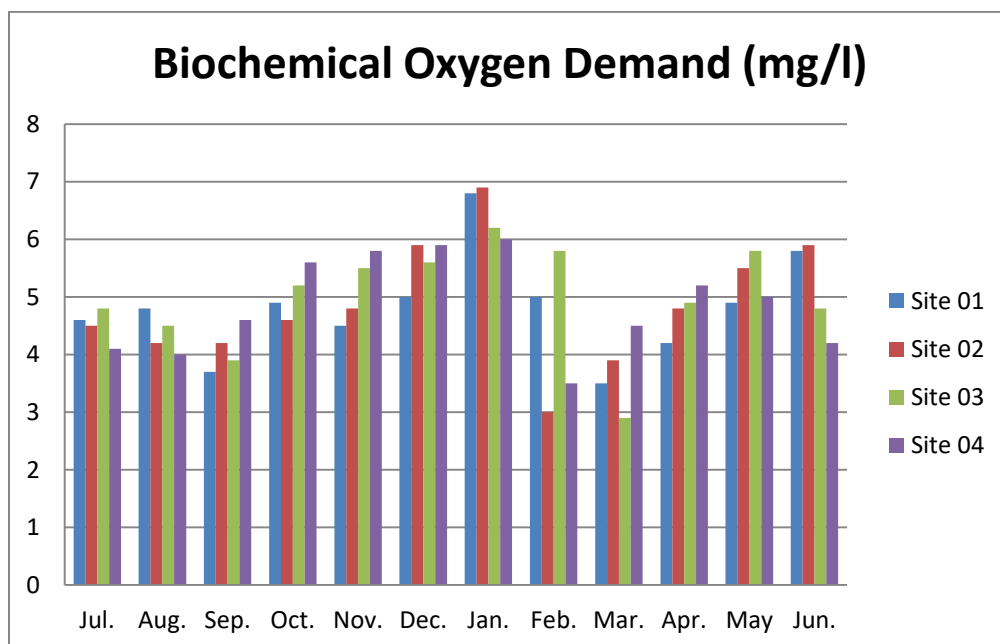


Fig.-18. Monthly Variations in Biochemical Oxygen Demand (mg/l) in Mansarovar Talab of Jeerapura (Dhar) Year July 2015- June 2016.

10. Total Hardness: -

July 2014 – June 2015

SamplingSite 01. The Total Hardness oscillated between 115.0mg/l to 240.0mg/l. The lowest Total Hardness was observed in February where as highest was in May (Table 19& fig.19).

SamplingSite 02. At this station Total Hardness varied from 105.0mg/l to 265.5mg/l. The lowest Total Hardness was observed in February where as highest was in November (Table 19& fig.19).

SamplingSite 03. The Total Hardness fluctuated between 130.0mg/l to 295.0mg/l. The lowest Total Hardness was observed in January where as highest was in June (Table 19& fig.19).

SamplingSite 04. At this station the Total Hardness varied from 145.8mg/l to 330.0mg/l. The lowest Total Hardness was observed in February where as highest was in June (Table 19& fig.19).

July 2015 – June 2016

SamplingSite 01. The Total Hardness fluctuated between 125.0mg/l to 290.0mg/l. The lowest Total Hardness was recorded in February where as highest was in June (Table 20& fig.20).

SamplingSite 02. At this station Total Hardness varied from 135.0mg/l to 290.0mg/l. The lowest Total Hardness was observed in February where as highest was in November (Table 20& fig.20).

SamplingSite 03.The Total Hardness fluctuated between 134.0mg/l to 300.0mg/l. The lowest Total Hardness was recorded in February where as highest was in June (Table 20 & fig.20).

SamplingSite 04.At this station the Total Hardness fluctuated between 155.0mg/l to 305.0mg/l. The lowest Total Hardness was observed in February where as highest was in December (Table 20 & fig.20).

It was fluctuated between 105.0 to 330.0 mg/l. The lowest total hardness was observed at sampling site 02 in February in the year of 2014-15 and highest total hardness at sampling site 04 in June in the year of 2014-15 (Table 19 & fig.19).

**Table 19. Monthly Variations in Total Hardness (mg/l) in Mansarovar
Talab of Jeerapura (Dhar) Year July 2014- June 2015.**

Month	Site 01	Site 02	Site 03	Site 04
Jul.	220.5	235.8	240.0	250.2
Aug.	195.8	198.5	205.5	215.0
Sep.	235.5	225.6	255.0	265.0
Oct.	190.0	175.2	205.0	225.0
Nov.	230.8	265.5	245.0	275.5
Dec.	220.5	210.9	245.5	230.2
Jan.	120.0	145.0	130.0	155.8
Feb.	115.0	105.0	135.0	145.8
Mar.	222.0	195.5	245.0	215.5
Apr.	225.8	218.8	255.0	262.9
May	240.0	232.0	260.0	275.0
Jun.	198.0	230.0	295.0	330.0

**Table 20. Monthly Variations in Total Hardness (mg/l) in Mansarovar
Talab of Jeerapura (Dhar) Year July 2015- June 2016.**

Month	Site 01	Site 02	Site 03	Site 04
Jul.	240.0	250.0	252.0	260.0
Aug.	220.0	215.0	235.0	225.0
Sep.	255.0	259.0	272.0	285.0
Oct.	220.0	195.0	233.0	215.0
Nov.	256.0	290.0	2680	300.0
Dec.	250.0	225.0	280.0	305.0
Jan.	130.0	165.0	145.0	177.0
Feb.	125.0	135.0	134.0	155.0
Mar.	220.0	205.5	240.0	225.8
Apr.	245.5	225.2	267.3	236.2
May	255.0	265.0	268.0	279.0
Jun.	290.0	285.0	300.0	291.0

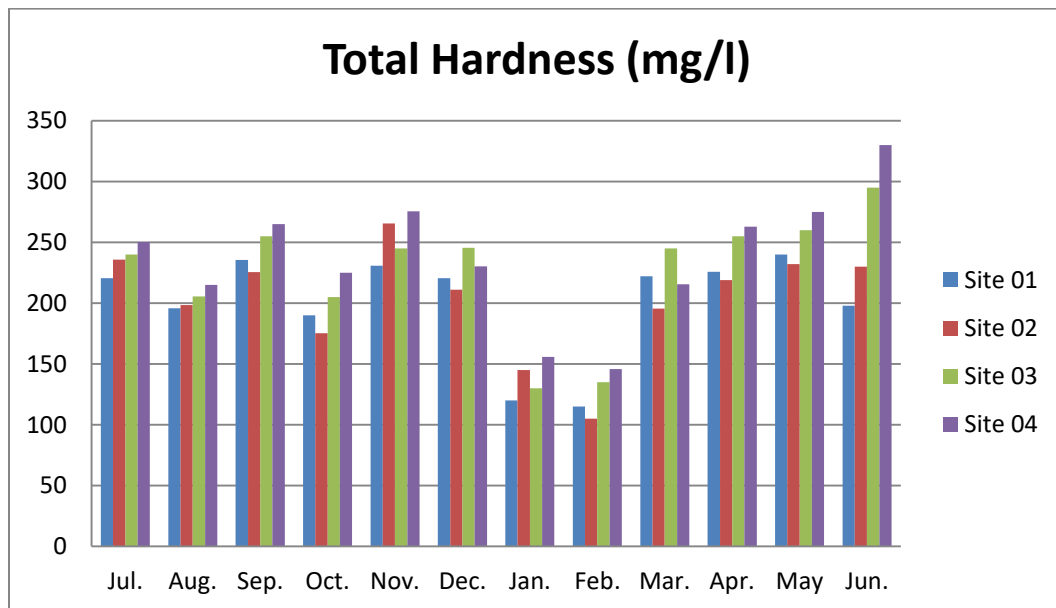


Fig. -19. Monthly Variations in TotalHardness (mg/l) in Mansarovar Talab of Jeerapura (Dhar) Year July 2014- June 2015.

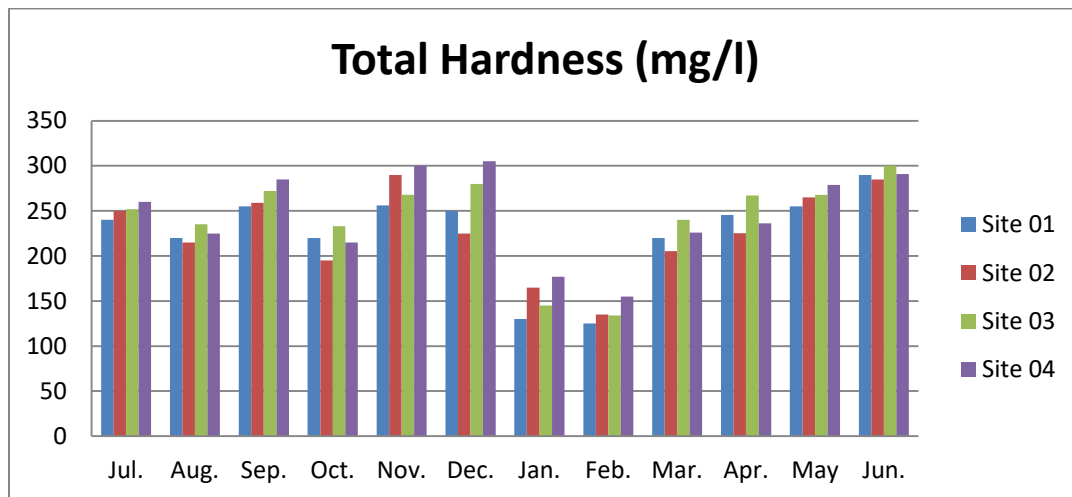


Fig. - 20. Monthly Variations inTotal Hardness (mg/l) in Mansarovar Talab of Jeerapura (Dhar) Year July 2015- June 2016.

11. Magnesium: -

July 2014 – June 2015

SamplingSite 01.The Magnesium ranged from 18.9mg/l to 35.5mg/l. The lowest Magnesium was recorded in January and May where as highest was in November (Table 21 & fig.21).

SamplingSite 02.At this station Magnesium fluctuated between 18.3mg/l to 40.5mg/l. The lowest Magnesium was recorded in February where as highest was in November (Table 21 & fig.21).

SamplingSite 03.The Magnesium oscillated between 18.5mg/l to 36.5mg/l. The lowest Magnesium was observed in February where as highest was in November (Table 21 & fig.21).

SamplingSite 04.At this station the Magnesium ranged between 19.9mg/l to 39.5mg/l. The lowest Magnesium was recorded in February where as highest was in November (Table 21 & fig.21).

July 2015 – June 2016

SamplingSite 01.The Magnesium varied from 18.9mg/l to 38.5mg/l. The lowest Magnesium was observed in February where as highest was in November (Table 22 & fig.22).

SamplingSite 02.At this station Magnesium fluctuated between 19.3mg/l to 43.5mg/l. The lowest Magnesium was observed in February where as highest was in November (Table 22 & fig.22).

SamplingSite 03. The Magnesium oscillated between 19.2mg/l to 39.5mg/l. The lowest Magnesium was observed in February where a highest was in November (Table 22 & fig.22).

SamplingSite 04. At this station the Magnesium ranged between 20.8mg/l to 44.0mg/l. The lowest Magnesium was recorded in February where a highest was in November (Table 22 & fig.22).

It was fluctuated between 18.3 to 44.0 mg/l. The lowest Magnesium was observed at sampling site 02 in February in the year 2014-2015 and highest range was observed at sampling site 04 in November in the year 2015-2016 (Table 21 & fig.21 and Table 22 & fig.22).

**Table 21. Monthly Variations in Magnesium (mg/l) in Mansarovar
Talab of Jeerapura (Dhar) Year July 2014- June 2015.**

Month	Site 01	Site 02	Site 03	Site 04
Jul.	30.5	32.9	31.8	33.5
Aug.	27.1	26.5	28.2	27.5
Sep.	27.1	28.9	28.4	29.5
Oct.	24.2	22.5	25.4	23.5
Nov.	35.5	40.5	36.5	39.5
Dec.	29.5	27.2	30.8	29.0
Jan.	18.9	20.5	19.5	22.2
Feb.	19.5	18.3	18.5	19.9
Mar.	21.5	22.0	22.7	23.5
Apr.	25.5	19.5	26.7	21.5
May	18.9	20.2	20.6	23.5
Jun.	25.1	26.5	27.3	28.8

**Table 22. Monthly Variations in Magnesium (mg/l) in Mansarovar
Talab of Jeerapura (Dhar) Year July 2015- June 2016.**

Month	Site 01	Site 02	Site 03	Site 04
Jul.	31.5	33.0	32.5	32.9
Aug.	28.5	27.9	29.5	28.2
Sep.	28.5	29.9	29.0	30.0
Oct.	26.2	24.5	28.0	25.5
Nov.	38.5	43.5	39.5	44.0
Dec.	32.5	35.2	33.8	36.7
Jan.	20.5	21.2	22.5	24.2
Feb.	18.9	19.3	19.2	20.8
Mar.	22.5	23.0	24.2	25.8
Apr.	26.2	21.5	27.5	24.5
May	22.9	21.2	25.2	23.5
Jun.	24.2	25.5	26.5	28.2

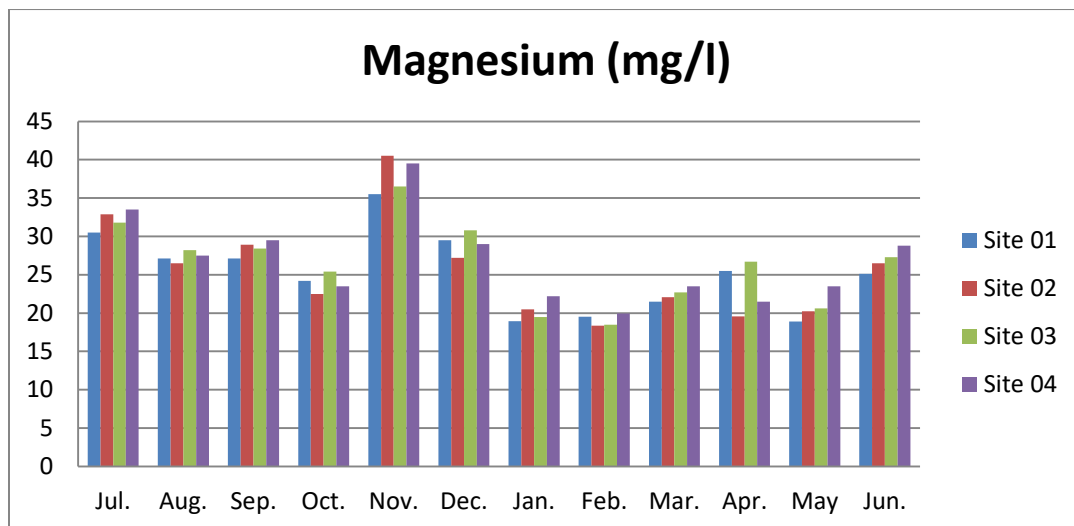


Fig. - 21. Monthly Variations in Magnesium (mg/l) in Mansarovar Talab of Jeerapura (Dhar) Year July 2014- June 2015.

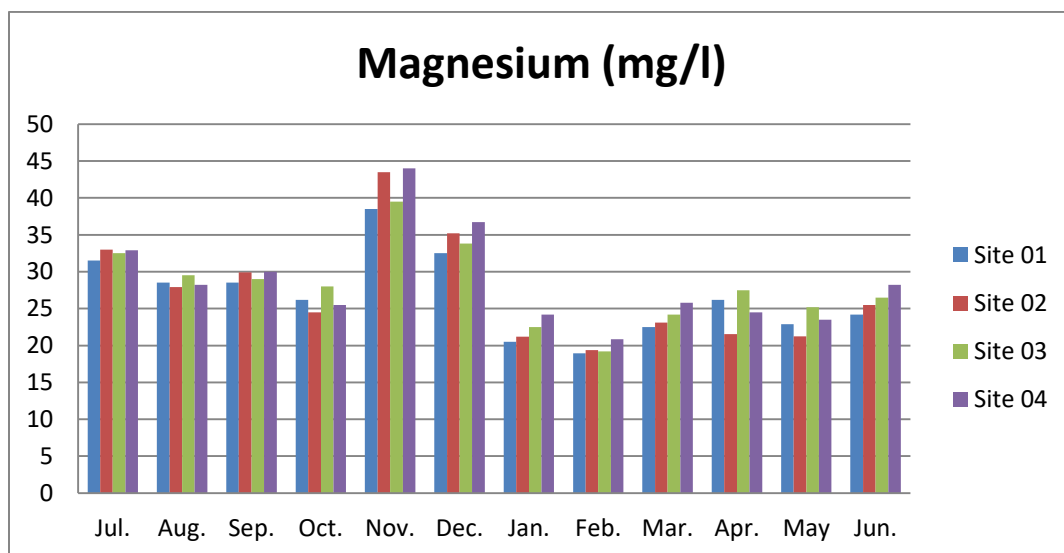


Fig. - 22. Monthly Variations in Magnesium (mg/l) in Mansarovar Talab of Jeerapura (Dhar) Year July 2015- June 2016.

12. Calcium: -

July 2014 – June 2015

SamplingSite 01.The Calcium varied from 35.5mg/l to 68.9mg/l. The lowest Calcium was recorded in August where as highest was in May (Table 23& fig.23).

SamplingSite 02.At this station Calcium fluctuated between 36.2mg/l to 72.4mg/l. The lowest Calcium was observed in August where as highest was in May (Table 23& fig.23).

SamplingSite 03.The Calcium fluctuated between 36.5mg/l to 69.0mg/l. The lowest Calcium was observed in August where as highest was in May (Table 23& fig.23).

SamplingSite 04.At this station the Calcium varied from 37.0mg/l to 74.2mg/l. The lowest Calcium was observed in August where as highest was in May (Table 23& fig.23).

July 2015 – June 2016

SamplingSite 01.The Calcium varied from 36.5mg/l to 68.2mg/l. The lowest Calcium was observed in August where as highest was in June (Table 24& fig.24).

SamplingSite 02.At this station Calcium fluctuated between 37.7mg/l to 70.4mg/l. The lowest Calcium was recorded in August where as highest was in May (Table 24& fig.24).

SamplingSite 03. The Calcium oscillated between 37.5mg/l to 69.8mg/l. The lowest Calcium was observed in August where as highest was in June (Table 24 & fig.24).

SamplingSite 04. At this station the Calcium ranged between 38.2mg/l to 72.5mg/l. The lowest Calcium was observed in August where as highest was in May (Table 24 & fig.24).

It was fluctuated between 35.5 to 74.2 mg/l. The lowest Calcium was observed at sampling site 01 in August in the year 2014-2015 and highest range was observed at sampling site 04 in May in the year 2014-2015 (Table 23 & fig.23).

Table 23. Monthly Variations in Calcium (mg/l) in Mansarovar Talab of Jeerapura (Dhar) Year July 2014- June 2015.

Month	Site 01	Site 02	Site 03	Site 04
Jul.	36.4	38.9	37.0	39.5
Aug.	35.5	36.2	36.5	37.0
Sep.	40.0	38.2	42.0	40.2
Oct.	41.2	44.5	43.2	45.5
Nov.	40.9	39.2	42.2	40.5
Dec.	46.2	44.5	48.0	42.5
Jan.	45.2	46.2	47.4	45.5
Feb.	47.5	49.1	48.2	50.2
Mar.	50.8	52.5	51.5	53.3
Apr.	54.6	55.7	55.5	56.7
May	68.9	72.4	69.0	74.2
Jun.	65.2	60.1	68.5	63.5

Table 24. Monthly Variations in Calcium (mg/l) in Mansarovar Talab of J

Month	Site 01	Site 02	Site 03	Site 04
Jul.	38.5	39.9	37.9	40.2
Aug.	36.5	37.7	37.5	38.2
Sep.	42.5	44.2	43.5	45.0
Oct.	45.2	47.5	46.4	48.5
Nov.	43.9	41.5	44.2	42.5
Dec.	48.2	46.5	49.0	45.5
Jan.	55.2	49.2	54.1	50.5
Feb.	46.5	45.2	47.5	48.2
Mar.	52.8	55.5	53.5	56.5
Apr.	56.6	59.7	57.5	60.5
May	67.9	70.4	68.7	72.5
Jun.	68.2	62.5	69.8	66.2

Y

ear July 2015- June 2016.

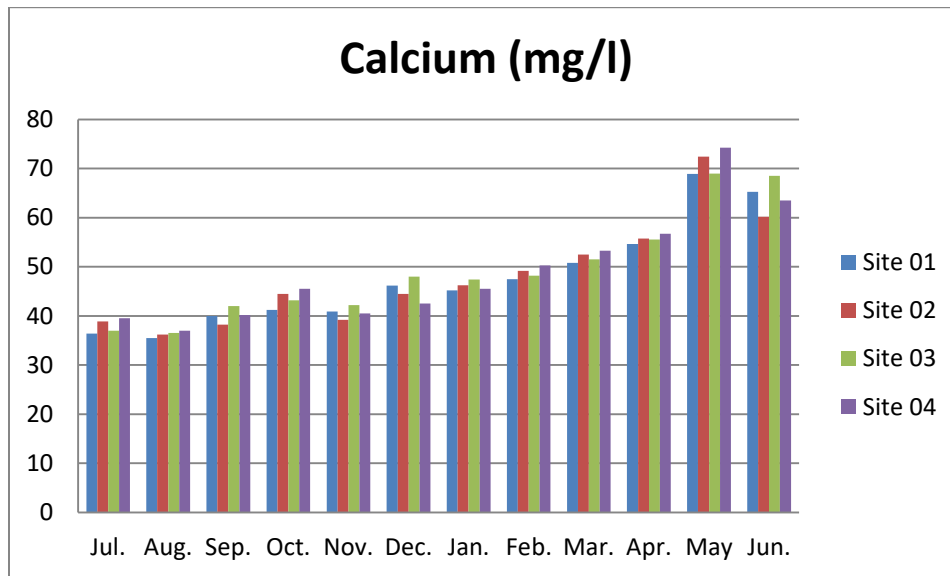


Fig. - 23. Monthly Variations in Calcium (mg/l) in Mansarovar Talab of Jeerapura (Dhar) Year July 2014- June 2015.

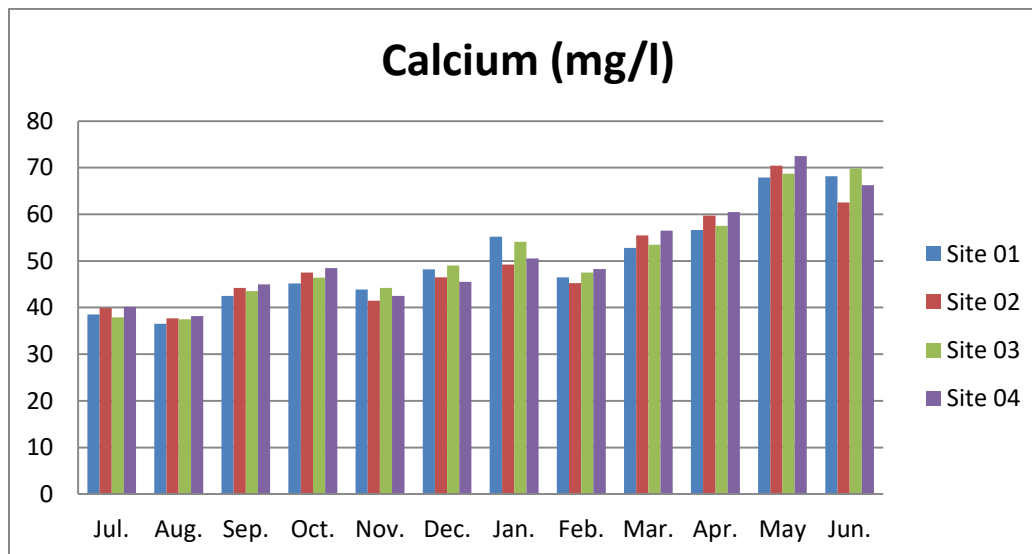


Fig. - 24. Monthly Variations in Calcium (mg/l) in Mansarovar Talab of Jeerapura (Dhar) Year July 2015- June 2016.

13. Chloride: -

July 2014 – June 2015

SamplingSite 01.The Chloride varied from 23.4mg/l to 60.1mg/l. The lowest Chloride was recorded in December where ashighest was in June (Table 25& fig.25).

SamplingSite 02.At this station Chloride fluctuated between 25.5mg/l to 65.2mg/l. The lowest Chloride was observed in October where ashighest was in June (Table 25& fig.25).

SamplingSite 03.The Chloride oscillated between 25.2mg/l to 62.5mg/l. The lowest Chloride was obsereved in December where ashighest was in June (Table 25& fig.25).

SamplingSite 04.At this station the Chloride ranged between 28.2mg/l to 69.3mg/l. The lowest Chloride was observed in Octoberwhere ashighest was in June (Table 25& fig.25).

July 2015 – June 2016

SamplingSite 01.The Chloride varied from 35.0mg/l to 59.6mg/l. The lowest Chloride was recorded in December where ashighest was in June (Table 26& fig.26).

SamplingSite 02.At this station Chloride fluctuated between 36.0mg/l to 62.2mg/l. The lowest Chloride was observed in December where ashighest was in July (Table 26& fig.26).

SamplingSite 03.The Chloride oscillated between 40.0mg/l to 59.0mg/l. The lowest Chloride was recorded in December where as highest was in June (Table 26& fig.26).

SamplingSite 04.At this station the Chloride ranged between 38.0mg/l to 64.0mg/l. The lowest Chloride was observed in December where as highest was in July (Table 26& fig.26).

It was fluctuated between 23.4 to 69.3 mg/l. The lowest range was observed at sampling site 01 in December in the year 2014-15 and highest range was at sampling site 04 in June in the year of 2014-15 (Table 25& fig.25).

**Table 25. Monthly Variations in Chloride (mg/l) in Mansarovar Talab
of Jeerapura (Dhar) Year July 2014- June 2015.**

Month	Site 01	Site 02	Site 03	Site 04
Jul.	45.5	58.2	42.0	55.5
Aug.	40.7	43.5	44.2	45.5
Sep.	44.5	41.2	47.5	42.2
Oct.	28.1	25.5	32.0	28.2
Nov.	32.5	26.8	35.5	28.5
Dec.	23.4	27.3	25.2	29.2
Jan.	40.5	45.2	42.5	47.2
Feb.	46.5	47.9	48.5	49.0
Mar.	48.8	49.1	50.2	50.9
Apr.	50.2	52.1	52.2	53.2
May	55.1	59.2	58.5	60.1
Jun.	60.1	65.2	62.5	69.3

**Table 26. Monthly Variations in Chloride (mg/l) in Mansarovar Talab
of Jeerapura (Dhar) Year July 2015- June 2016.**

Month	Site 01	Site 02	Site 03	Site 04
Jul.	48.0	62.2	50.0	64.0
Aug.	55.0	58.0	56.2	60.1
Sep.	45.4	50.0	54.0	52.0
Oct.	42.0	45.1	48.0	46.0
Nov.	48.0	39.0	50.0	42.2
Dec.	35.0	36.0	40.0	38.0
Jan.	47.5	48.2	48.5	49.2
Feb.	50.5	52.2	52.2	52.2
Mar.	51.5	50.1	52.2	52.1
Apr.	49.2	50.1	50.5	52.2
May	53.2	55.2	54.2	56.2
Jun.	59.6	61.2	59.0	62.5

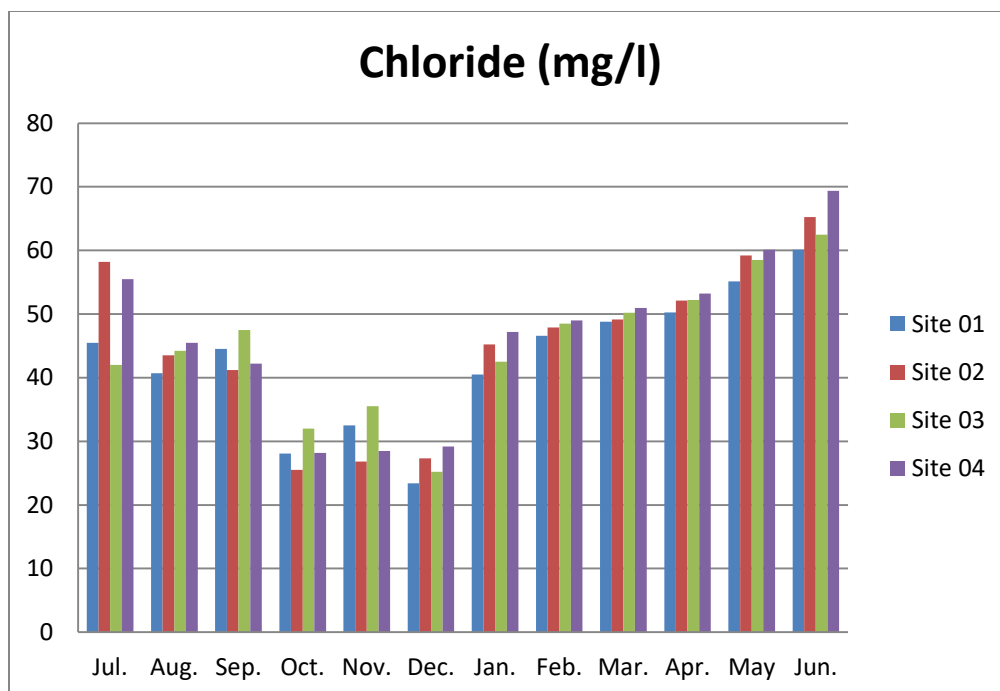


Fig. - 25. Monthly Variations in Chloride (mg/l) in Mansarovar Talab of Jeerapura (Dhar) Year July 2014- June 2015.

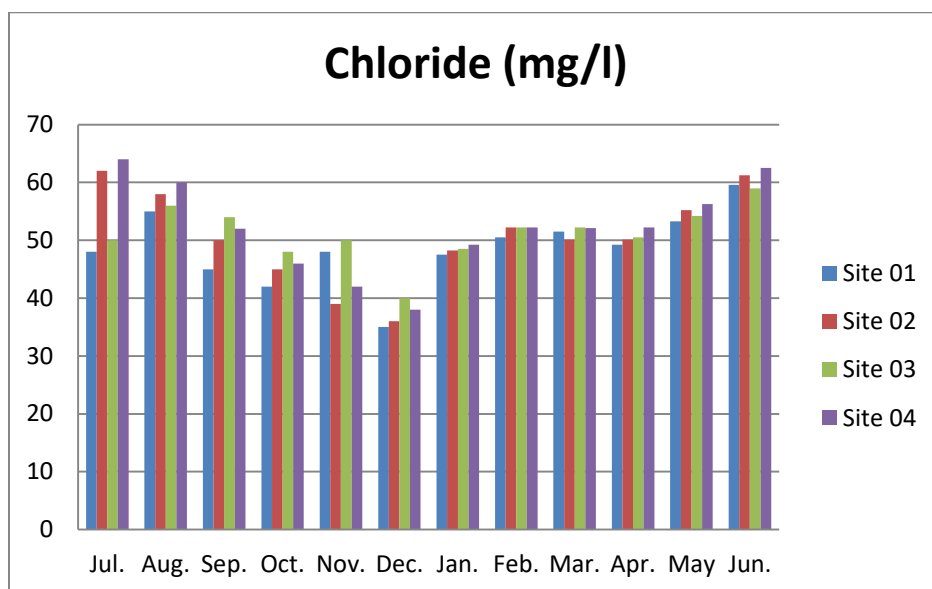


Fig. - 26. Monthly Variations in Chloride (mg/l) in Mansarovar Talab of Jeerapura (Dhar) Year July 2015- June 2016.

14.Nitrate: -

July 2014 – June 2015

SamplingSite 01.The Nitrate varied from 27.0mg/l to 48.0mg/l. The lowest Nitrate was recorded in November where ashighest was in June (Table 27& fig.27).

SamplingSite 02.At this station Nitrate fluctuated between 26.0mg/l to 50.3mg/l. The lowest Nitrate was observed in October where ashighest was in May (Table 27& fig.27).

SamplingSite 03.The Nitrate oscillated between 30.2mg/l to 51.3mg/l. The lowest Nitrate was observed in July where ashighest was in June (Table 27& fig.27).

SamplingSite 04.At this station the Nitrate ranged between 32.5mg/l to 54.0mg/l. The lowest Nitrate was observed in July where ashighest was in May (Table 27& fig.27).

July 2015 – June 2016

SamplingSite 01.The Nitrate varied from 35.0mg/l to 59.4mg/l. The lowest Nitrate was recorded in July where ashighest was in May (Table 28& fig.28).

SamplingSite 02.At this station Nitrate fluctuated between 38.0mg/l to 57.0mg/l. The lowest Nitrate was observed in July where ashighest was in April (Table 28& fig.28).

SamplingSite 03.The Nitrate oscillated between 38.0mg/l to 68.0mg/l. The lowest Nitrate was recorded in July where ashighest was in June (Table 28& fig.28).

SamplingSite 04.At this station the Nitrate ranged between 41.2mg/l to 65.1mg/l. The lowest Nitrate was observed in July where as highest was in May (Table 28& fig.28).

In general, Nitrate value varied from 26.0 to 68.0 mg/l. The lowest ranged was observed at sampling site 02 in month of October in the year of 2014-15 and highest ranged was observed at sampling site 03 in June in the year of 2015-16 (Table 27 & fig.27 and Table 28 & fig.28).

Table 27. Monthly Variations in Nitrate (mg/l) in Mansarovar Talab of Jeerapura (Dhar) Year July 2014- June 2015.

Month	Site 01	Site 02	Site 03	Site 04
Jul.	29.2	30.5	30.2	32.5
Aug.	32.8	34.4	33.5	35.7
Sep.	36.0	39.0	38.0	40.0
Oct.	28.0	26.0	32.0	36.0
Nov.	27.0	29.0	32.0	35.0
Dec.	35.0	37.0	39.0	36.2
Jan.	32.0	35.0	34.0	38.0
Feb.	38.0	40.4	41.0	43.0
Mar.	45.1	48.0	48.2	50.0
Apr.	45.0	48.0	47.0	52.3
May	47.2	50.3	50.0	54.0
Jun.	48.0	44.0	51.3	49.4

Table 28. Monthly Variations in Nitrate (mg/l) in Mansarovar Talab of Jeerapura (Dhar) Year July 2015- June 2016.

Month	Site 01	Site 02	Site 03	Site 04
Jul.	35.0	38.0	38.0	41.2
Aug.	42.0	45.0	44.2	46.0
Sep.	48.0	50.4	49.0	53.3
Oct.	55.0	45.0	58.0	47.0
Nov.	38.2	40.3	42.3	45.2
Dec.	45.0	45.0	48.0	50.0
Jan.	40.0	45.0	44.0	49.0
Feb.	47.5	49.1	48.0	51.1
Mar.	50.0	51.0	53.4	55.0
Apr.	55.0	57.0	58.0	59.0
May	59.4	55.2	62.1	65.1
Jun.	58.0	54.0	68.0	60.0

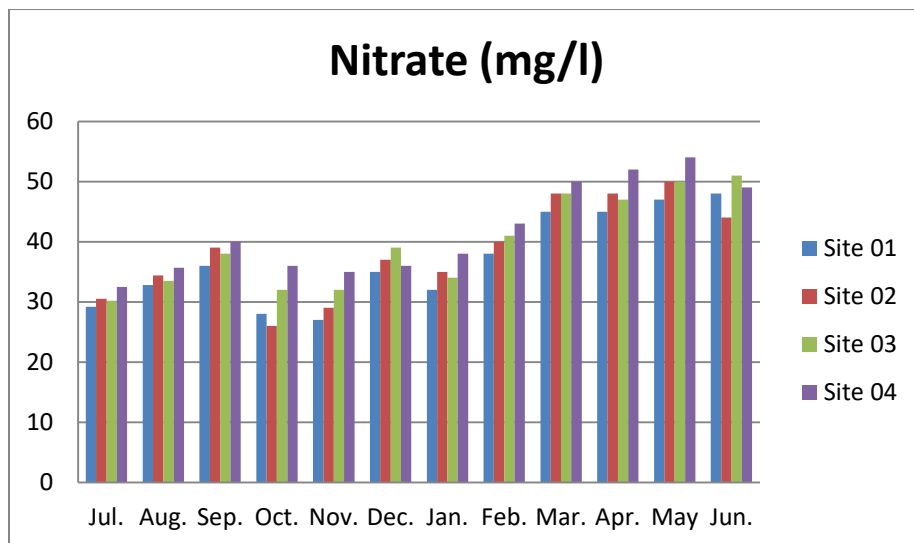


Fig. - 27. Monthly Variations in Nitrate (mg/l) in Mansarovar Talab of Jeerapura (Dhar) Year July 2014- June 2015.

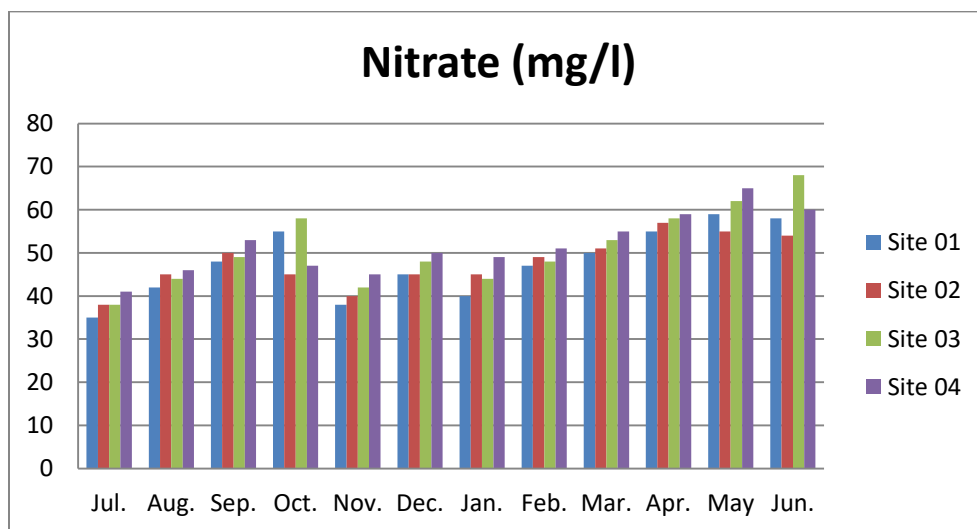


Fig. - 28. Monthly Variations in Nitrate (mg/l) in Mansarovar Talab of Jeerapura (Dhar) Year July 2015- June 2016.

15. Phosphate: -

July 2014 – June 2015

SamplingSite 01.The Phosphate varied from 1.18mg/l to 2.38mg/l. The lowest Phosphate was observed in August and October where a highest was in May (Table 29& fig.29).

SamplingSite 02.At this station Phosphate fluctuated between 1.15mg/l to 2.50mg/l. The lowest Phosphate was observed in June where a highest was in May (Table 29& fig.29).

SamplingSite 03.The Phosphate oscillated between 1.95mg/l to 3.80mg/l. The lowest Phosphate was observed in July and November where a highest was in June (Table 29& fig.29).

SamplingSite 04.At this station the Phosphate ranged between 2.15mg/l to 3.75mg/l. The lowest Phosphate was observed in November where a highest was in Many (Table 29& fig.29).

July 2015 – June 2016

SamplingSite 01.The Phosphate varied from 1.40mg/l to 2.50mg/l. The lowest Phosphate was observed in January where a highest was in November (Table 30& fig.30).

SamplingSite 02.At this station Phosphate fluctuated between 1.45mg/l to 2.65mg/l. The lowest Phosphate was observed in January where a highest was in November (Table 30& fig.30).

SamplingSite 03. The Phosphate oscillated between 1.60mg/l to 3.50mg/l. The lowest Phosphate was recorded in July where as highest was in May (Table 30 & fig.30).

SamplingSite 04. At this station the Phosphate ranged between 1.90mg/l to 3.68mg/l. The lowest Phosphate was observed in July where as highest was in May (Table 30 & fig.30).

It was fluctuated between 1.15 to 3.80 mg/l. The lowest range observed at sampling site 02 in June in the year of 2014-15. The highest range at sampling site 03 in June in the year of 2014-15 (Table 29 & fig.29).

Table 29. Monthly Variations in Phosphate (mg/l) in Mansarovar Talab of Jeerapura (Dhar) Year July 2014- June 2015.

Month	Site 01	Site 02	Site 03	Site 04
Jul.	1.25	1.52	1.95	2.20
Aug.	1.18	1.25	2.30	2.45
Sep.	1.25	1.35	2.25	2.35
Oct.	1.18	1.25	2.10	2.35
Nov.	1.46	1.55	1.95	2.15
Dec.	1.70	1.85	2.90	2.75
Jan.	1.20	1.48	2.30	2.40
Feb.	1.61	1.75	2.69	2.85
Mar.	1.87	1.90	2.99	3.10
Apr.	2.15	2.24	3.55	3.20
May	2.38	2.50	3.55	3.75
Jun.	1.58	1.15	3.80	2.95

Table 30. Monthly Variations in Phosphate (mg/l) in Mansarovar Talab of Jeerapura (Dhar) Year July 2015- June 2016.

Month	Site 01	Site 02	Site 03	Site 04
Jul.	1.50	1.70	1.60	1.90
Aug.	1.80	1.95	2.20	2.50
Sep.	2.10	2.25	2.80	2.45
Oct.	2.35	2.45	2.55	2.68
Nov.	2.50	2.65	2.72	2.95
Dec.	1.85	2.00	2.55	2.60
Jan.	1.40	1.45	1.90	2.15
Feb.	1.55	1.65	2.55	2.65
Mar.	1.85	1.95	2.85	2.95
Apr.	2.20	2.35	3.05	3.25
May	2.45	2.51	3.50	3.68
Jun.	2.10	1.95	3.10	3.55

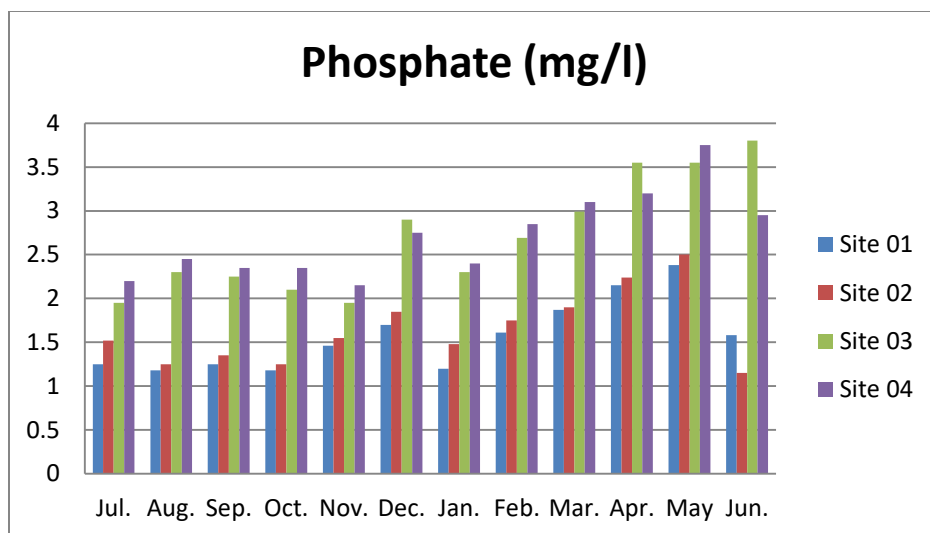


Fig. -29. Monthly Variations in Phosphate (mg/l) in Mansarovar Talab of Jeerapura (Dhar) Year July 2014- June 2015.

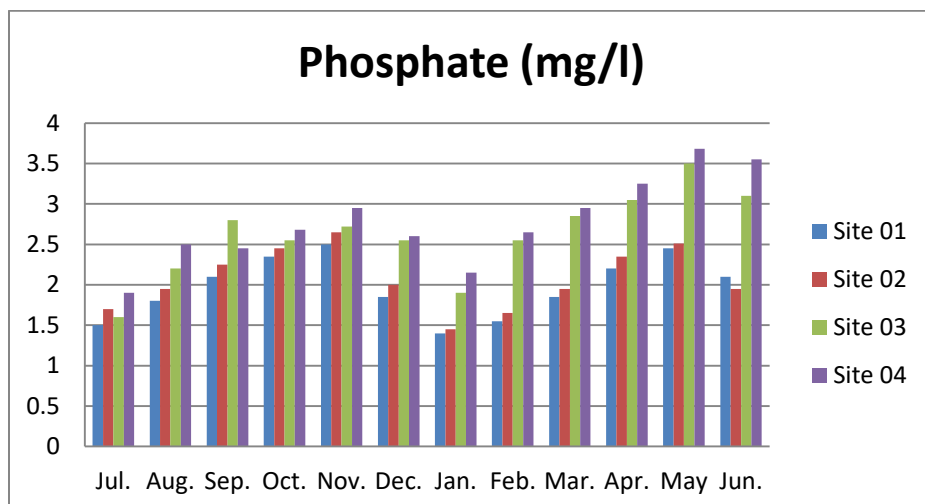


Fig. - 30. Monthly Variations in Phosphate (mg/l) in Mansarovar Talabof Jeerapura (Dhar) Year July 2015- June 2016.

16. Sulphate: -

July 2014 – June 2015

SamplingSite 01.The Sulphate varied from 5.3 mg/l to 9.6 mg/l. The lowest Sulphate was observed in February where as highest was in September (Table 31& fig.31).

SamplingSite 02.At this station Sulphate fluctuated between 4.9 mg/l to 9.6 mg/l. The lowest Sulphate was observed in February where as highest was in September (Table 31& fig.31).

SamplingSite 03.The Sulphate varied from 5.5 mg/l to 8.6 mg/l. The lowest Sulphate was observed in February where as highest was in September (Table 31& fig.31).

SamplingSite 04.At this station the Sulphate varied from 5.2 mg/l to 9.4 mg/l. The lowest Sulphate was observed in February where as highest was in September (Table 31& fig.31).

July 2015 – June 2016

SamplingSite01.The Sulphate varied from 6.0 mg/l to 9.5 mg/l. The lowest Sulphate was observed in January where as highest was in September (Table 32& fig.32).

SamplingSite 02.At this station Sulphate varied from 7.2 mg/l to 9.8 mg/l. The lowest Sulphate was observed in May where as highest was in October (Table 32& fig.32).

SamplingSite 03.The Sulphate varied from 7.0 mg/l to 9.6 mg/l. The lowest Sulphate was observed in February where a highest was in September (Table 32& fig.32).

SamplingSite 04.At this station the Sulphate varied from 7.1 mg/l to 9.9 mg/l. The lowest Sulphate was observed in February where a highest was in September (Table 32& fig.32).

It was fluctuated between 4.9 to 9.9 mg/l. The lowest value was observed at sampling site 02 in February in the year of 2014-15. The highest value was at sampling site 04 in September in the year of 2015-16 (Table 31& fig.31 and Table 32& fig.32).

**Table 31. Monthly Variations in Sulphate (mg/l) in Mansarovar Talab
of Jeerapura (Dhar) Year July 2014- June 2015.**

Month	Site 01	Site 02	Site 03	Site 04
Jul.	8.9	9.0	6.9	8.8
Aug.	9.4	9.4	8.3	9.2
Sep.	9.6	9.6	8.6	9.4
Oct.	8.3	8.5	8.2	8.2
Nov.	7.5	7.9	7.8	7.7
Dec.	6.9	6.8	7.4	6.9
Jan.	6.7	6.6	5.8	6.5
Feb.	5.3	4.9	5.5	5.2
Mar.	5.8	6.2	6.5	5.8
Apr.	6.2	6.4	5.8	6.4
May	6.5	6.6	6.3	6.6
Jun.	6.9	6.8	6.8	7.0

**Table 32. Monthly Variations in Sulphate (mg/l) in Mansarovar Talab
of Jeerapura (Dhar) Year July 2015- June 2016.**

Month	Site 01	Site 02	Site 03	Site 04
Jul.	9.0	9.2	8.9	8.8
Aug.	9.2	9.4	9.0	8.9
Sep.	9.5	9.6	9.6	9.9
Oct.	8.8	9.8	8.6	8.5
Nov.	8.6	9.2	8.4	8.3
Dec.	7.2	9.5	8.9	8.8
Jan.	6.0	8.4	7.8	7.7
Feb.	6.6	8.6	7.0	7.1
Mar.	7.2	8.5	8.4	8.3
Apr.	6.4	8.4	8.6	9.4
May	6.8	7.2	8.2	8.8
Jun.	7.2	7.5	8.7	9.2

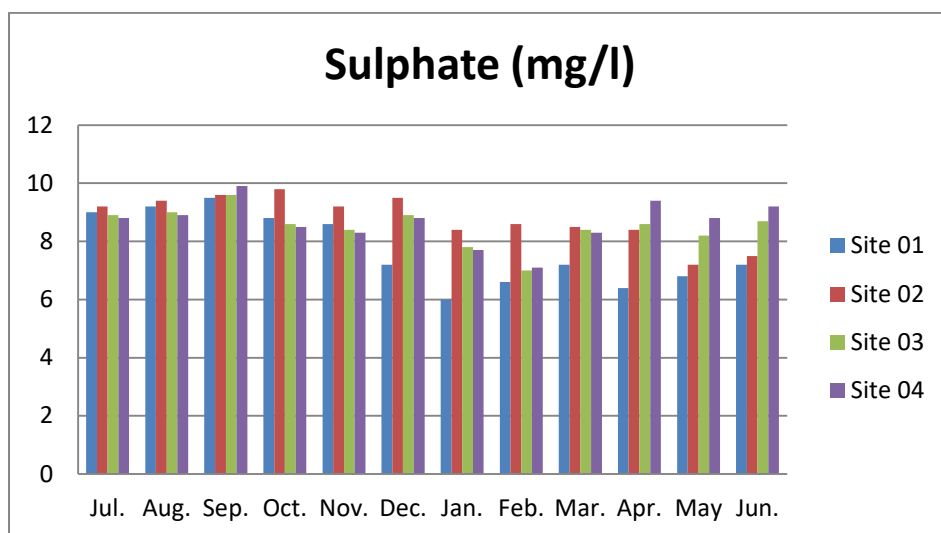


Fig. -31. Monthly Variations in Sulphate (mg/l) in Mansarovar Talab of Jeerapura (Dhar) Year July 2014- June 2015.

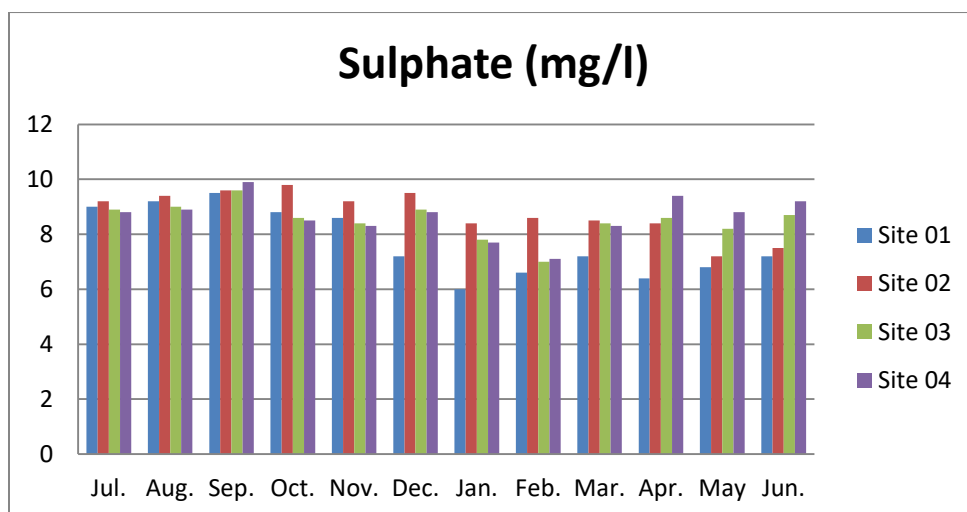


Fig. - 32. Monthly Variations in Sulphate (mg/l) in Mansarovar Talab of Jeerapura (Dhar) Year July 2015- June 2016.

17. Total Coli Form (MPN/100 ml): -

July 2014 – June 2015

SamplingSite 01.The Total Coli Form oscillated between 300 to 530 (MPN/100 ml). The lowest Total Coli Form was recorded in July where ashighest was in June (Table 33& fig. 33).

SamplingSite 02.At this station Total Coli Form value fluctuated between 320 to 570 (MPN/100 ml). The lowest Total Coli Form was observed in July where ashighest was in June (Table 33& fig.33).

SamplingSite 03.The Total Coli Form oscillated between 330 to 590 (MPN/100 ml). The lowest Total Coli Form was recorded in July where ashighest was in June (Table 33& fig. 33).

SamplingSite 04.At this station the Total Coli Form ranged between 340 to 650 (MPN/100 ml). The lowest Total Coli Form was observed in July where ashighest was in May (Table 33& fig. 33).

July 2015 – June 2016

SamplingSite 01.The Total Coli Form oscillated between 355 to 625 (MPN/100 ml). The lowest Total Coli Form was observed in July where ashighest was in December (Table 34& fig. 34).

SamplingSite 02.At this station Total Coli Form fluctuated between 350 to 620 (MPN/100 ml). The lowest Total Coli Form was recorded in January where ashighest was in December (Table 34& fig. 34).

SamplingSite 03. The Total Coli Form oscillated between 358 to 645 (MPN/100 ml). The lowest Total Coli Form was observed in July where as highest was in December (Table 34 & fig. 34).

SamplingSite 04. At this station the Total Coli Form ranged between 402 to 660 (MPN/100 ml). The lowest Total Coli Form was recorded in July where as highest was in December (Table 34 & fig. 34).

It was fluctuated between 300 to 660 (MPN/100 ml). The lowest range was observed at sampling site 01 in July in the year 2014-15. The highest range was observed at sampling site 04 in December in the year of 2015-16 (Table 33 & fig. 33 and Table 34 & fig. 34).

Table 33. Monthly Variations in Total Coli Form (MPN/100 ml) in Mansarovar Talab of Jeerapura (Dhar) Year July 2014- June 2015.

Month	Site 01	Site 02	Site 03	Site 04
Jul.	300	320	330	340
Aug.	355	370	375	390
Sep.	395	410	400	420
Oct.	430	455	450	478
Nov.	420	436	439	495
Dec.	475	490	485	500
Jan.	360	350	400	380
Feb.	395	410	425	450
Mar.	425	450	475	500
Apr.	470	485	550	565
May	490	510	520	650
Jun.	530	570	590	570

Table 34. Monthly Variations in Total Coli Form (MPN/100 ml) in Mansarovar Talab of Jeerapura (Dhar) Year July 2015- June 2016.

Month	Site 01	Site 02	Site 03	Site 04
Jul.	355	395	358	402
Aug.	420	450	435	470
Sep.	495	500	505	520
Oct.	520	550	540	580
Nov.	560	575	594	615
Dec.	625	620	645	660
Jan.	380	350	485	455
Feb.	415	420	465	490
Mar.	445	460	515	550
Apr.	480	492	580	592
May	500	525	620	625
Jun.	541	510	585	610

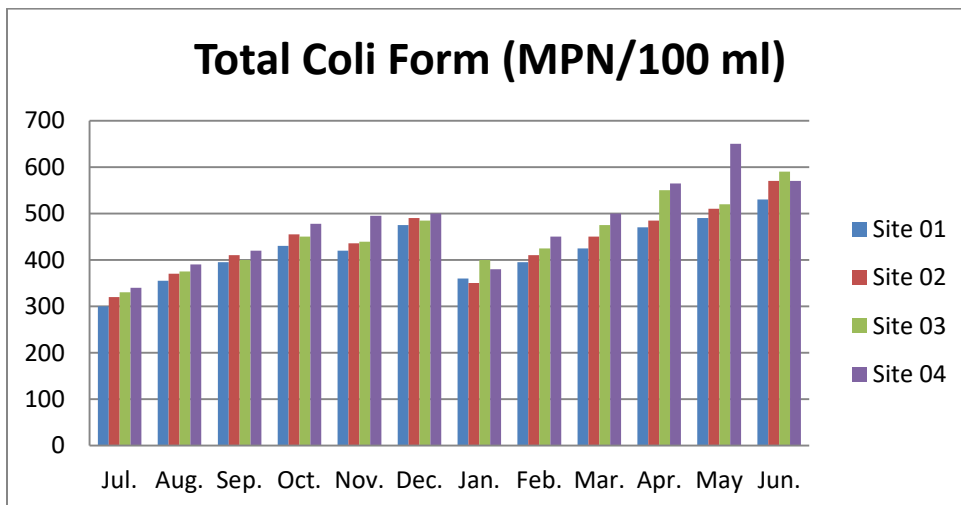


Fig.-33. Monthly Variations in Total Coli Form (MPN/100 ml) in Mansarovar Talab of Jeerapura (Dhar) Year July 2014- June 2015.

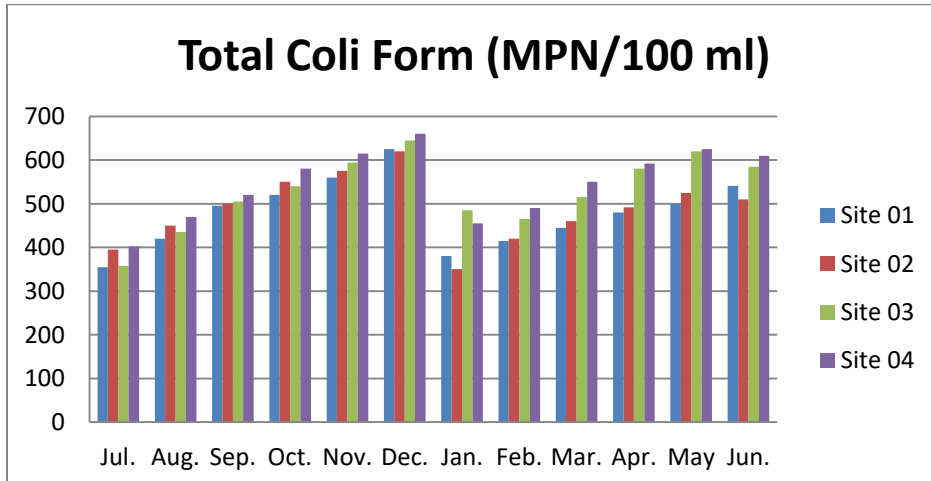


Fig.-34. Monthly Variations in Total Coli Form (MPN/100 ml) in Mansarovar Talab of Jeerapura (Dhar) Year July 2015- June 2016.

18. Fecal Coli Form (MPN/100 ml): -

July 2014 – June 2015

SamplingSite 01.The Fecal Coli Form oscillated between 145 to 365 (MPN/100 ml). The lowest Fecal Coli Form was observed in July where ashighest was in June (Table 35& fig.35).

SamplingSite 02.At this station Fecal Coli Form fluctuated between 162 to 380 (MPN/100 ml). The lowest Fecal Coli Form was recorded in July where ashighest was in June (Table 35& fig.35).

SamplingSite 03.The Fecal Coli Form oscillated between 155 to 390 (MPN/100 ml). The lowest Fecal Coli Form was observed in July where ashighestwas in May (Table 35& fig.35).

SamplingSite 04.At this station the Fecal Coli Form ranged between 170 to 410 (MPN/100 ml). The lowest Fecal Coli Form was observed in July where ashighest was in May (Table 35& fig.35).

July 2015 – June 2016

SamplingSite 01.The Fecal Coli Form oscillated between 170 to 385 (MPN/100 ml). The lowest Fecal Coli Form was recorded in January where ashighest was in December (Table 36& fig.36).

SamplingSite 02.At this station Fecal Coli Form fluctuated between 199 to 398 (MPN/100 ml). The lowest Fecal Coli Form was observed in January where ashighest was in December (Table 36& fig.36).

SamplingSite 03.The Fecal Coli Formoscillated between 220 to 455 (MPN/100 ml). The lowest Fecal Coli Form was recorded in July where as highest was in June (Table 36 & fig.36).

SamplingSite 04.At this station the Fecal Coli Form ranged between 250 to 480 (MPN/100 ml). The lowest Fecal Coli Form was observed in July where as highest was in June (Table 36 & fig.36).

It was fluctuated between 145 to 480 (MPN/100 ml). The lowest range was observed at sampling site 01 in July in the year 2014-15. The highest range was observed at sampling site 04 in June in the year of 2015-16 (Table 35 & fig.35 and Table 36 & fig. 36).

Table-35. Monthly Variations in Fecal Coli Form (MPN/100 ml) in Mansarovar Talab of Jeerapura (Dhar) Year July 2014- June 2015.

Month	Site 01	Site 02	Site 03	Site 04
Jul.	145	162	155	170
Aug.	178	192	180	199
Sep.	215	222	225	242
Oct.	242	235	255	265
Nov.	260	272	280	295
Dec.	284	295	280	325
Jan.	180	190	288	295
Feb.	210	240	245	270
Mar.	255	280	295	310
Apr.	300	335	330	355
May	350	330	390	410
Jun.	365	380	385	350

Table -36. Monthly Variations in Fecal Coli Form (MPN/100 ml) in Mansarovar Talab of Jeerapura (Dhar) Year July 2015- June 2016.

Month	Site 01	Site 02	Site 03	Site 04
Jul.	180	210	220	250
Aug.	220	240	235	266
Sep.	260	280	275	297
Oct.	275	295	315	345
Nov.	320	340	362	381
Dec.	385	398	401	418
Jan.	170	199	225	266
Feb.	220	250	290	320
Mar.	262	290	355	390
Apr.	320	355	380	385
May	370	390	395	420
Jun.	345	360	455	480

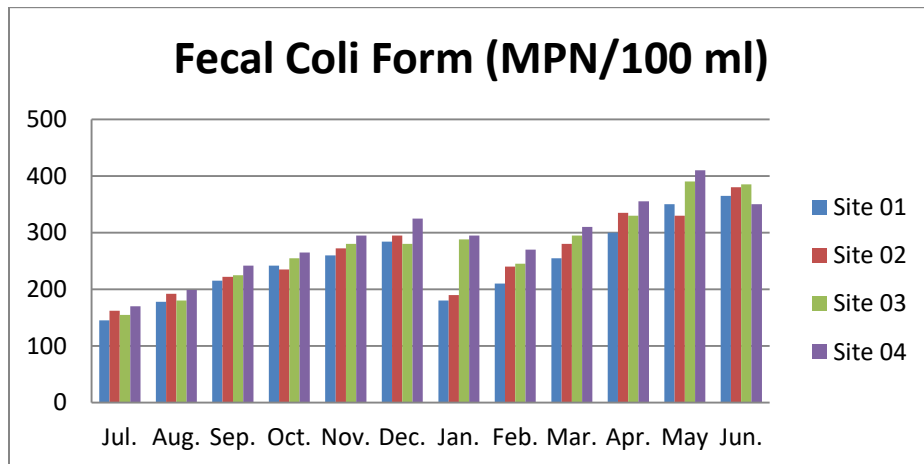


Fig.-35. Monthly Variations in Fecal Coli Form (MPN/100 ml) in Mansarovar Talab of Jeerapura (Dhar) Year July 2014- June 2015.

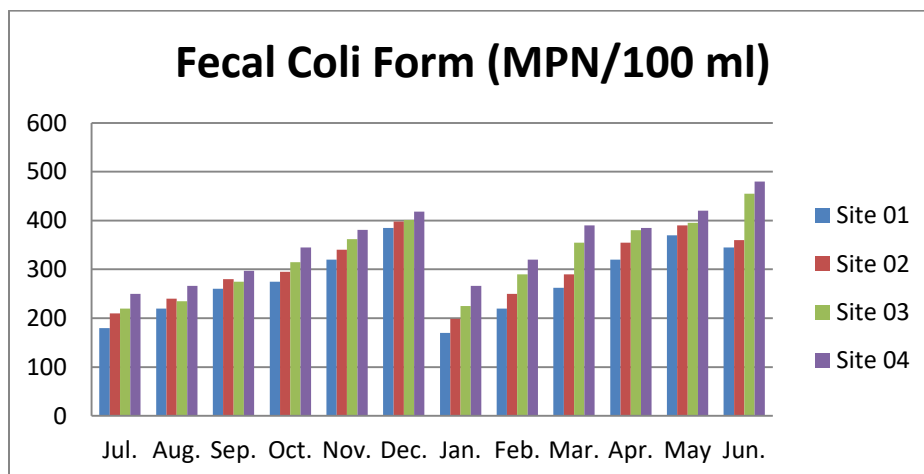


Fig.-36. Monthly Variations in Fecal Coli Form (MPN/100 ml) in Mansarovar Talab of Jeerapura (Dhar) Year July 2015- June 2016.

Table -37. Monthly variation in Diversity of Macro-invertebrates reported in Mansarovar Talab of Jeerapura (Dhar) Year July 2014 to June 2015.

S.No.	Macro-invertebrates (Bottom fauna)	July	Aug.	Sep.	Oct.	Nov.	Dec.	Jan.	Feb.	March	April	May	June	Total	Remarks
1	Oligochaeta	67	59.4	50.1	43	45.1	42.5	32	30.3	31	41.5	49.8	61.4	554 553.1	I
2	Hirudinea (Leeches)	4.2	3.3	2	2.3	2.2	2	4.3	3.1	3	3.8	3.5	4	37.7	VI
3	Gastropoda	6.5	7.9	5.1	7.3	8.8	12.9	15	17.2	26.7	28.5	19.2	8.2	163	III
4	Pelecypoda (Bivalvia)	4.9	6.9	4.3	3.6	2.8	3.1	4.2	9.7	7.3	5.3	4.8	5.9	62.8	IV
5	Insecta	11	14.2	28.5	34	30.9	33.9	36	30.9	24.8	15.6	16.2	13.5	288 289.5	II
6	Ostracoda	2.2	3.9	6.5	5.2	3	2.5	4.5	4.8	3.5	2.4	2.2	2	42.7	V
7	Shrimps	2.8	2.5	2	1.9	1.7	2	3	2.8	2.2	1.9	2.5	3	28.3	VII
8	Miscellaneous	1.8	1.9	1.5	2.8	5.5	1.1	1.5	1.2	1.5	1	1.8	2	23.6	VIII

Table -38. Monthly variation in Diversity of Macro-invertebrates reported in Mansarovar Talab of Jeerapura (Dhar) Year July 2015 to June 2016.

S.No.	Macro-invertebrates (Bottom fauna)	July	Aug.	Sep.	Oct.	Nov.	Dec.	Jan.	Feb.	March	April	May	June	Total	Remarks
1	Oligochaeta	70	60.8	43.5	40	44.1	39.2	36	32.5	30.3	46.3	62.9	68.7	575 574.3	I
2	Hirudinea (Leeches)	5.4	3.1	3.4	3	2.1	2.5	1.8	1.5	0.7	1.5	1.5	7.5	34	V
3	Gastropoda	8.5	11.2	8.7	8.2	9.3	8.2	14	19.8	25.6	29.6	16.5	6.2	166	III
4	Pelecypoda (Bivalvia)	4.8	3.2	3.5	2.5	1.3	1.5	2.5	4.8	5.8	4.2	2.1	4.2	40.4	IV
5	Insecta	6.3	12.9	35.8	37	34.9	42.8	40	33.2	31.5	11.5	12.3	8.2	307 306.4	II
6	Ostracoda	1.3	3.5	2.5	3.2	1.2	2.3	2.9	3.2	3	2.2	1.9	1.2	28.4	VI
7	Shrimps	1.8	2.8	1.5	1.2	2.2	1.5	2.2	3.4	1.2	1.9	2.3	3.5	25.5	VII
8	Miscellaneous	1.7	2.5	1.1	4.4	4.9	2	0.2	1.6	1.9	2.8	0.5	0.5	24.1	VIII

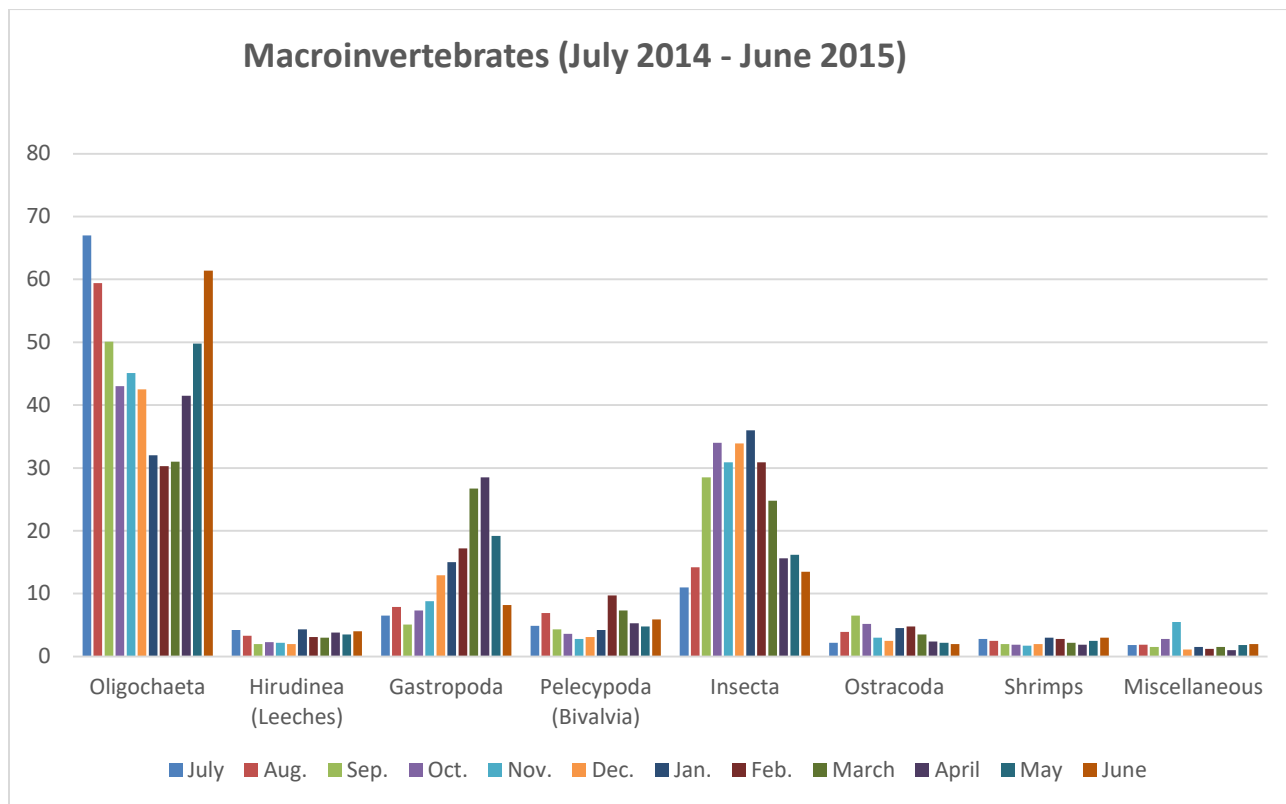


Fig.-37. Monthly variation in Diversity of Macro-invertebrates reported in Mansarovar Talab of Jeerapura (Dhar) Year July 2014 to June 2015.

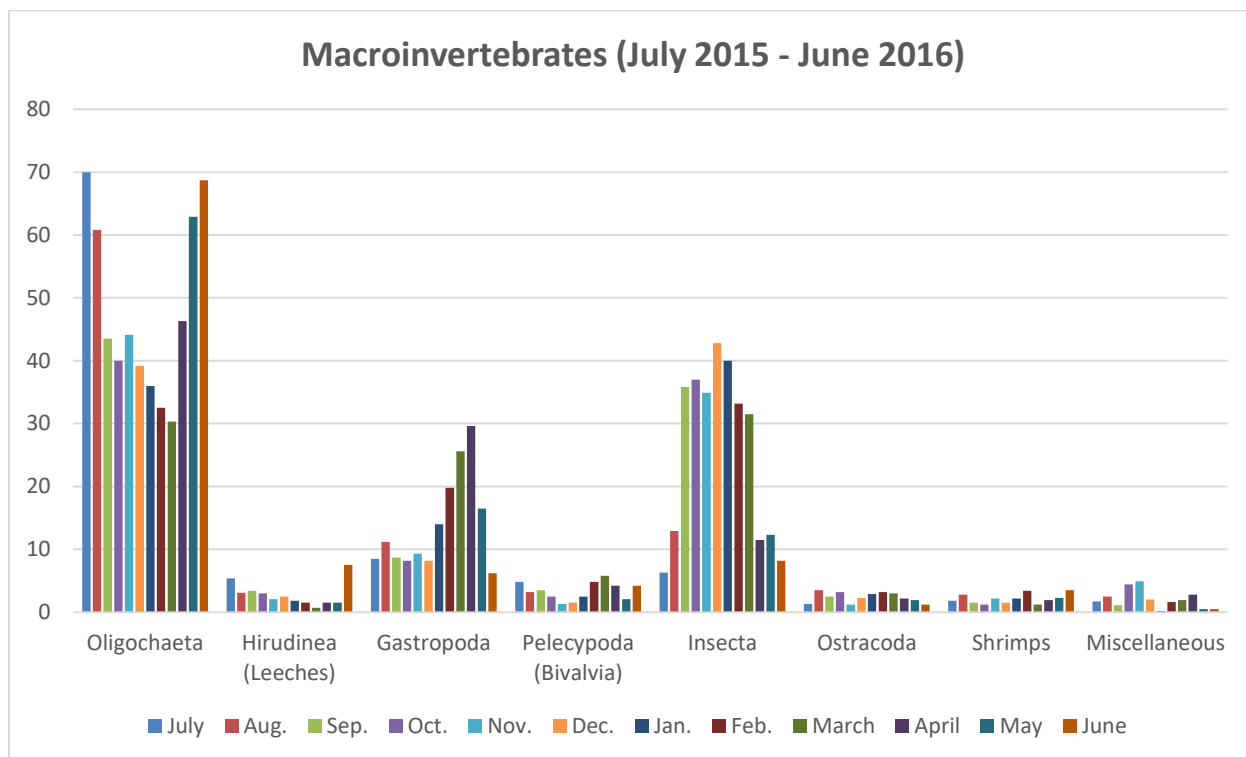


Fig.-38. Monthly variation in Diversity of Macro-invertebrates reported in Mansarovar Talab of Jeerapura (Dhar) Year July 2015 to June 2016.

Biological analysis

Table -39. List of Benthic Macro-invertebrates species reported in Mansarovar Talab from July 2014 to June 2016.

Phylum	Class	Species
Annelida	Oligocheata	<i>1. Tubifex tubifex</i> <i>2. Chaetogaster species</i> <i>3. Nais simplex</i> <i>4. Derolimnosa</i> <i>5. Branchiurusowerbyi</i> <i>6. Stylariafossularis</i> <i>7. Dero digitata</i> <i>8. Srachiondrilussemperi</i> <i>9. Branchiodrilussemperi</i>
	Hirudinea	<i>10. Helobdella species</i> <i>11. Glossiphonia species</i>
Mollusca	Gastropoda	<i>12. Planorbis species</i> <i>13. Limnaeaauricularia</i> <i>14. Limnaeaacumainata</i> <i>15. Vivipara bengalensis</i> <i>16. Viviperaoxytropsis</i> <i>17. Bellamya bengalensis</i> <i>18. Pisidium clarkeanum</i> <i>19. Digoniostomapulchella</i> <i>20. Melanoidestuberculatus</i> <i>21. Melanoideslineatus</i>
	Pelecypoda (Bivalvia)	<i>22. Lamellidensmarginalis</i> <i>23. Lamellidensconsobrinus</i>
Arthropoda	Insecta	<i>24. Chironomus phumosus</i> <i>25. Chaoborus species</i> <i>26. Baetis simplex</i> <i>27. Berosus species</i>
	Crustacea	<i>28. Daphnia cercinata</i> <i>29. Dephnia magna</i> <i>30. Cyclopes</i> <i>31. Nauplius</i> <i>32. Cypris</i>

Table -40. Depthwise Status of Benthic Macro-invertebrates.

Group	Macro-invertebrates	0.2m	0.5m	1m	1.5m
1. Phylum Annelida (A) Class-Oligochaeta	1. <i>Tubifex tubifex</i>	RA	RA	RA	RA
	2. <i>Chaetogaster species</i>	I	I	I	SR
	3. <i>Nais simplex</i>	I	I	I	I
	4. <i>Dero limnosa</i>	I	I	I	NR
	5. <i>Branchiura sowerbyi</i>	I	I	RA	RA
	6. <i>Stylaria fossularis</i>	RA	RA	RA	RA
	7. <i>Dero digitata</i>	RA	RA	RA	RA
	8. <i>Sratchiondrilus semperi</i>	I	I	NR	NR
	9. <i>Branchiodrilus semperi</i>	I	I	NR	NR
	10. <i>Helobdella species</i>	NR	NR	NR	ILA
(B) Class-Hirudinea (Leeches) 2. Phylum Mollusca (A) Class-Gastropoda	11. <i>Glossiphonia species</i>	RA	RA	RA	SR
	12. <i>Planorbis species</i>	RA	RA	SR	NR
	13. <i>Limnaea auricularia</i>	RA	RA	SR	NR
	14. <i>Limnaea acuminata</i>	I	I	I	I
	15. <i>Vivipara bengalensis</i>	RA	RA	RA	ILA
	16. <i>Vivipera oxytropis</i>	RA	RA	RA	ILA
	17. <i>Bellamya bengalensis</i>	RA	ILA	RA	RA
	18. <i>Pisidium clarkeanum</i>	ILA	ILA	NR	NR
	19. <i>Digoniostoma pulchella</i>	ILA	ILA	NR	NR
	20. <i>Melanoides tuberculatus</i>	RA	RA	RA	RA
	21. <i>Melanoides lineatus</i>	RA	RA	RA	RA

3. Phylum Arthropoda (A) Class-Insecta (B) Class-Crustacea 1. Branchipoda (Shrimps)	22. <i>Lamellidens marginalis</i>	ILA	ILA	ILA	-
	23. <i>Lamellidens consobrinus</i>	I	I	RA	-
	24. <i>Chironomus phumosus</i>	I	I	I	I
	25. <i>Chaoborus species</i>	ILA	ILA	I	-
	26. <i>Baetis simplex</i>	RA	RA	RR	RR
	27. <i>Berosus species</i>	RA	RA	I	I
	28. <i>Daphnia cercinata</i>	RA	IR	-	-
	29. <i>Dephnia magna</i>	ILA	ILA	-	-
	30. <i>Cyclopes</i>	ILA	ILA	ILA	ILA
	31. <i>Nauplius</i>	ILA	ILA	ILA	ILA
	32. <i>Cypris</i>	ILA	ILA	-	-
		ILA	-	-	-
		RLA	ILA	-	-

Key:

RA = Regular and abundant.

I = Irregular

- = Absent

ILA = Irregular and less abundant

NR = Not Recorded

RLA = Regular & less abundant

**Photographs showing the species of Annelids in Mansarovar Talab
Jeerapura (Dhar).**



Tubifex tubifex *Chaetogaster species*



Dero limnosa *Branchiura sowerbyi*



Stylaria fossularis Derodigitate



Glossiphonia species

**Photographs showing the species of Molluscans in Mansarovar
Talab Jeerapura (Dhar).**



Limnaea auricularia *Limnaea acuminata*



Vivipara bengalensis *Bellamya bengalensis*



Pisidium clarkeanum *Melanoidestuberculatus*



Lamellidens marginalis *Lamellidens consobrinus*

**Photographs showing the species of Arthropods Mansarovar Talab
Jeerapura (Dhar).**



Baetis simplex *Berosus species*



Daphnia cercinata

Daphnia magna



Cyclops

Nauplius



Cypris

Table -41. Karl Pearsons coefficient of correlation between Physico-chemical parameters and Macro Invertebrates and Fishes at Station-I

Parameter	Macro Invertebrates	Fishes
Temperature	0.37	0.21
Ph	-0.96	-0.97
BOD	0.83	0.72
Fecal Coliform	-0.11	0.07
Nitrate	0.16	-0.01
Phosphate	-0.20	-0.05
DO % Saturation	-0.29	-0.46
Turbidity	-0.93	-0.88
Total Solids	-0.83	-0.74

Table -42. Karl Pearsons coefficient of correlation between Physico-chemical parameters and Macro Invertebrates and Fishes at Station-II

Parameter	Macro Invertebrates	Fishes
Temperature	0.37	0.34
Ph	-0.96	-0.97
BOD	0.90	0.88
Fecal Coliform	-0.13	-0.11
Nitrate	0.23	0.19
Phosphate	-0.14	-0.13
DO % Saturation	-0.32	-0.35
Turbidity	-0.94	-0.94
Total Solids	-0.84	-0.83

Table -43. Karl Pearsons coefficient of correlation between Physico-chemical parameters and Macro Invertebrates and Fishes at Station-III

Parameter	Macro Invertebrates	Fishes
Temperature	0.41	0.38
Ph	-0.86	-0.89
BOD	0.98	0.96
Fecal Coliform	-0.25	-0.21
Nitrate	-0.17	-0.22
Phosphate	0.26	0.29
DO % Saturation	-0.16	-0.21
Turbidity	-0.86	-0.83
Total Solids	-0.70	-0.68

Table -44. Karl Pearsons coefficient of correlation between Physico-chemical parameters and Macro Invertebrates and Fishes at Station-IV

Parameter	Macro Invertebrates	Fishes
Temperature	0.51	0.49
Ph	-0.92	-0.94
BOD	0.75	0.71
Fecal Coliform	-0.01	0.06
Nitrate	0.10	0.04
Phosphate	0.07	0.11
DO % Saturation	-0.18	-0.25
Turbidity	-0.97	-0.97
Total Solids	-0.90	-0.89

Table -45. Seasonal Variation in fishes of Mansarovar Talab of Jeerapura (Dhar) Year Jul. 2014 – June 2015.

2014-15									
Family	Species	Jul.-Sep. (Monsoon)	Oct.-Dec. (Post Monsoon)	Jan.-march (Winter)	Apr.-June (Summer)	Jul.- Sep. (Monsoon)	Oct.-Dec. (Post Monsoon)	Jan.-march (Winter)	Apr.-June (Summer)
Cyprinidae	<i>Labeorohita</i>	+	++	+++	+	+	+	+++	+
	<i>Labeocalbasu</i>	+	-	+++	-	+	++	+++	+
	<i>Labeobata</i>	-	+	-	++	-	++	++	-
	<i>Cirrhinus mrigale</i>	-	+	+++	++	+	+	+++	+
	<i>Puntius sarana</i>	-	+	++	-	-	+	++	-
	<i>Puntius ticto</i>	+	-	+++	++	-	-	++	-
	<i>Cyprinus carpio</i>	-	-	-	+	-	+	-	-
	<i>Rasbora daniconius</i>	+	+	++	+	+	++	+++	-
	<i>Catlacatla</i>	+	++	+++	++	+	+	+++	+
Bagridae	<i>Mystus bleekari</i>	+	++	+++	+	+	+	+++	+
	<i>Mystus seenghala</i>	+	+	+++	-	+	-	+++	++
Mastacembelidae	<i>Mastacembelus armatus</i>	+	++	++	-	+	+	+++	+
	<i>Mastacembelus pancalus</i>	+	+++	++	+++	+	++	++	+
Notopteridae	<i>Notopterus notopterus</i>	-	+	-	++	-	++	++	-
anabantidae	<i>Colisa fasciatus</i>	+	+	+	-	-	-	+	-
Heteropneustidae	<i>Heteropneustes fossilis</i>	+	+++	++	+++	+	++	++	+
Nemacheilidae	<i>Noemacheilus botia</i>	+	+	+++	++	-	++	++	+
Clariidae	<i>Clarias batrachus</i>	-	+	++	-	-	+	++	+
Ambassidae	<i>Chanda nama</i>	+	+	++	+	+	+	+	+
Ophiocephalidae	<i>Channa punctatus</i>	-	-	-	+	-	-	+	-
Clupeidae	<i>Hilsa hilsa</i>	+	+	-	+	-	-	-	+
Nandidae	<i>Nandus nandus</i>	-	+	-	-	-	+	-	-
Siluridae	<i>Wallago attu</i>	+	+	++	++	+	++	+	++

Indication: +++ (Abundant), ++ (Less Abundant, Always Visible), + (Rare, Sometime Visible), - (Absent).

Table -46. Seasonal Variation in fishes of Mansarovar Talab of Jeerapura (Dhar) Year Jul. 2015– June 2016.

2015-16									
Family	Species	Jul.-Sep. (Monsoon)	Oct.-Dec. (Post Monsoon)	Jan.-march (Winter)	Apr.-June (Summer)	Jul.- Sep. (Monsoon)	Oct.-Dec. (Post Monsoon)	Jan.-march (Winter)	Apr.-June (Summer)
Cyprinidae	<i>Labeorohita</i>	+	+	+++	++	+	+	+++	+
	<i>Labeocalbasu</i>	+	-	+++	+	+	++	+++	-
	<i>Labeobata</i>	-	-	++	+	-	+	+	-
	<i>Cirrhinusmrigale</i>	+	+	+++	+	+	++	+++	+
	<i>Puntius sarana</i>	+	+	+++	-	-	+	++	-
	<i>Puntius ticto</i>	+	-	++	-	-	-	++	-
	<i>Cyprinus carpio</i>	-	-	-	+	+	-	-	-
	<i>Rasbora daniconius</i>	-	-	++	-	-	-	-	-
	<i>Catlacatla</i>	+	+	+++	+	+	++	++	+
Bagridae	<i>Mystusbleekari</i>	+	+	+++	++	+	+	++	+
	<i>Mystusseenghala</i>	+	-	+++	-	+	-	+++	-
Mastacembelidae	<i>Mastacembalusarmatus</i>	+	-	+++	+	+	-	++	-
	<i>Mastacembeluspancalus</i>	+	++	+	++	+	++	++	++
Notopteridae	<i>Notopterusnotopterus</i>	-	-	++	+	-	+	+	-
anabantidae	<i>Colisa fasciatus</i>	-	-	-	-	-	-	-	-
Heteropneustidae	<i>Heteropneustesfossillis</i>	+	++	+	++	+	++	++	+++
Nemacheilidae	<i>Noemacheilusbotia</i>	-	-	-	-	-	-	-	-
Clariidae	<i>Clariasbatrachus</i>	-	-	+++	++	-	+	+++	++
Ambassidae	<i>Chanda nama</i>	-	-	++	+	-	-	-	-
Ophiocephalidae	<i>Channa punctatus</i>	-	-	-	-	-	-	-	-
Clupeidae	<i>Hilsa hilsa</i>	+	+	-	-	-	++	+	-
Nandidae	<i>Nandus nandus</i>	-	-	-	-	-	-	-	-
Siluridae	<i>Wallago attu</i>	+	+	++	++	+	++	++	+

Indication:+++ (Abundant), ++ (Less Abundant, Always Visible),+ (Rare, Sometime Visible), - (Absent).

Table -47. Monthly Variation in Shannon & Weaver index and Simpson's Diversity Index during the study period (July 2014 to June 2016).

Months	Shannon & Weaver Index		Simpson's diversity Index	
	2014-2015	2015-2016	2014-2015	2015-2016
July	1.91	1.93	0.83	0.84
Aug.	1.33	1.37	0.59	0.61
September	1.37	1.26	0.67	0.63
October	1.85	1.99	0.82	0.85
Nov.	1.37	1.37	0.67	0.67
December	1.25	1.28	0.64	0.66
Jan.	1.93	1.91	0.84	0.84
February	1.55	1.66	0.74	0.77
Marh	1.43	1.47	0.72	0.70
Apr.	1.42	1.46	0.68	0.71
May	1.17	1.47	0.56	0.68
June	1.16	1.01	0.50	0.50

Diversity Index during the study period July 2014 to June 2016:

Shannon and Weaver Index from July 2014 to June 2015 was observed in the varied from 1.16 to 1.93. The lowest value observed in June and highest value observed in January. July 2015 to June 2016 was observed in the varied from 1.01 to 1.99. The lowest value observed in June and highest value observed in October. This indicates that station has moderate water quality ($H = 1 - 3$).

The value of Simpson index varies between 0 - 1. Zero represents no diversity and 1 represents infinite diversity. This indicates that bigger the value of D , greater is the diversity and smaller the value, smaller the diversity.

The value of Simpson diversity index from July 2014 to June 2015 was observed in the varied from 0.50 to 0.84. The lowest value observed in June and highest value observed in January. July 2015 to June 2016 was observed in the varied from 0.50 to 0.85. The lowest value observed in June and highest range observed in October. Thus, value of D is higher (close to 1) and indicates higher diversity.

Table -48.Fish Species percentage.

Family	Species	No. of Species	Percentage (%)
Bagridae	<i>Mystusbleekari</i>	2	8.695652174
	<i>Mystusseenghala</i>		
Cyprinidae	<i>Labeorohita</i>	9	39.13043478
	<i>Labeocalbasu</i>		
	<i>Labeobata</i>		
	<i>Cirrhinusmrigale</i>		
	<i>Puntius sarana</i>		
	<i>Puntius ticto</i>		
	<i>Cyprinus carpio</i>		
	<i>Rasbora daniconius</i>		
	<i>Catlacatla</i>		
Siluridae	<i>Wallago attu</i>	1	4.347826087
Notopteridae	<i>Notopterusnotopterus</i>	1	4.347826087
Mastacembelidae	<i>Mastacembalusarmatus</i>	2	8.695652174
	<i>Mastacembeluspancalus</i>		
anabantidae	<i>Colisa fasciatus</i>	1	4.347826087
Heteropneustidae	<i>Heteropneustesfossillis</i>	1	4.347826087
Nemacheilidae	<i>Noemacheilusbotia</i>	1	4.347826087
Clariidae	<i>Clariasbatrachus</i>	1	4.347826087
Ambassidae	<i>Chanda nama</i>	1	4.347826087
Ophiocephalidae	<i>Channa punctatus</i>	1	4.347826087
Clupeidae	<i>Hilsa hilsa</i>	1	4.347826087
Nandidae	<i>Nandus nandus</i>	1	4.347826087

Table -49. Seasonal Variations in fish species from July 2014 to June 2015.

S.No.	Name of fish	Jul.-Sep. (Monsoon)	Oct.-Dec. (Post Monsoon)	Jan.-March (Winter)	Apr.-June (Summer)	Jul.-Sep. (Monsoon)	Oct.-Dec. (Post Monsoon)	Jan.-March (Winter)	Apr.-June (Summer)
1	<i>Labeorohita</i>	3.57	0.00	7.78	0.00	8.00	8.75	9.03	4.76
2	<i>Labeocalbasu</i>	0.00	1.22	0.00	7.95	0.00	8.75	3.87	0.00
3	<i>Labeobata</i>	0.00	1.22	0.00	6.82	0.00	7.50	3.23	0.00
4	<i>Notopterusnotopterus</i>	7.14	8.54	8.98	9.09	8.00	2.50	9.03	4.76
5	<i>Catlacatla</i>	3.57	1.22	2.99	1.14	8.00	10.00	1.94	0.00
6	<i>Puntius sarana</i>	3.57	0.00	7.78	7.95	0.00	0.00	4.52	0.00
7	<i>Rasbora daniconius</i>	0.00	1.22	2.99	0.00	0.00	2.50	4.52	0.00
8	<i>Cirrhinusmrigale</i>	3.57	7.32	2.99	0.00	8.00	2.50	8.39	4.76
9	<i>Puntius ticto</i>	0.00	1.22	6.59	9.09	8.00	2.50	7.74	4.76
10	<i>Cyprinus carpio</i>	3.57	2.44	3.59	7.95	8.00	8.75	1.29	14.29
11	<i>Mastacembalusarmatus</i>	7.14	13.41	4.79	14.77	8.00	8.75	4.52	4.76
12	<i>Mastacembeluspancalus</i>	0.00	0.00	0.00	1.14	0.00	1.25	0.00	0.00
13	<i>Mystusseenghala</i>	3.57	8.54	7.78	1.14	4.00	2.50	9.03	4.76
14	<i>Mystusbleekari</i>	7.14	15.85	4.79	15.91	8.00	8.75	4.52	2.38
15	<i>Wallago attu</i>	3.57	2.44	7.78	0.00	8.00	0.00	7.74	19.05
16	<i>Clariasbatrachus</i>	3.57	2.44	3.59	2.27	8.00	1.25	0.65	2.38
17	<i>Heteropneustesfossillis</i>	0.00	1.22	2.99	0.00	0.00	2.50	0.00	4.76
18	<i>Colisa fasciatus</i>	0.00	1.22	0.00	0.00	0.00	1.25	0.00	0.00
19	<i>Chanda nama</i>	3.57	1.22	0.00	0.00	0.00	0.00	0.65	0.00
20	<i>Channa punctatus</i>	28.57	15.85	7.19	2.27	8.00	7.50	3.87	16.67
21	<i>Nandus nandus</i>	0.00	0.00	0.00	0.00	0.00	0.00	0.65	0.00
22	<i>Noemacheilusbotia</i>	7.14	1.22	0.00	1.14	0.00	0.00	0.00	2.38
23	<i>Hilsa hilsa</i>	3.57	2.44	7.19	9.09	0.00	10.00	4.52	4.76

Table -50. Seasonal Variation in fish species from July 2015 to June 2016.

**PHOTOGRAPHS OF FISH SPECIES OF MANSAROVIR TALAB OF
JEERAPURA (DHAR).**

S.No.	Name of fish	Jul.-Sep. (Monsoon)	Oct.-Dec. (Post Monsoon)	Jan.-March (Winter)	Apr.-June (Summer)	Jul.-Sep. (Monsoon)	Oct.-Dec. (Post Monsoon)	Jan.-March (Winter)	Apr.-June (Summer)
1	<i>Labeorohita</i>	5.26	0.00	7.39	2.67	11.11	13.11	10.95	0.00
2	<i>Labeocalbasu</i>	0.00	0.00	2.84	1.33	0.00	1.64	0.73	0.00
3	<i>Labeobata</i>	0.00	0.00	3.41	2.67	0.00	3.28	0.73	0.00
4	<i>Notopterusnotopterus</i>	10.53	6.67	7.39	2.67	5.56	11.48	3.65	2.13
5	<i>Catlacatla</i>	0.00	0.00	3.98	0.00	0.00	0.00	0.00	0.00
6	<i>Puntius sarana</i>	5.26	0.00	2.84	0.00	0.00	0.00	3.65	0.00
7	<i>Rasbora daniconius</i>	5.26	6.67	7.95	0.00	0.00	3.28	3.65	0.00
8	<i>Cirrhinusmrigale</i>	5.26	0.00	6.25	2.67	5.56	0.00	5.11	0.00
9	<i>Puntius ticto</i>	10.53	6.67	7.95	2.67	11.11	11.48	10.22	4.26
10	<i>Cyprinus carpio</i>	10.53	3.33	3.41	9.33	11.11	9.84	4.38	2.13
11	<i>Mastacembalusarmatus</i>	5.26	20.00	1.14	10.67	11.11	9.84	4.38	17.02
12	<i>Mastacembeluspancalus</i>	0.00	0.00	0.00	1.33	5.56	0.00	0.00	0.00
13	<i>Mystusseenghala</i>	10.53	6.67	8.52	10.67	11.11	3.28	5.11	2.13
14	<i>Mystusbleekari</i>	5.26	20.00	1.14	12.00	11.11	13.11	5.84	31.91
15	<i>Wallago attu</i>	10.53	0.00	9.09	0.00	5.56	0.00	8.03	0.00
16	<i>Clariasbatrachus</i>	0.00	0.00	2.84	2.67	0.00	0.00	0.00	0.00
17	<i>Heteropneustesfossillis</i>	0.00	0.00	7.39	9.33	0.00	3.28	10.22	17.02
18	<i>Colisa fasciatus</i>	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00
19	<i>Chanda nama</i>	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00
20	<i>Channa punctatus</i>	0.00	23.33	4.55	18.67	0.00	3.28	10.95	19.15
21	<i>Nandus nandus</i>	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00
22	<i>Noemacheilusbotia</i>	5.26	3.33	3.41	0.00	0.00	9.84	0.73	0.00
23	<i>Hilsa hilsa</i>	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00



Labeo rohita *Labeo calbasu*



Labeo bata *Notopterus notopterus*



Catla catla *Rasbora daniconius*



Puntius sarana *Puntius ticto*



Cirrhinus mrigale



Cyprinus carpio



Wallago attu *Mystus seenghala*



Mystus bleekeri *Noemacheilus botia*



Hilsa hilsa

Table -51. Water Quality Index-Analysed Status Station I & II (July 2014 to June 2016)

	Jul-14		Aug-14		Sep-14		Oct-14		Nov-14		Dec-14	
Stations	S1	S2	S1	S2	S1	S2	S1	S2	S1	S2	S1	S2
NSFQI Value	54.4	54.22	54.91	52.97	49.41	50.45	50.57	52.06	53.19	48.29	54.75	53.41
Status	Medium	Medium	Medium	Medium	Poor	Medium	Medium	Medium	Medium	Poor	Medium	Medium
	Jan-15		Feb-15		Mar-15		Apr-15		May-15		Jun-15	
Stations	S1	S2	S1	S2	S1	S2	S1	S2	S1	S2	S1	S2
NSFQI Value	53.27	52.85	53.27	53.18	51.37	50.91	48.05	50.46	41.15	42.22	40.67	40.55
Status	Medium	Medium	Medium	Medium	Medium	Medium	Poor	Medium	Poor	Poor	Poor	Poor
	Jul-15		Aug-15		Sep-15		Oct-15		Nov-15		Dec-15	
Stations	S1	S2	S1	S2	S1	S2	S1	S2	S1	S2	S1	S2
NSFQI Value	50.73	50.14	51.3	49.59	49.01	47.01	47.81	49.49	51.79	50.62	51.85	49.38
Status	Medium	Medium	Medium	Poor	Poor	Poor	Poor	Poor	Medium	Medium	Medium	Poor
	Jan-16		Feb-16		Mar-16		Apr-16		May-16		Jun-16	
Stations	S1	S2	S1	S2	S1	S2	S1	S2	S1	S2	S1	S2
NSFQI Value	52.54	51.45	52.07	50.89	50.89	49.29	48.12	47.1	43.93	44.31	38.34	41.47
Status	Medium	Medium	Medium	Medium	Medium	Poor	Poor	Poor	Poor	Poor	Poor	Poor

	July 2014 to Dec 2014															
	Jul-14								Aug-14							
Parameter	S1	Q	W	QW	S2	Q	W	QW	S1	Q	W	QW	S2	Q	W	QW
Temperature	26.5	13.5	0.1	1.35	26.4	13.6	0.1	1.36	28.5	11.5	0.1	1.15	29.5	10.5	0.1	1.05
PH	7.7	91	0.11	10.01	7.9	87	0.11	9.57	7.9	87	0.11	9.57	8.1	80	0.11	8.8
BOD	2.9	68	0.11	7.48	2.5	70	0.11	7.7	2	80	0.11	8.8	2.2	76	0.11	8.36
Fecal coliform	142	41	0.16	6.56	165	39	0.16	6.24	178	38	0.16	6.08	192	38	0.16	6.08
Nitrate	29.2	28	0.1	2.8	30.5	27	0.1	2.7	32.8	24	0.1	2.4	34.4	23	0.1	2.3
Phosphate	1.2	36	0.1	3.6	1.48	32	0.1	3.2	1.61	30	0.1	3	1.75	29	0.1	2.9
DO % saturation	83	89	0.17	15.13	85	91	0.17	15.47	85	91	0.17	15.47	84	90	0.17	15.3
Turbidity	38	47	0.08	3.76	35	49	0.08	3.92	30	53	0.08	4.24	36	48	0.08	3.84
Total solids	350	53	0.07	3.71	315	58	0.07	4.06	300	60	0.07	4.2	280	62	0.07	4.34
	NSFWQI			54.4	NSFWQI			54.22	NSFWQI			54.91	NSFWQI			52.97
	Sep-14								Oct-14							
Parameter	S1	Q	W	QW	S2	Q	W	QW	S1	Q	W	QW	S2	Q	W	QW
Temperature	30.5	9.5	0.1	0.95	31	9	0.1	0.9	28.8	11.2	0.1	1.12	29.2	10.8	0.1	1.08
PH	8.2	77	0.11	8.47	8.5	66	0.11	7.26	8.5	66	0.11	7.26	8.2	77	0.11	8.47
BOD	2.5	70	0.11	7.7	2.6	69	0.11	7.59	2.8	68	0.11	7.48	3	67	0.11	7.37
Fecal coliform	215	37	0.16	5.92	222	36	0.16	5.76	242	36	0.16	5.76	235	36	0.16	5.76
Nitrate	36.0	21	0.1	2.1	39.0	18	0.1	1.8	28.0	29	0.1	2.9	26.0	31	0.1	3.1
Phosphate	1.87	28	0.1	2.8	1.4	33	0.1	3.3	2.15	26	0.1	2.6	2.24	26	0.1	2.6
DO % saturation	75	81	0.17	13.77	87	93	0.17	15.81	114	93	0.17	15.81	111	95	0.17	16.15
Turbidity	45	42	0.08	3.36	48	40	0.08	3.2	55	36	0.08	2.88	62	32	0.08	2.56
total solids	278	62	0.07	4.34	225	69	0.07	4.83	235	68	0.07	4.76	210	71	0.07	4.97
	NSFWQI			49.41	NSFWQI			50.45	NSFWQI			50.57	NSFWQI			52.06

	Nov-14								Dec-14							
Parameter	S1	Q	W	QW	S2	Q	W	QW	S1	Q	W	QW	S2	Q	W	QW
Temperature	26.5	13.5	0.1	1.35	25.5	15	0.1	1.5	24.9	16.1	0.1	1.61	25.2	15.3	0.1	1.53
PH	7.6	92	0.11	10.12	7.8	90	0.11	9.9	7.9	87	0.11	9.57	8.1	80	0.11	8.8
BOD	3.2	66	0.11	7.26	3.3	65	0.11	7.15	2.7	69	0.11	7.59	2.4	72	0.11	7.92
Fecal coliform	260	35	0.16	5.6	272	34	0.16	5.44	284	34	0.16	5.44	295	34	0.16	5.44
Nitrate	27.0	30	0.1	3.0	29.0	28	0.1	2.8	35.0	22	0.1	2.2	37.0	20	0.1	2.0
Phosphate	2.38	25	0.1	2.5	2.5	24	0.1	2.4	1.58	30	0.1	3.0	1.25	35	0.1	3.5
DO % saturation	134	82	0.17	13.94	146	50	0.17	8.5	111	95	0.17	16.15	121	89	0.17	15.13
Turbidity	30	53	0.08	4.24	16	66	0.08	5.28	12	72	0.08	5.76	14	69	0.08	5.52
Total solids	188	74	0.07	5.18	170	76	0.07	5.32	380	49	0.07	3.43	365	51	0.07	3.57
	NSFWQI			53.19	NSFWQI			48.29	NSFWQI			54.75	NSFWQI			53.41

	Jan. 2015 to June 2015															
	JAN-2015								FEB-2015							
Parameter	S1	Q	W	QW	S2	Q	W	QW	S1	Q	W	QW	S2	Q	W	QW
Temperature	27	13	0.1	1.3	26.5	13.5	0.1	1.35	29.5	10.5	0.1	1.05	28.9	11.1	0.1	1.11
pH	7.4	93	0.11	10.23	7.6	92	0.11	10.12	7.9	87	0.11	9.57	8.2	77	0.11	8.47
BOD	6.3	49	0.11	5.39	6.7	47	0.11	5.17	5.08	55	0.11	6.05	3	67	0.11	7.37
Fecal coliform	180	38	0.16	6.08	190	38	0.16	6.08	210	37	0.16	5.92	240	36	0.16	5.76
Nitrate	32.0	25	0.1	2.5	35.0	22	0.1	2.2	38.0	19	0.1	1.9	40.4	17	0.1	1.7
Phosphate	1.2	36	0.1	3.6	1.48	32	0.1	3.2	1.61	30	0.1	3.0	1.75	29	0.1	2.9
DO % saturation	119	90	0.17	15.3	114	93	0.17	15.81	98	99	0.17	16.83	102	99	0.17	16.83
Turbidity	20	61	0.08	4.88	22	59	0.08	4.72	28	55	0.08	4.4	25	57	0.08	4.56
Total solids	320	57	0.07	3.99	295	60	0.07	4.2	255	65	0.07	4.55	268	64	0.07	4.48
		NSFWQI		53.27		NSFWQI		52.85		NSFWQI		53.27		NSFWQI		53.18
	Mar-2015								Apr-2015							
Parameter	S1	Q	W	QW	S2	Q	W	QW	S1	Q	W	QW	S2	Q	W	QW
Temperature	31.2	8.8	0.1	0.88	32.5	7.5	0.1	0.75	33.9	6.1	0.1	0.61	24.2	16.8	0.1	1.68
pH	8.4	70	0.11	7.7	8.3	73	0.11	8.03	8.4	70	0.11	7.7	7.9	87	0.11	9.57
BOD	2.8	68	0.11	7.48	3.1	66	0.11	7.26	3.8	62	0.11	6.82	4.3	59	0.11	6.49
Fecal coliform	255	35	0.16	5.6	280	34	0.16	5.44	300	34	0.16	5.44	335	32	0.16	5.12
Nitrate	45.1	14	0.1	1.4	48.0	12	0.1	1.2	45.0	14	0.1	1.4	48.0	12	0.1	1.2
Phosphate	1.87	28	0.1	2.8	1.9	28	0.1	2.8	2.15	26	0.1	2.6	2.24	26	0.1	2.6
DO % saturation	115	93	0.17	15.81	113	94	0.17	15.98	130	84	0.17	14.28	125	87	0.17	14.79
Turbidity	15	67	0.08	5.36	18	63	0.08	5.04	19	62	0.08	4.96	20	61	0.08	4.88

Total solids	280	62	0.07	4.34	270	63	0.07	4.41	290	61	0.07	4.24	302	59	0.07	4.13	
	NSFWQI			51.37		NSFWQI			50.91	NSFWQI			48.05		NSFWQI		50.46

	May-2015								Jun-2015							
Parameter	S1	Q	W	QW	S2	Q	W	QW	S1	Q	W	QW	S2	Q	W	QW
Temperature	35.8	3.2	0.1	0.32	35.9	4.1	0.1	0.41	36.2	4.8	0.1	0.48	36.8	4.2	0.1	0.42
pH	8	84	0.11	9.24	7.8	90	0.11	9.9	8.2	77	0.11	8.47	8.5	66	0.11	7.26
BOD	4.7	57	0.11	6.27	5.1	55	0.11	6.05	5.5	53	0.11	5.83	5.7	52	0.11	5.72
Fecal coliform	370	31	0.16	4.96	330	33	0.16	5.28	365	32	0.16	5.12	380	31	0.16	4.96
Nitrate	47.2	12	0.1	1.2	50.3	10	0.1	1	48.0	12	0.1	1.2	44.0	15	0.1	1.5
Phosphate	2.38	25	0.1	2.5	2.5	24	0.1	2.4	1.58	30	0.1	3	1.15	37	0.1	3.7
DO % saturation	153	50	0.17	8.5	168	50	0.17	8.5	180	50	0.17	8.5	166	50	0.17	8.5
Turbidity	30	53	0.08	4.24	26	56	0.08	4.48	32	51	0.08	4.08	24	58	0.08	4.64
Total solids	325	56	0.07	3.92	298	60	0.07	4.2	320	57	0.07	3.99	335	55	0.07	3.85
		NSFWQI		41.15		NSFWQI		42.22		NSFWQI		40.67		NSFWQI		40.55

	July 2015 to Dec.2015
--	------------------------------

	Jul-2015								Aug.-2015							
Parameter	S1	Q	W	QW	S2	Q	W	QW	S1	Q	W	QW	S2	Q	W	QW
Temperature	27.5	12.5	0.1	1.25	28.5	11.5	0.1	1.15	29.2	10.8	0.1	1.08	29.9	10.1	0.1	1.01
pH	7.9	87	0.11	9.57	7.8	90	0.11	9.9	7.7	91	0.11	10.01	8.3	73	0.11	8.03
BOD	4.6	58	0.11	6.38	4.5	58	0.11	6.38	4.8	57	0.11	6.27	4.2	60	0.11	6.6
Fecal coliform	180	38	0.16	6.08	210	36	0.16	5.76	220	36	0.16	5.76	240	36	0.16	5.76
Nitrate	35.0	22	0.1	2.2	38.0	19	0.1	1.9	42.0	16	0.1	1.6	45.0	14	0.1	1.4
Phosphate	1.5	31	0.1	3.1	1.7	29	0.1	2.9	1.8	29	0.1	2.9	1.95	27	0.1	2.7
DO% saturation	78	85	0.17	14.45	77	84	0.17	14.28	90	95	0.17	16.15	95	98	0.17	16.66
Turbidity	45	42	0.08	3.36	40	45	0.08	3.6	50	39	0.08	3.12	55	36	0.08	2.88
Total solids	280	62	0.07	4.34	285	61	0.07	4.27	270	63	0.07	4.41	255	65	0.07	4.55
	NSFWQI			50.73	NSFWQI			50.14	NSFWQI			51.3	NSFWQI			49.59
	Sep-2015								Oct-2015							
Parameter	S1	Q	W	QW	S2	Q	W	QW	S1	Q	W	QW	S2	Q	W	QW
Temperature	30.8	9.2	0.1	0.92	31.2	8.8	0.1	0.88	29.1	10.9	0.1	1.09	29.5	10.5	0.1	1.05
pH	8.4	70	0.11	7.7	8.54	66	0.11	7.26	8.6	63	0.11	6.93	8.2	77	0.11	8.47
BOD	3.7	63	0.11	6.93	4.2	60	0.11	6.6	4.9	56	0.11	6.16	4.6	58	0.11	6.38
Fecal coliform	260	35	0.16	5.6	280	34	0.16	5.44	275	34	0.16	5.44	295	34	0.16	5.44
Nitrate	48.0	12	0.1	1.2	50.4	10	0.1	1	55.0	8	0.1	0.8	45.0	14	0.1	1.4
Phosphate	2.1	26	0.1	2.6	2.25	26	0.1	2.6	2.35	25	0.1	2.5	2.45	24	0.1	2.4
DO % saturation	88	93	0.17	15.81	81	88	0.17	14.96	98	99	0.17	16.83	103	99	0.17	16.83
Turbidity	46	41	0.08	3.28	44	43	0.08	3.44	55	36	0.08	2.88	65	31	0.08	2.48
Total solids	215	71	0.07	4.97	225	69	0.07	4.83	190	74	0.07	5.18	205	72	0.07	5.04
	NSFWQI			49.01	NSFWQI			47.01	NSFWQI			47.81	NSFWQI			49.49

	Nov-2015								Dec-2015							
Parameter	S1	Q	W	QW	S2	Q	W	QW	S1	Q	W	QW	S2	Q	W	QW
Temperature	26.5	13.5	0.1	1.35	27	13	0.1	1.3	25.5	15	0.1	1.5	26.2	13.8	0.1	1.38
pH	7.9	87	0.11	9.57	8.1	80	0.11	8.8	8.2	77	0.11	8.47	8.4	70	0.11	7.7
BOD	4.5	58	0.11	6.38	4.8	57	0.11	6.27	5	56	0.11	6.16	5.9	51	0.11	5.61
Fecal coliform	320	33	0.16	5.28	340	32	0.16	5.12	385	31	0.16	4.96	398	31	0.16	4.96
Nitrate	38.2	19	0.1	1.9	40.3	17	0.1	1.7	45.0	14	0.1	1.4	45.0	14	0.1	1.4
Phosphate	2.5	24	0.1	2.4	2.65	23	0.1	2.3	1.85	28	0.1	2.8	2	27	0.1	2.7
DO % saturation	109	96	0.17	16.32	115	93	0.17	15.81	100	99	0.17	16.83	117	92	0.17	15.64
Turbidity	38	47	0.08	3.76	25	57	0.08	4.56	18	63	0.08	5.04	20	61	0.08	4.88
Total solids	228	69	0.07	4.83	235	68	0.07	4.76	245	67	0.07	4.69	200	73	0.07	5.11
	NSFWQI			51.79	NSFWQI			50.62	NSFWQI			51.85	NSFWQI			49.38

	Jan.2016 to June 2016															
	Jan-2016								Feb-2016							
Parameter	S1	Q	W	QW	S2	Q	W	QW	S1	Q	W	QW	S2	Q	W	QW
Temperature	28	12	0.1	1.2	27.5	12.5	0.1	1.25	28.5	11.5	0.1	1.15	28	12	0.1	1.2
pH	7.6	92	0.11	10.12	7.8	90	0.11	9.9	8	84	0.11	9.24	8.3	73	0.11	8.03
BOD	6.8	47	0.11	5.17	6.9	46	0.11	5.06	5	56	0.11	6.16	3	67	0.11	7.37
Fecal coliform	170	39	0.16	6.24	199	37	0.16	5.92	220	36	0.16	5.76	250	35	0.16	5.6
Nitrate	40.0	18	0.1	1.8	45	14	0.1	1.4	47.5	12	0.1	1.2	49.1	11	0.1	1.1
Phosphate	1.4	33	0.1	3.3	1.45	32	0.1	3.2	1.55	31	0.1	3.1	1.65	30	0.1	3
DO % saturation	112	95	0.17	16.15	113	94	0.17	15.98	102	99	0.17	16.83	89	94	0.17	15.98
Turbidity	25	57	0.08	4.56	28	55	0.08	4.4	32	51	0.08	4.08	35	49	0.08	3.92
Total solids	295	60	0.07	4.2	278	62	0.07	4.34	260	65	0.07	4.55	240	67	0.07	4.69
		NSFWQI		52.54		NSFWQI		51.45		NSFWQI		52.07		NSFWQI		50.89
	Mar-2016								Apr-2016							
Parameter	S1	Q	W	QW	S2	Q	W	QW	S1	Q	W	QW	S2	Q	W	QW
Temperature	32.2	7.8	0.1	0.78	32.7	7.3	0.1	0.73	34	6	0.1	0.6	34.6	5.4	0.1	0.54
pH	8.6	63	0.11	6.93	8.7	59	0.11	6.49	8.9	52	0.11	5.72	8.8	56	0.11	6.16
BOD	3.5	64	0.11	7.04	3.9	62	0.11	6.82	4.2	60	0.11	6.6	4.8	57	0.11	6.27
Fecal coliform	262	35	0.16	5.6	290	34	0.16	5.44	320	33	0.16	5.28	355	32	0.16	5.12
Nitrate	50.0	10	0.1	1	51.0	10	0.1	1	55.0	8	0.1	0.8	57.0	7	0.1	0.7
Phosphate	1.85	28	0.1	2.8	1.95	27	0.1	2.7	2.205	26	0.1	2.6	2.35	25	0.1	2.5
DO % saturation	105	98	0.17	16.66	107	97	0.17	16.49	120	90	0.17	16.3	114	93	0.17	15.81
Turbidity	18	63	0.08	5.04	22	59	0.08	4.72	18	63	0.08	5.04	19	62	0.08	4.96
Total solids	205	72	0.07	5.04	220	70	0.07	4.9	190	74	0.07	5.18	205	72	0.07	5.04
		NSFWQI		50.89		NSFWQI		49.29		NSFWQI		48.12		NSFWQI		47.1

	May-2016								Jun-2016							
Parameter	S1	Q	W	QW	S2	Q	W	QW	S1	Q	W	QW	S2	Q	W	QW
Temperature	35.9	4.1	0.1	0.41	36.2	3.8	0.1	0.38	37	3	0.1	0.3	37.5	2.5	0.1	0.25
pH	8.8	56	0.11	6.16	8.9	52	0.11	5.72	8.6	63	0.11	6.93	9	49	0.11	5.39
BOD	4.9	56	0.11	6.16	5.5	53	0.11	5.83	5.8	52	0.11	5.72	5.9	51	0.11	5.61
Fecal coliform	370	31	0.16	4.96	390	31	0.16	4.96	345	32	0.16	5.12	360	32	0.16	5.12
Nitrate	59.4	7	0.1	0.7	55.2	8	0.1	0.8	58.0	7	0.1	0.7	54.0	8	0.1	0.8
Phosphate	2.45	24	0.1	2.4	2.51	24	0.1	2.4	2.1	26	0.1	2.6	1.95	27	0.1	2.7
DO % saturation	139	79	0.17	13.43	129	85	0.17	14.45	146	50	0.17	8.5	138	79	0.17	13.43
Turbidity	20	61	0.08	4.88	21	60	0.08	4.8	35	49	0.08	3.92	38	47	0.08	3.76
Total solids	225	69	0.07	4.83	210	71	0.07	4.97	255	65	0.07	4.55	272	63	0.07	4.41
		NSFWQI		43.93		NSFWQI		44.31		NSFWQI		38.34		NSFWQI		41.47

Table -52. Water Quality Index-Analysed Status Station III & IV (July 2014 to June 2016)

	Jul-14		Aug-14		Sep-14		Oct-14		Nov-14		Dec-14	
Stations	S3	S4	S3	S4	S3	S4	S3	S4	S3	S4	S3	S4
NSFQI Value	52.29	52.22	51.88	51.02	47.58	49.08	51.07	50.05	53.42	52.89	52.08	52.4
Status	Medium	Medium	Medium	Medium	Poor	Poor	Medium	Medium	Medium	Medium	Medium	Medium
	Jan-15		Feb-15		Mar-15		Apr-15		May-15		Jun-15	
Stations	S3	S4	S3	S4	S3	S4	S3	S4	S3	S4	S3	S4
NSFQI Value	52.17	51.66	52.08	50.1	50.65	49.85	45.96	45.27	44.55	45.68	37.08	36.92
Status	Medium	Medium	Medium	Medium	Medium	Poor	Poor	Poor	Poor	Poor	Poor	Poor
	Jul-15		Aug-15		Sep-15		Oct-15		Nov-15		Dec-15	
Stations	S3	S4	S3	S4	S3	S4	S3	S4	S3	S4	S3	S4
NSFQI Value	49.18	48.79	48.25	48.02	47.08	44.16	44.29	45.22	48.16	47.53	49.19	48.39
Status	Poor	Poor	Poor	Poor	Poor	Poor	Poor	Poor	Poor	Poor	Poor	Poor
	Jan-16		Feb-16		Mar-16		Apr-16		May-16		Jun-16	
Stations	S3	S4	S3	S4	S3	S4	S3	S4	S3	S4	S3	S4
NSFQI Value	51.24	50.49	49.45	49.53	49.85	46.95	47.03	46.3	44.88	43.42	41.88	41.17
Status	Medium	Medium	Poor	Poor	Poor	Poor	Poor	Poor	Poor	Poor	Poor	Poor

	July 2014 to Dec 2014															
	Jul-14								Aug-14							
Parameter	S3	Q	W	QW	S4	Q	W	QW	S3	Q	W	QW	S4	Q	W	QW
Temperature	25.5	14.5	0.1	1.45	26.9	13.1	0.1	1.31	27.5	12.5	0.1	1.25	28.8	11.2	0.1	1.12
PH	7.8	90	0.11	9.9	8.0	84	0.11	9.24	7.7	91	0.11	10.01	8.2	77	0.11	8.47
BOD	2.2	76	0.11	8.36	2.6	69	0.11	7.59	2.8	68	0.11	7.48	2.6	69	0.11	7.59
Fecal coliform	155	40	0.16	6.4	170	39	0.16	6.24	180	38	0.16	6.08	199	37	0.16	5.92
Nitrate	30.2	27	0.1	2.7	32.5	25	0.1	2.5	33.5	24	0.1	2.4	35.7	21	0.1	2.1
Phosphate	1.95	27	0.1	2.7	2.20	26	0.1	2.6	2.30	25	0.1	2.5	2.45	24	0.1	2.4
DO % saturation	76	82	0.17	13.94	81	88	0.17	14.96	76	82	0.17	13.94	80	87	0.17	14.79
Turbidity	36	48	0.08	3.84	34	50	0.08	4	31	52	0.08	4.16	33	51	0.08	4.08
Total solids	370	50	0.07	3.5	340	54	0.07	3.78	315	58	0.07	4.06	260	65	0.07	4.55
	NSFWQI			52.29	NSFWQI			52.22	NSFWQI			51.88	NSFWQI			51.02
	Sep-14								Oct-14							
Parameter	S3	Q	W	QW	S4	Q	W	QW	S3	Q	W	QW	S4	Q	W	QW
Temperature	30.0	10	0.1	1	30.5	9.5	0.1	0.95	28.2	11.8	0.1	1.18	29.0	11	0.1	1.1
PH	8.1	80	0.11	8.8	8.5	66	0.11	7.26	8.6	63	0.11	6.93	8.4	70	0.11	7.7
BOD	2.4	72	0.11	7.92	2.5	70	0.11	7.7	2.8	68	0.11	7.48	3.8	62	0.11	6.82
Fecal coliform	225	36	0.16	5.76	242	36	0.16	5.76	255	35	0.16	5.6	265	35	0.16	5.6
Nitrate	38.0	19	0.1	1.9	40.0	18	0.1	1.8	32.0	25	0.1	2.5	36.0	21	0.1	2.1
Phosphate	2.25	26	0.1	2.6	2.35	25	0.1	2.5	2.10	26	0.1	2.6	2.35	25	0.1	2.5
DO % saturation	69	73	0.17	11.73	83	89	0.17	15.13	103	99	0.17	16.83	101	99	0.17	16.83
Turbidity	40	45	0.08	3.6	45	42	0.08	3.36	50	39	0.08	3.12	60	33	0.08	2.64
total solids	290	61	0.07	4.27	250	66	0.07	4.62	225	69	0.07	4.83	235	68	0.07	4.76
	NSFWQI			47.58	NSFWQI			49.08	NSFWQI			51.07	NSFWQI			50.05

	Nov-14								Dec-14							
Parameter	S3	Q	W	QW	S4	Q	W	QW	S3	Q	W	QW	S4	Q	W	QW
Temperature	26.9	13.1	0.1	1.31	26.5	13.5	0.1	1.35	25.2	14.8	0.1	1.48	24.8	15.2	0.1	1.52
PH	7.8	90	0.11	9.9	7.9	87	0.11	9.57	8.1	80	0.11	8.8	8.3	73	0.11	8.03
BOD	3.4	65	0.11	7.15	3.5	64	0.11	7.04	3.8	62	0.11	6.82	3.2	66	0.11	7.26
Fecal coliform	280	34	0.16	5.44	295	34	0.16	5.44	280	34	0.16	5.44	325	33	0.16	5.28
Nitrate	32.0	25	0.1	2.5	35.0	22	0.1	2.2	39.0	18	0.1	1.8	36.2	21	0.1	2.1
Phosphate	1.95	27	0.1	2.7	2.15	26	0.1	2.6	2.90	22	0.1	2.2	2.75	23	0.1	2.3
DO % saturation	122	89	0.17	15.13	124	88	0.17	14.96	106	98	0.17	16.66	108	97	0.17	16.49
Turbidity	29	54	0.08	4.32	18	63	0.08	5.04	14	69	0.08	5.52	11	74	0.08	5.92
Total solids	210	71	0.07	4.97	240	67	0.07	4.69	390	48	0.07	3.36	375	50	0.07	3.5
	NSFWQI			53.42	NSFWQI			52.89	NSFWQI			52.08	NSFWQI			52.4

	Jan. 2015 to June 2015															
	JAN-2015								FEB-2015							
Parameter	S3	Q	W	QW	S4	Q	W	QW	S3	Q	W	QW	S4	Q	W	QW
Temperature	27.5	12.5	0.1	1.25	26..9	13.1	0.1	1.31	30.2	9.8	0.1	0.98	29.5	10.5	0.1	1.05
pH	7.7	91	0.11	10.01	7.8	90	0.11	9.9	7.8	90	0.11	9.9	8.4	70	0.11	7.7
BOD	5.9	51	0.11	5.61	6.0	51	0.11	5.61	5.2	55	0.11	6.05	3.8	62	0.11	6.82
Fecal coliform	288	34	0.16	5.44	295	34	0.16	5.44	245	35	0.16	5.6	270	35	0.16	5.6
Nitrate	34.0	23	0.1	2.3	38.0	19	0.1	1.9	41.0	17	0.1	1.7	43.0	15	0.1	1.5
Phosphate	2.30	25	0.1	2.5	2.40	25	0.1	2.5	2.69	23	0.1	2.3	2.85	22	0.1	2.2
DO % saturation	108	97	0.17	16.49	103	99	0.17	16.83	96	99	0.17	16.83	92	97	0.17	16.49
Turbidity	23	59	0.08	4.72	20	61	0.08	4.88	30	53	0.08	4.24	28	55	0.08	4.4
Total solids	333	55	0.07	3.85	395	47	0.08	3.29	266	64	0.07	4.48	280	62	0.07	4.34
		NSFWQI		52.17		NSFWQI		51.66		NSFWQI		52.08		NSFWQI		50.1
	Mar-2015								Apr-2015							
Parameter	S3	Q	W	QW	S4	Q	W	QW	S3	Q	W	QW	S4	Q	W	QW
Temperature	30.8	9.2	0.1	0.92	31.9	8.1	0.1	0.81	33.5	6.5	0.1	0.65	35.0	5	0.1	0.5
pH	8.6	63	0.11	6.93	8.5	66	0.11	7.26	8.8	56	0.11	6.16	8.9	52	0.11	5.72
BOD	2.2	76	0.11	8.36	3.2	66	0.11	7.26	4.8	57	0.11	6.27	4.0	61	0.11	6.71
Fecal coliform	295	34	0.16	5.44	310	33	0.16	5.28	330	33	0.16	5.28	355	32	0.16	5.12
Nitrate	48.2	11	0.1	1.1	50.0	10	0.1	1	47.0	12	0.1	1.2	52.3	9	0.1	0.9
Phosphate	2.99	21	0.1	2.1	3.10	21	0.1	2.1	3.55	19	0.1	1.9	3.20	20	0.1	2
DO % saturation	107	97	0.17	16.49	112	95	0.17	16.15	123	88	0.17	14.96	119	90	0.17	15.3
Turbidity	18	63	0.08	5.04	20	61	0.08	4.88	17	65	0.08	5.2	19	62	0.08	4.96
Total solids	290	61	0.07	4.27	300	73	0.07	5.11	280	62	0.07	4.34	312	58	0.07	4.06
	NSFWQI			50.65		NSFWQI		49.85	NSFWQI			45.96		NSFWQI		45.27

	May-2015								Jun-2015							
Parameter	S3	Q	W	QW	S4	Q	W	QW	S3	Q	W	QW	S4	Q	W	QW
Temperature	35.2	4.8	0.1	0.48	36.2	3.8	0.1	0.38	36.8	3.8	0.1	0.38	37.1	2.9	0.1	0.29
pH	8.3	73	0.11	8.03	8.2	77	0.11	8.47	8.6	63	0.11	6.93	8.9	52	0.11	5.72
BOD	3.9	62	0.11	6.82	3.0	67	0.11	7.37	4.5	58	0.11	6.38	4.1	60	0.11	6.6
Fecal coliform	390	31	0.16	4.96	410	31	0.16	4.96	385	31	0.16	4.96	350	32	0.16	5.12
Nitrate	50.0	10	0.1	1	54.0	8	0.1	0.8	51.3	9	0.1	0.9	49.4	10	0.1	1
Phosphate	3.55	19	0.1	1.9	3.75	18	0.1	1.8	3.80	18	0.1	1.8	2.95	22	0.1	2.2
DO % saturation	139	79	0.17	13.43	133	82	0.17	13.94	151	50	0.17	8.5	146	50	0.17	8.5
Turbidity	32	51	0.08	4.08	29	54	0.08	4.32	33	51	0.08	4.08	35	49	0.08	3.92
Total solids	335	55	0.07	3.85	358	52	0.07	3.64	340	54	0.07	3.87	365	51	0.07	3.57
		NSFWQI		44.55		NSFWQI		45.68		NSFWQI		37.08		NSFWQI		36.92

	July 2015 to Dec.2015															
	Jul-2015								Aug.-2015							
Parameter	S3	Q	W	QW	S4	Q	W	QW	S3	Q	W	QW	S4	Q	W	QW
Temperature	28.2	11.8	0.1	1.18	29.0	11	0.1	1.1	27.9	12.1	0.1	1.21	28.5	11.5	0.1	1.15
pH	8.0	84	0.11	9.24	7.9	87	0.11	9.57	7.8	90	0.11	9.9	8.4	70	0.11	7.7
BOD	4.8	57	0.11	6.27	4.1	60	0.11	6.6	4.5	58	0.11	6.38	4.0	61	0.11	6.71
Fecal coliform	220	36	0.16	5.76	250	35	0.16	5.6	235	36	0.16	5.76	266	35	0.16	5.6
Nitrate	38.0	19	0.1	1.9	41.2	17	0.1	1.7	44.2	14	0.1	1.4	46.2	13	0.1	1.3
Phosphate	1.60	30	0.1	3	1.90	28	0.1	2.8	2.20	26	0.1	2.6	2.50	24	0.1	2.4
DO% saturation	77	84	0.17	14.28	75	81	0.17	13.77	75	81	0.17	13.77	89	94	0.17	15.98
Turbidity	47	41	0.08	3.28	42	44	0.08	3.52	53	37	0.08	2.96	55	36	0.08	2.88
Total solids	290	61	0.07	4.27	305	59	0.07	4.13	290	61	0.07	4.27	268	64	0.07	4.48
	NSFWQI			49.18	NSFWQI			48.79	NSFWQI			48.25	NSFWQI			48.02
	Sep-2015								Oct-2015							
Parameter	S3	Q	W	QW	S4	Q	W	QW	S3	Q	W	QW	S4	Q	W	QW
Temperature	29.5	10.5	0.1	1.05	30.5	9.5	0.1	0.95	30.0	10	0.1	1	28.5	11.5	0.1	1.15
pH	8.6	63	0.11	6.93	8.8	56	0.11	6.16	8.9	52	0.11	5.72	8.7	59	0.11	6.49
BOD	3.9	62	0.11	6.82	4.6	58	0.11	6.38	5.2	55	0.11	6.05	5.6	53	0.11	5.83
Fecal coliform	275	34	0.16	5.44	297	34	0.16	5.44	315	33	0.16	5.28	345	32	0.16	5.12
Nitrate	49.0	11	0.1	1.1	53.3	8	0.1	0.8	58.0	7	0.1	0.7	47.0	12	0.1	1.2
Phosphate	2.80	22	0.1	2.2	2.45	24	0.1	2.4	2.55	24	0.1	2.4	2.68	23	0.1	2.3
DO % saturation	92	97	0.17	16.49	77	84	0.17	14.28	82	89	0.17	15.13	88	93	0.17	15.81
Turbidity	48	40	0.08	3.2	49	40	0.08	3.2	52	38	0.08	3.04	68	30	0.08	2.56
Total solids	335	55	0.07	3.85	255	65	0.07	4.55	210	71	0.07	4.97	235	68	0.07	4.76
	NSFWQI			47.08	NSFWQI			44.16	NSFWQI			44.29	NSFWQI			45.22

	Nov-2015								Dec-2015							
Parameter	S3	Q	W	QW	S4	Q	W	QW	S3	Q	W	QW	S4	Q	W	QW
Temperature	27.5	12.5	0.1	1.25	28.3	11.7	0.1	1.17	26.5	13.5	0.1	1.35	27.3	12.7	0.1	1.27
pH	8.5	66	0.11	7.26	8.7	59	0.11	6.49	8.5	66	0.11	7.26	8.6	63	0.11	6.93
BOD	5.5	53	0.11	5.83	5.8	52	0.11	5.72	5.6	53	0.11	5.83	5.9	51	0.11	5.61
Fecal coliform	362	32	0.16	5.12	381	31	0.16	4.96	401	31	0.16	4.96	418	30	0.16	4.8
Nitrate	42.3	16	0.1	1.6	45.2	14	0.1	1.4	48.0	12	0.1	1.2	50.0	10	0.1	1
Phosphate	2.72	23	0.1	2.3	2.95	21	0.1	2.1	2.55	24	0.1	2.4	2.60	24	0.1	2.4
DO % saturation	95	98	0.17	16.66	103	99	0.17	16.83	98	99	0.17	16.83	103	99	0.17	16.83
Turbidity	42	44	0.08	3.52	30	53	0.08	4.24	20	61	0.08	4.88	23	59	0.08	4.72
Total solids	252	66	0.07	4.62	247	66	0.07	4.62	265	64	0.07	4.48	230	69	0.07	4.83
	NSFWQI			48.16	NSFWQI			47.53	NSFWQI			49.19	NSFWQI			48.39

	Jan.2016 to June 2016															
	Jan-2016								Feb-2016							
Parameter	S3	Q	W	QW	S4	Q	W	QW	S3	Q	W	QW	S4	Q	W	QW
Temperature	28.8	11.2	0.1	1.12	28.4	11.6	0.1	1.16	29.5	10.5	0.1	1.05	29.0	11	0.1	1.1
pH	7.9	87	0.11	9.57	8.0	84	0.11	9.24	8.2	77	0.11	8.47	8.4	70	0.11	7.7
BOD	6.2	50	0.11	5.5	6.0	51	0.11	5.61	5.8	52	0.11	5.72	3.5	64	0.11	7.04
Fecal coliform	225	36	0.16	5.76	266	35	0.16	5.6	290	34	0.16	5.44	320	33	0.16	5.28
Nitrate	44.0	15	0.1	1.5	49.0	11	0.1	1.1	48.0	12	0.1	1.2	51.1	9	0.1	0.9
Phosphate	1.90	28	0.1	2.8	2.15	26	0.1	2.6	2.55	24	0.1	2.4	2.65	23	0.1	2.3
DO % saturation	109	96	0.17	16.32	105	98	0.17	16.66	97	99	0.17	16.83	99	99	0.17	16.83
Turbidity	27	55	0.08	4.4	29	54	0.08	4.32	34	50	0.08	4	37	47	0.08	3.76
Total solids	285	61	0.07	4.27	298	60	0.07	4.2	277	62	0.07	4.34	250	66	0.07	4.62
		NSFWQI		51.24		NSFWQI		50.49		NSFWQI		49.45		NSFWQI		49.53
	Mar-2016								Apr-2016							
Parameter	S3	Q	W	QW	S4	Q	W	QW	S3	Q	W	QW	S4	Q	W	QW
Temperature	31.9	8.1	0.1	0.81	32.5	7.5	0.1	0.75	35.0	5	0.1	0.5	35.5	4.5	0.1	0.45
pH	8.8	56	0.11	6.16	8.9	52	0.11	5.72	8.7	59	0.11	6.49	8.6	63	0.11	6.93
BOD	2.9	68	0.11	7.48	4.5	58	0.11	6.38	4.9	56	0.11	6.16	5.2	55	0.11	6.05
Fecal coliform	355	32	0.16	5.12	390	31	0.16	4.96	380	31	0.16	4.96	385	31	0.16	4.96
Nitrate	53.4	8	0.1	0.8	55.0	8	0.1	0.8	58.0	7	0.1	0.7	59.0	9	0.1	0.9
Phosphate	2.85	22	0.1	2.2	2.95	21	0.1	2.1	3.05	21	0.1	2.1	3.25	20	0.1	2
DO % saturation	107	97	0.17	16.49	103	99	0.17	16.83	115	93	0.17	15.81	120	90	0.17	15.3
Turbidity	19	62	0.08	4.96	23	59	0.08	4.72	17	65	0.08	5.2	20	61	0.08	4.88
Total solids	225	69	0.07	4.83	240	67	0.07	4.69	200	73	0.07	5.11	225	69	0.07	4.83
		NSFWQI		48.85		NSFWQI		46.95		NSFWQI		47.03		NSFWQI		46.3

	May-2016								Jun-2016							
Parameter	S3	Q	W	QW	S4	Q	W	QW	S3	Q	W	QW	S4	Q	W	QW
Temperature	36.0	4	0.1	0.4	36.5	3.5	0.1	0.35	37.2	2.8	0.1	0.28	37.9	2.1	0.1	0.21
pH	8.4	70	0.11	7.7	9.0	49	0.11	5.39	8.9	52	0.11	5.72	9.1	46	0.11	5.06
BOD	5.8	52	0.11	5.72	5.0	56	0.11	6.16	4.8	57	0.11	6.27	4.2	60	0.11	6.6
Fecal coliform	395	31	0.16	4.96	420	30	0.16	4.8	455	30	0.16	4.8	480	29	0.16	4.64
Nitrate	62.1	6	0.1	0.6	65.1	6	0.1	0.6	68.0	5	0.1	0.5	60.0	7	0.1	0.7
Phosphate	3.50	19	0.1	1.9	3.68	18	0.1	1.8	3.10	21	0.1	2.1	3.55	19	0.1	1.9
DO % saturation	132	83	0.17	14.11	122	89	0.17	15.13	132	83	0.17	14.11	131	83	0.17	14.11
Turbidity	21	60	0.08	4.8	24	58	0.08	4.64	38	47	0.08	3.76	39	46	0.08	3.68
Total solids	245	67	0.07	4.69	250	65	0.07	4.55	278	62	0.07	4.34	288	61	0.07	4.27
		NSFWQI		44.88		NSFWQI		43.42		NSFWQI		41.88		NSFWQI		41.17

DISCUSSION

DISCUSSION

The current study was conducted in 02 years from July 2014 to June 2016. Four sampling sites were selected at the **Mansarovar Talab of Jeerapura** for the present research work.

Physico-chemical parameters:

1. Temperature:

The Temperature is one of the most important factors in an aquatic environment and profoundly influences the nature of water body. In general, water temperature varied from 24.8 to 37.9°C. The lowest water temperature was observed at sampling site 04 in December in the year 2014-2015 and highest range was observed at sampling site 04 in June in the year 2015-2016. *Sharma et al., (2013)* observed temperature varied from 25°C to 42°C in Kunda river, Khargone (M.P.). *Sharma et al., (2008)* observed temperature fluctuation between 20.6°C to 38.4°C while studying the hydrological parameters of Narmada river at Hoshangabad. The result of current study is assisted by the research work of *Sharma et al., (2008)*.

2. Transparency:

Transparency of the water body is mainly affected by the suspended molecule and other component like plankton growth, nature of water, rainfall and temperature conditions. In general, transparency varied from 19.8 to 50.0 cm. The lowest transparency was observed at sampling site 02 in July in the year of 2015-2016. This was due to the increased number of particles which were added by surface runoff. The highest transparency was at sampling site 04 in November in the year 2015-2016. This was due to settling

of particles. *Yadav and Shrivastava (2011)* observed the water transparency varied from 25 to 110 cm in the river Ganga at Ghazipur. *Prabhakar et al., (2012)* observed Transparency range from 7.30 cm to 20.89 cm. In Palar River, Vellore district Tamilnadu. *Zahoor et al., (2012)* recorded a similar range of transparency the physico-chemical parameters of Narmada river. *Chaurasia (2013)* recorded same range of water transparency in Kunda river (M.P).

3. Turbidity:

Turbidity is the measure of relative clarity of a liquid. In general, turbidity ranged from 11 to 68. The lowest turbidity was observed at sampling site 04 in December in the year 2014-2015 and highest value was observed at sampling site 04 in October in the year 2015-2016.

4. pH:

pH is very affected by photosynthetic activity of aquatic plants, disposal of sewage and disposal of industrial water and exposure of air temperature, etc. pH ranged from 7.4 to 9.1. The lowest pH was observed at sampling site 01 in January in the year 2014-15. The highest pH was observed at sampling site 04 in June in 2015-16. *Chaudhari et al., (2001)* observed pH ranged from 7.6 to 9.8 from Chattri lake. *Gupta et al., (2001)* observed pH varied from 7.8 to 9.3 in Udaipur lakes. *Shitika Barkale (2013-2015)* found the pH varied from 7.4 to 9.4 in Bilawali talab, Indore. This study is supported by *Shitika Barkale (2013-2015)*.

5. Dissolved Oxygen:

Dissolved Oxygen is important for aquatic system and also essential for the metabolism in the organisms. Air and photosynthetic activity There are two main sources of dissolved oxygen in water. In general, dissolved

oxygen oscillated between 5.2 to 12.3 mg/l. The lowest range was observed at sampling site 03 in September in the year of 2014-2015. The highest dissolved oxygen value was recorded at sampling site 01 in June in the year of 2014-2015. *Sharma & Chowdhary (2011)* recorded DO oscillated between 0.4 to 8 mg/l in river Tawi of Jammu and Kashmir. *Sisodiya and Moundiotiya (2006)* recorded dissolved oxygen range from 5.2 to 12.08 mg/l in Kalakho Lake, Rajasthan and same range was also observed *Sharam et al., (2011)*. The result of current study is supported by *Sisodiya and Moundiotiya (2006)*.

6. Total Solid:

The Total Solids are the sum of dissolved, suspended and settleable solids in water. It is also used to determine a sludge dry weight. The Total Solids fluctuated between 170 to 390 mg/l. The lowest Total Solids was observed at sampling site 02 in November in the year 2014-2015 and highest value was observed at sampling site 03 in December in the year 2014-2015. *Bhutiani et al (2014)* reported total solids varied from 112.00 mg/l lowest value and 650.60 mg/l highest value in Ganga river. *Tripathi et al (2014)* reported total solids value oscillated between 325.6 to 411.3 mg/l in river Ganga at Allahabad. *Patil and Sharma (2016)* recorded total solid varied from 216 to 617 mg/l in river Narmada. With the results of this study similar results were recorded by *Tripathi et al (2014)* and *Patil and Sharma (2016)*.

7. Free CO₂:

The quantity of free CO₂ depends on the quantity of water or the quantity of gas in the atmosphere. The CO₂ content fluctuated from 1.2 to

8.5 mg/l. in the year 2014-2016. The lowest Free CO₂ was recorded at samplingsite01 in April in the year 2014-2015 and highest value was observed at samplingsite04 in November in the year 2015-2016. This might be due to increased and decreased biological activities of aquatic of fauna & flora. *Singh et al. (1997)* and *Lakra (1990)* recorded low value of Free CO₂ during winter months. *Singh et al. (1987)* also observed highest and lowest of CO₂ in different months. *Mandal and Haking (1977)* observed the highest & lowest CO₂ concentration in Febuaryand April months.

8. Total Alkalinity:

Total alkalinity is varied from 225.4 to 499.0 mg/l. The lowestrage was observed at samplingsite02 in March in the year of 2014-2015. The highestrange was observed at samplingsite02 in November in the year of 2015-2016. During present highest total alkalinity was observed in winter and lowest in summer, was mostly because of bicarbonate as the ranged for carbonate alkalinity were observed to be much lower. In the present study, on the basis of bicarbonate concentration, various seasons may be arranged in the order of summer, decreasingly followed by rainy and winter seasons. The observations of Ganapati (1960), that increase of bicarbonate alkalinity is attributed to the lowering of oxygen concentration, was supported by *Adoni (1975)*, *Sankhla (1981)*. *Selot (1977)*, and *Dhakar (1979)*, however, did not observe this relationship. In the present study a relationship of this kind was clearly observed. *Selot (1977)*, *Jain (1978)* and *Dhakar (1979)* have observed a rise of bicarbonate values with a rise in temperature, reaching their maximum in June. In the present study also a similar trend of this kind was observed.

9. Biochemical Oxygen Demand:

Biochemical oxygendemand involving of oxygen consumed by the micro-organism in stabilizing the biologically decomposable organic matter under aerobic conditions. The biochemical oxygen demand rangedfom 2.0 to 6.9 mg/l. The lowestBOD value was observed at samplingsite01 in August in the year 2014-2015. In the highestrange was recorded at samplingsite02 in January in the year of 2015-2016. **Pandey (2007)**observed BOD ranged from 2 to 5.7 mg/l in Yeshwantsagar reservoir, Indore. **Sharma et al., (2013)**observed BOD ocillated between 3.1 to 5.63mg/l in Kunda river, Khargone (M.P.). In this research work BOD value is higher than the result of **Sharma et al., (2013)**.

10. Hardness:

The hardness is governed by contents of magnesium salt, and calcium, largely combine with carbonates and bicarbonates, and with sulphate, chloride and other elements. The total hardness ocillatedfrom 105.0 to 330.0 mg/l. The lowest total hardness was observed at samplingsite02 in February in the year of 2014-2015 and highest total hardness at samplingsite04 in June in the year of 2014-2015. This highest value is due to continuous leaching an accumulation of salts in the absence of flow in summer months. **Murhekar (2011)** recoreded the total hardnesseranged from 312 to 687 mg/l in and around Akot city. **Bhawna Dawar (2013-14)**got the range of total hardness from 220 mg/l to 440 mg/l. at Shahid Chandra shekharazad dam, Jobat. The result of present study is lower than **Bhawna Dawar (2013-14)** &**Murhekar (2011)**.

11.Magnesium:

In the present study Magnesium varied from 18.3 to 44.0 mg/l. The lowest Magnesium was observed at samplingsite02 in February in the year 2014-2015 and highest range was recorded at samplingsite04 in November in the year 2015-2016. *Shitika Barkale (2014-15)* magnesium value ranges from 11 to 48mg/L. *Mariappan et al., (2002)* the Magnesium value varied from 9 to 31 mg/l. in Sivakasi. *Jain & Shrivastava (1998)* the magnesium value oscillated between 12 mg/l. to 40.6 mg/l. in Bharar River, district Chhatarpur. *Jain et al., (2001)* observed the magnesium value 3.9 mg/l to 292 mg/l. in Karnataka. *Shoukat Ara et al., (2000-2001)* found the magnesium varied from 4.7 to 11 mg/l. *Garg (2003)* observed the Magnesium oscillated between 5 to 25 mg/l. in Chitrakut. *Baruah et al., (2003)* observed the magnesium fluctuated 10 to 30 mg/l. in Subansiri River, Assam. The present study was supported by the research work of *Shitika Barkale (2014-15)*.

12. Calcium:

Calcium varied from 35.5 to 74.2 mg/l. The lowest Calcium was observed at samplingsite01 in August in the year 2014-2015 and highest range was observed at samplingsite04 in May in the year 2014-2015. It being an important supporter to hardness in water and it reduce the useful of water for domestic use. *Shitika Barkale (2014-15)* recorded the value of calcium 17.2 to 58 Mg/l. at Bilawali talab, Indore. *Jadhav et al., (2006)* observed calcium varied from 52 to 73 mg/l in Krishna river at Kharad, Maharashtra. *Murhekar (2011)* recorded the calcium ranged from 147 to 472 mg/l in and around Akot city. *Bajpai (2012)* observed calcium oscillated between 49 to 50 mg/l in Hasdeo river and 112.8 to 113.2 mg/l in Arpa river, Bilaspur region. *Hamaidi et al., (2013)* calcium fluctuated

between 90 to 104 mg/l in Chiffa river, Algeria. This study supported by the research work of *Jadhav et al., (2006)*.

13.Chloride:

Chloride is present in fresh water in the form of calcium, magnesium and sodium salts. The concentration of chloride content is also used as an indicator of pollution in fresh water. In general, chloride varied from 23.4 to 69.3 mg/l. The lowestrage was observed at samplingsite01 in December in the year 2014-2015 and highestrange was at samplingsite04 in June in the year of 2014-2015. *Sharma & Chowdhary (2011)* observed chloride ranged from 21.95 to 59.88 mg/l in river Tawi Jammu and Kashmir. *Sharma et al., (2013)* observed Chloride varied from 0.3 mg/l to 53.4 mg/l in Kunda river, Khargone (M.P.). *Sharma & Chowdhary (2011)* reported low chloride range than the present work.

14.Nitrate:

Nitrogen is present in the form of organic nitrogen compounds, nitrate, ammonia, as nitrite. In general, nitrate varied from 26.0 to 68.0 mg/l. The lowestrage was observed at samplingsite02 in month of October in the year of 2014-2015 and highestrange was observed at samplingsite03 in June in the year of 2015-2016. *Manjare et al., (2010)* recorded nitrate ranged from 4.4 to 36.8 mg/l in Kolhapur district Maharashtra. *Bhawna dawar (2013)* recorded nitrate value varied from 0.05 to 55.0 mg/l. in Shahid Chandra shekharazad dam, jobat. This result is slightly higher than the result of *Bhawna dawar (2013)*.

15.Phosphate:

The main sources of phosphorus are domestic sewage detergents, industrial waste water and agricultural effluents with fertilizers. The study was phosphate value varied from 1.15 to 3.80 mg/l. The lowestrage was observed at samplingsite02 in June in the year of 2014-2015. The highestrange was at samplingsite03 in June in the year of 2014-2015. *Sharma et al., (2008)* while studying on Narmada river observed phosphate ranged from 6.2 to 21 mg/l. Phosphate value variedfrom 0.05 to 3.80 mg/l. *Bhawna dawar* at shahid Chandra shekharazad dam, jobat in **2015**. The similarvalue of *Sharam et al., (2014)*, *Sharma et al., (2013)*, *Chouhan (2013)* and *Choursiya (2013)*.

16.Sulphate:

Sulphates are generally pesent in appreciable concentration and impart hardness to water. Mostly they are present in amounts more then adequate for fresh water productivity. It was fluctuated between 4.9 to 9.9 mg/l. The lowestrage was observed at samplingsite 02 in February in the year of 2014-2015. The highest value was at samplingsite 04 in September in the year of 2015-2016. *Shitika Barkale (2014-15)* recorded the value of sulphate varied from 4.8-115 mg/l. at bilawali talab, Indore. *Sharma et al., (2011)* recorded sulphate varied from 325 to 449 mg/l and 415 to 493 mg/l in pre-monsoon and monsoon seasons. *Krishna (2012)* recorded sulphate varied from 38.13 to 68.65 mg/l in Kaveri river, Kudige, Kodagu, Karnataka. This current research work supported by *Shitika Barkale (2014-15)*.

17.Total Coli Form:

The total coliform is found in the intestines of warm-blooded animal and therefor are present in sewage, on and in soil, surface waters and

vegetation. The total coliform group has been used for some time as an “indicator organism”. An indicator organism by itself is considered to cause no diseases in man or animal, but its presence usually indicates the presence of pathogenic or disease-causing organism. It was fluctuated between 300 to 660 MNP/100 ml. The lowest range was observed at samplingsite01 in July in the year 2014-2015. The highest range was observed at samplingsite04 in December in the year of 2015-2016. *Hrushikesh Behra et al. (2000)* recorded value of Total coliform from 1800 to 97000 MPN/100 ml. *Bhawna dawar (2014-15)* recorded total coliform from 200 to 460 ml. in Shahid Chandra shekhar azad dam, jobat. Present work is supported by *Bhawna Dawar (2014-15)*.

18. Faecal Coli form:

Faecal coliform bacteria are normally present in the feces of humans and animals. Faecal coliform bacteria enter the rivers directly or indirectly from the sewage system and agricultural runoffs. Disease causing organisms including bacteria, viruses, and other parasites can be a part of faecal coliform. Faecal coliform varied from 145 to 480 MNP/100 ml. The lowest Faecal coliform observed at samplingsite01 in July in the year 2014-2015. The highest Faecal coliform observed at samplingsite04 in June in the year of 2015-2016. Same results with the current study of Faecal coliform was observed by *Basu et al (2012)*, *Pratibha and Murulidhar (2015)*, *Nicholson et al (2017)*.

19. Biological Analysis:

The bottom fauna of this pond qualitatively diversified and rich. All together 32 species of bottom fauna were collected and identified. They

showed remarkable fluctuation both its quantity and quality in different season. The densities of oligochaetes and insects formed the main bulk consisting about 80% of the total collected benthos. The percentage abundance of oligochaetes was maximum in June and July while minimum in colder months. Insects, gastropods and ostracods showed their highest abundance during colder months and lowest during monsoon. However, variation in other groups were also registered but in haphazard fashion.

During biostatistical analysis of collected data, water temperature was correlated with DO but positive correlation of water temperature and phosphate as well as free CO₂. DO was found inverse relationship with Free CO₂. Inverse relationship such as Free CO₂ & pH, pH and phosphate but positive correlation with Free CO₂ and phosphate were exhibited.

Collected fishes of this pond were represented by 7 orders, 13 families, 17 genera and 23 species. Fishes were of all sizes but there were no any artificial manuring and introduction of fry, fingerlings and fishes could be recognized as the modest one.

Macro-invertebrates- During the study period, Oligochaetes, Hirudinea (Leeches), Gastropods, palacypods, Insects, and Crustaceans were recorded.

20.Diversity Indices:

In Mansarovar Talab the physico-chemical characteristics develop many limnological habitats, with varying community structure and species composition. The higher the range of index, the greater the diversity and obviously the cleaner the environment. *Willhm and Dorris (1966)* have

distinguished three zones based on the status of water given with index values, which was found applicable to River Narmada.

The Shannon Weaver Species diversity Index (H) can be used for the evaluation of the stresses to the benthic macro invertebrate population due to changes in water quality. This will serve as a direct indication of the cumulative effect of environmental stress on benthic organisms.

In the current study, Shannon and Weaver Index from July 2014 to June 2015 was observed in the varied from 1.16 to 1.93. July 2015 to June 2016 was observed in the varied from 1.01 to 1.99. **Shukla and Shrivastava (2004)** recorded value of H index in the range from 1.73 to 2.94 in Gandhisagar reservoir India. **Nandan (2003)** reported that diversity index for different species was generally low in the retting zones (H= 0.68 to 1.20) when compared to the non retting zone (H= 0.88 to 2.97) of Kadinamkulam backwaters. The lowest range of H was observed in the month of July and August, **Bass and Potts (2001)** reported the same decrease in diversity index during July and they attributed it to the emergence of several groups of aquatic insects prior to sampling. **Bhat et al. (2011)** recorded the value of H index in the range between H= 1.99 to 2.42 in Kashmir Himalayas. **Sharma and Chowdhary (2011)** observed H value between H= 0.20 to 1.98 in River Tawi. **Sharma et al (2010)** observed H index between H= 0.79 to 1.62 in river Chenab.

In the present study value of Simpson diversity index from July 2014 to June 2015 was observed in the varied from 0.50 to 0.84. July 2015 to June 2016 was observed in the varied from 0.50 to 0.85. **Shukla and Shrivastava (2004)** recorded Simpson's index value between D= 0.74 to 0.86 at

Gandhisagar reservoir MP. Similar results were also reported by *Nkwoji (2010)* giving the community diversity and species richness of macro invertebrates. *Arab et al (2004)* also reported high species richness in the upstream part of Chelif course, which decreases at the downstream site, where the water was heavily polluted. According to *Duran and Suicmez (2007)*, the Cekerez stream of Turkey was characterised as class-I water quality with high species richness. *Sharma et al (2010)* recorded the value of Simpson's index between $D = 0.21$ to 0.54 in Chenab river. *Sharma and Chowdhary (2011)* recorded the value of Simpson's index between $D = 0.31$ to 0.91 in river Tawi, India.

21. Fish Diversity:

Fish production in talab, lake & reservoir is indirectly or directly dependent on the abundance of plankton and bottom fauna. The physico-chemical character of water determines the quantity and quality of the fauna. A total of twenty-three species of fish belonging to seventeen genera, under thirteen families and seven orders were recorded from Mansarovar talab. The most dominant family was cyprinidae 39.13% followed by Bargridae and Mastacembelidae 8.70% each. This is because these three groups are dominant in lentic water reservoirs of India & Bangladesh and are more tolerant towards pollution (*Pathak, S. K. & Mudgal L.K., 2005*). The family cyprinidae represents the bulk of fish species, which includes nine species. The important fish species recorded from this family are *Rasbora daniconius*, *Puntius sarana*, *Puntius ticto*, Family Bargridae represents second dominant family 8.70% comprised of two species namely *Mystus bleekeri* and *Mystus seenghala*. The family Siluridae represented by *Wallago attu* whereas Heteropneustidae represented by *Heteropneustes fossilis*.

Family Clariidae is represented by a single species *Clarias batrachus*, family Belontiidae also recorded by single species *Colisa fasciatus*. The species recorded is *Chanda nama* and Channidae *Channa punctatus*, Family Notopteridae is also recorded by single species *Notopterus notopterus*. The Mastacembelidae family recorded two species *Mastacembelus armatus* & *Mastacembelus pancalus*. Family Clupeidae is also recorded by single species *Hilsa hilsa* and family Nemacheilidae represented by *Noemacheilus botia*. The Change in the composition of a fish assemblage often indicates a variation in the water quality parameters. Such as Temperature, pH, D.O. and Nutrient (**Jhingran, 1982**). Due to more fecundity of major carp and suitable environmental condition there exists a relatively higher number of cypriniformes. Such type of observation was reported by **Talwar & Jhingran (1991)** and **Das & Chand (2003)** **Pathak & Mudgal (2005)** Indian fishes.

National Sanitation Foundation Water Quality Index (NSFWQI): -

It is a commonly-used water quality index (WQI) was developed by the National Sanitation Foundation (NSF) in 1970 (**Brown and others, 1970**). The NSF WQI was developed to provide a standardized method for comparing the water quality of various bodies of water. In the current study the same was used to check the status of water as per the standards. From the study it was observed that the value lies in the bad zone that is less than 50 but at some sites the value was just above the 50. The values of NSF WQI for different months of the two years are shown in the given. The lowest value was observed 36.92 in the month of June at S4 in the year 2014-2015 and 38.34 in the month of June at S1 in the year 2015 -2016. This is mainly due to high temperature, high influence of humans as the site is mostly invaded by locals, high metabolic activities of living organisms due to the

suitable temperature which results in the high accumulation of excretion. The site receives the waste water drain which also increases its pollution level. The highest range was observed 54.91 in the month of August at S1 in 2014 – 2015 and 52.54 in the month of January at S1 in the year 2015-16. This result is mostly due to low temperature.

CONCLUSION

CONCLUSION

In the present study, Benthic macro-invertebrates were identified which belongs to class Oligochaeta, Hirudinea, Gastropoda, Palacypoda, Insect and Crutaceans. We found most abundant species from the class Oligochaeta, Insecta and than from the class Gastropoda but shrimp & Miscellaneous were found less during the study period July 2014-June 2016.

The bottom fauna of this pond is qualitatively diversified and rich. All together 32 species of bottom fauna were collected and identified. They showed remarkable fluctuation both its quantity and quality in different season. The densities of oligochaetes and insects formed the main bulk consisting about 80% of the total collected benthos. The percentage abundance of oligochaetes was maximum in June and July while minimum in colder months. Insects, gastropods and ostracods showed their highest abundance during colder months and lowest during monsoon. However, variation in other groups were also registered but in haphazard fashion.

During biostatistical analysis of collected data, water temperature was correlated with DO but positive correlation of water temperature and phosphate as well as free CO₂. DO was found inverse relationship with Free CO₂. Inverse relationship such as Free CO₂ & pH, pH and phosphate but positive correlation with Free CO₂ and phosphate were exhibited.

Collected fishes of this pond were represented by 7 orders, 13 families, 17 genera and 23 species. Fishes were of all sizes but there were no any artificial manuring and introduction of fry and fingerlings and fishes could be recognized as the modest one.

Fish productivity of Mansarovar Talab was considered as very low due to lack of scientific culture. It was found that all abiotic and biotic components were still maintaining the pond productivity. The abiotic components of water are in the tolerable range of fishes.

In last, several suggestions are also made for improvement of aquaculture, specially fishes. On the adoption of mentioned suggestions and guideline the Mansarovar Talab will improve the fish yield. It will increase per capita fish consumption of the area and consequently it will improve socio-economic conditions of the localities specially of those who are engaged in fish farming. Besides, it will help in solving the problem of protein deficiency in food of the society.

***RECOMMENDATIO
NS
&
SUGGESTIONS***

RECOMMENDATIONS & SUGGESTIONS

Based on the observations made and experience gained during the course of this study, the following recommendation are made to improve the water quality and in- situ conservation of biodiversity of Mansarovar Talab.

Scientific studies in regard to in situ conservation should focus on the following lines where very little data are available.

- Applied research for conservation of living resources.
- Interlink ages between plant and animal species.
- Quantitative assessment of the conservation status of the species.
- Successional status of key species in different ecosystems.
- Multiplication and restoration of endangered, rare and endemic species using biotechnology.
- Ecological restoration of degraded micro and macro – habitats.
- Identification of critical index species and their sensitive parameters.
- Assessment of the impact of exotic species on the ecosystem.
- Determination of the impact on the ecosystem of various activities in the protected areas.
- The possible climate change and its impact on biodiversity.
- Hydrological change including surface run-off and percolation in the protected areas.
- Primary production and cycling of nutrients in the soil.
- Studies on satellite mapping of all protected areas.

- Development of methodologies for classification of microhabitats.

BIBLIOGRAPHY

- **Adakole, J.A. and Anunne, P.A. (2003):** Benthic macroinvertebrates as indicators of environmental quality of an urban stream, Zaria, Northern, Nigeria. *J Aquat. Sc.* 18 (2), 85-92.
- **Arimoro F. O., Ikomi R. B., Efemuna E., (2007):** Macro-invertebrate community patterns and diversity in relation to water quality status of River Ase, Niger Delta, Nigeria. *J Fish Aquat Sci.*, Volume 2(5), pp 337-344.
- **Akolkar P., Trivedi R.C., Makhijani S.D. And Sharma S.S. (1999):** Application of benthic macro-invertebrates as Biological Reference Material (BRM) for surface water quality management in India, DUREM-2, IInd.Natl. Workshop on Development and use of Reference materials, New Delhi, LATS/12/1999 29.
- **APHA (2002):** Standard method for examination of water and waste water, American Public Health Association Inc. New York 22nd Ed.
- **Begiraj S., Licaj P., Luotonen H., Adhami E., Hellsten S., Pritzl G., (2006):** Situation of benthic macroinvertebrates in Vjosa river- Albania and their relationships with water quality and environmental state. <http://balwois.com/balwois/administration/full-paper/ffp-1190.pdf>.
- **Berg, C. O. (1949):** Limnological relation of insect to plant of the genus *potomageton*. *Trans. Am. Micr. Soc.* 68: 279-291.

- **Berg, L. S. (1964):** Freshwater fishes of the USSR and adjacent countries, Vol. 2, Jerusalem.
- **Boyd C. E. (1982):** Water quality in Warm fish ponds, Auburn University Agricultural Experiment Station. Auburn University Agricultural Experiment station. Auburn, Alabama United states of America. 9-44.
- **Boyd S.E. (2002):** Guidelines for the conduct of benthic studies at aggregate sites. Reported the Centre for Environment Fisheries & Aquaculture. dredgng Lowestoft Laboratory, United Kingdom, pp 117.
- **Bachmann R.W. (2001):** The limiting factor concept: What stops growth Lakeline, Spring 2001: 26-28.
- **Bajpai R., (2012):**Comparative analysis of physicochemical parameters of Hasdeo river barrage and Arpa river water samples of Bilaspur region. *International Journal of Scientific and Research Publications*, Volume2, ISSN 2250-3153.
- **Barbhuyan S.I. & Khan A. (1992):** Studies on the structure & function of benthic ecosystem in eutropic body of water temperatureoral& spatial distribution of benthos. *J Fresh water Biology*. 4(4): 239-247.
- **Baruah, B.K. and Baruah, D. (2003):** Studies of water quality of Subanshi river in Assam-An approach for a proposed hydroelectric power project. *Indian. J. Env. &Ecoplan*, 7(2): 380-384.
- **Bhatnagar, G.P. (1984):** Limnological of lower lake of Bhopal with reference to sewage pollution and eutrophication; Technical report MAB program. Dept. of Environ. Govt. of India, New Delhi, pp 1 – 77.

- **Bhowmick, M. L. (1968):** Environmental factors affecting fish food in freshwater fisheries, Kalyani, West Bengal, Ph. D. Thesis, University of Kalyani.
- **Brinkhurst, R. O. and Cook, D. G. (1974):** Aquatic Earthworm (Annelida: Oligochaeta) In Pollution Ecology of freshwater invertebrate (Ed. Hart, J. and Fulter, T) Academic Press, New York. 143 - 155.
- **Brinkhurst R.O., (1974):** The benthos of lakes. *The Macmillan press. London and Basingstake*, 150.
- **Brinkhurst R.O., (1972):**The role of Sludge Worms in eutrophication U.S. Environmental Protection Agency, Washington. *D.C. Ecol, Res. Ser. EPA- R3-72-104*.
- **Brinkhurst R.O. (1970):** Distribution and abundance of tubificid (oligochaeta) species in Toronto Harbour, Lake Ontario. *J. Fish. Res. Bd. Can.*, Volume 27, pp 1961-1969.
- **Brinkhurst, R.O. (1965):** Studies on the North American aquatic Oligochaete I. *Naididae and Opistocytidae. Proc. Acad. Sci. Amer.* 16-20.
- **Brown R.M., Mclelland N.I., Deininger R. and Ronald G. (1970):** A water quality Index- Do we dare? *Water and Sewage Work* 339-343.
- **Camur-Elipek, B. Arslan, N. Kirgis, T. Otlerler, B. Guher, H. and Ozkan, N. (2010):** Analysis of benthic macro invertebrates in relation to environmental variables of Lake Gala. A national park in Turkey. *Turkish Journal of Fisheries and Aquatic Science* 10: pp 235-243.
- **Chattergee A.A.K. (1994):** Benthos, the water quality indicator in river Brahmani, Ind. J. Env. Hlth, Vol. 36, No. 1:24-30.

- **Cowell B.C. & Vodopich D.S. (1981):** Distribution & seasonal abundance of benthic sub-tropical Florida lake. *Hydrobiologia*. 78:97-105.
- **Chaudhari U., Johari S., Chaudhari P.R., (2001):** Trophic status of Chatri lake in Vicinity of Amravati city. *Indian J. Environ. Health.*, Volume 43(3), pp 135-137.
- **Carlisle D.M., Meador M.R., Moulton S.R., Ruhl P.M. (2007):** Estimation and application of indicator values for common macro-invertebrates. *Ecol. Indicators*, 7:22-23.
- **David, A., Ray, P., Govind, B. V., Rajagopal, K. V. and Banerjee, R. K. (1969):** Limnology and fisheries of Tungabhadra reservoir; *Bull. Cent. Inland fish Res. Inst.*, 13: 1 – 188.
- **Danes K.W., Hynes H.B.N. (1980):** Some effects of agricultural land use on stream insect communities. *Environ. Pollut. Ser. A* 22, 19–28.
- **Das. S.K. and Chand B.K. (2003):** Limnology and Biodiversity of Ichthyofauna in a pond of Southren Orrissa India. *J. Ecotox, Env. Monit* (13)2 97-102.
- **Dudgeon D. (1991):** An experimental study on the effects of predatory fish on macro-invertebrate in a Hong Kong Stream. *Fresh Water Boil.* 25(32):1-330. 29). 3031).
- **Edmondson, W. T. (1977):** Population dynamics and secondary production; *Ergeb. Limnol.* 8: 56 – 64.
- **Edmondson, W.T. (1971):** *Freshwater biology, ward and whipple* New York. John Wiley and Sons., Inc., London Chapman and Hall, Limited.
- **Edmondson, W. T. (1959):** *Fresh water Biology ward and Shipple* (2nd Ed.) Willey & Sons, New york, London, 12-48 PP.

- **Edokpayi C.A., Okenyi J.C., Ogbeibu A.E. and Osimen E.C. (2000):** The effect of human activities on the macro benthic invertebrates of Ibiekuma Stream, Ekpoma, Nigeria. *Biosci. Res.Comm.* 12: 79-87.
- **Egborge ABM (1986):** Physical, chemical and biological properties of Benin river. Technical Report for Cage fishery Development at Kolo submitted to Nr. J. Ayomike, P. 32.
- **Environmental Protection Agency., (2001):** Protecting and Restoring America's Watersheds: Status Trends and Initiatives in Watershed Management. EPA-840-R-00-001. pp 5-7.
- **Emere M.C. (2000):** A Survey of Macroinvertebrate fauna along River Kaduna, Kaduna, Nigeria, *J. Basic and Applied Sci.* 9: 17-27.
- **Esenowo I. K. and Ugwunba A.A.A. (2010):** Composition and abundance of Macrobenthos in Majidun River Ikorodu Lagos State. Nigeria. *Research Journals of Biological Science* 5(8): 556-560.
- **Ganapati, S. V. (1960):** Ecology of tropical waters. In. P. Kachroo (ed.), proceedings in the symposium on Ecology. Indian council of Agricultural research, New Delhi.
- **Ganapati, S. V. (1941):** Seasonal changes in the physical and chemical conditions of a garden pond containing abundant aquatic vegetation. *J. Madras Univ.*, 13: 55 – 99.
- **Garg S.S., (2003):** Water quality of well and bore well of ten selected locations of Chitrakoot region. IN: ARAVIND Kumar (Ed.), Toxicology, Daya publishing house, Dekhi, India, pp 114.

- **Gauvin, A.R. and Tarzwell, C.M. (1956):** Aquatic macro-invertebrate communities as indicator of organic pollution in Lytte, creek, sewage & industrial waste. 28(7): pp 906-924.114P.
- **George A.D.I., Abowei J.F.N. and Daka E.R. (2009):** Benthic Macro Invertebrate Fauna and Physico-chemical Parameters in Okpoka Creek Sediments, Niger Delta, Nigeria International Journal of Animal and Veterinary Advances, ISSN; 2041- 2908 1(2): 59-65.
- **George, J. P. (1976):** Hydrobiological studies on the lower lake of Bhopal with special reference to the productivity of economic fishes. Ph. D. Thesis Bhopal Univ. Bhopal.
- **Gerber, A. and Gabriel, M.J.M. (2002):** Aquatic invertebrates of South Africa rivers. Field Guide. Institute for Water Quality Studies. *Department of Water Affairs and Forestry*: 1-150.
- **Goldman, C. R. and Wetzel, R. G. (1963):** A study of primary productivity of clear lake, lake county, California, Ecology, 44: 283 – 294.
- **Golterman, H.L. Clymo R.S. and Ohnstad, M.A.M. (1978):** Method for physical & chemical analysis of fresh water. *IBP hand books no. 8 Black Well Scientific Publication*, Oxford 213pp.
- **Golterman, H. L. (1975):** Physiological limnology: An approach to the physiology of lake ecosystem. Elsevier Scientific Publication Comp. Amsterdam. Oxford, New York, 249 – 277.
- **Govind, B. V. (1978):** Planktonological studies in the Tungabhadra reservoir and its comparison with other storage reservoir in India; Proc. Semi. Eco. And fish fresh water reservoirs, 66 – 72.

- **Govind, B. V. (1969):** Bottom fauna and macrovegetation in the Tungabhadra reservoir and their role in the food chain of fish communities Proc. Sem. Ecol. Fish F. W. reservoir, sponsored by ICAR at CIFRI, Barrackpore November 27 – 29.
- **Gupta S.C., Rathore G.S., Mathur G.C.D., (2001):**Hydro-chemistry of Udaipur lakes. *Indian J. Environ. Health.*, Volume 43(1), pp 38-44.
- **Haase C.S. & Blodgett K. D. (2009):** The nature conservancy's mississippi River program: Sustainable Conservation of a Working River that works, in stevestarreett (ed.) (41036 edn. 342; Kansas City, Missouri: ASCE), 610-610pp.
- **Hamaidi-Chergui F., Errahmani M.B., Benouaklil F., Hamaidi M.S., (2013):**Preliminary Study on Physico-Chemical Parameters and Phytoplankton of Chiffa River (Blida, Algeria). *Journal of Ecosystems Article ID 148793*.
- **Hart A.I. and Zabbey N. (2005):** Physico-chemistry and Benthic Fauna of Woji Creek in the Lower Niger Delta, Nigeria. *Environment and Ecology* 23(2): 361-368.
- **Hellawell J.M. (1986):** Biological indicator of fresh water pollution and environmental management. Pollution monitoring series. Advisory editor: Kenneth Mellanby, England 546.
- **Hynes K.E. (1998):** Benthic macro-invertebrate Diversity & Biotic indices for monitoring of 5 urban urbanizing lakes within the Halifax Regional Municipality (HRM) Nova Scotia, Canada. Soil & water conservation society of Metro Halifax. XIV: 114.
- **Idowu E.O. and Ugwumba A.A.A. (2005):** Physical, chemical and benthic faunal characteristics of a Southern Nigeria Reservoir. *The Zoologist*, 3: 15-25.

- **Indabawa I.I. (2010):** The assessment of water quality at Challawa River via physico-chemical and macro-invertebrate analysis, bioscience research communications Vol.22: 227-233pp.
- **Jadhav S.D., Kanase D.G., Lawale R.W., Jadhav M.S., (2006):**Physico chemical assessment of Krishna River at Kharad, Satara district, Maharashtra.
- **Jain P.,Shrivastva, (1998):** Heavy metal contamination in Kerwan dam water at Bhopal, India. Journal of Ecobiol., Volume 10, pp 63-66.
- **Jayaram, K.C. (1994):** The fresh water fishesh of India, Pakistan, Bangladesh,Burma and Srilanka, Zoological Survey of India, Calcutta.475.
- **Jenkin, P. M. (1942):** Seasonal changes in the temperature of Windermere (English lake district); J. Anim. Ecol., 11: 248 – 269.
- **Jhingran, V. G. (1992):** Fish and Fisheries of India. 3rd Edn. Hindustan Publ. Corp. India Ltd. New Delhi.
- **Jhingran V.G. (1982):** Fish and Fisheries of India. Hindustan Publishing Corporation (India), Delhi: 1-666.
- **Karr J.R. and Chu E.W. (1999):** Restoring Life in Running Waters: Better biological monitoring. Island Press. Washington, District of Columbia, U.S.A. 206.
- **Khanna D.R. & Bhatia A. (2003):** Limnological characteristics of the river Ganga at Haridwar, Uttaranchal. U.P.J. Zool. 23(3) 179-18.
- **Khanna D.R. and Vats Deepika (2006):** Macrobenthic diversity and Its ecology of river Ganga at foot hills of Garhwal Himalaya J. Exp. Zool. India 9 (1) 125-130.

- **Koperski, P. (2010):** Diversity of macro invertebrates in low land streams ecological determinants and taxonomic specificity. *J. Limnol.* Vol. 69(1) pp 88-101.
- **Krishna J.M., (2012):** Physicochemical and bacteriological study of Kaveri river at Kudige, Kodagu District, Karnataka. *International journal of Environmental Sciences*, Volume 2, pp 4.
- **Kumar A. & Singh A.K. (1997):** Role of macrobenthos in monitoring the water quality of the river Mayrakshi in South Bihar. *J. Mondel* 15: pp 145-151.
- **Kumar K. (2003):** Bioassessment of water quality of river Yamuna using benthic macro-invertebrates MSc. Thesis Delhi University.
- **Lindman, R. L. (1942):** The trophic dynamic aspect of ecology; 23: 399 – 418.
- **Lund, J. W. G. (1972):** Eutrophication; *Proc. R. Soc. London Ser. B*, 180: 371 – 382.
- **Lydeard C., Cowie R.H., Ponder W.F., Bogan A.E. and Bouchet P. (2004):** The global decline of Non-marine mollusks. *Bioscience*, 54: 321-330p.
- **Mahendra Pal Sharma, Arun Kumar, Praveen Sharma, Vivel Goel. And Shailendra Sharma (2010):** Water quality assessment of Kosi River, using BMI as Bioindicator. *Proceeding of IInd International river festival – March 2010*: 41-42.
- **Mahapatra, Y.R. & Panday (2001):** Seasonal fluctuation of physico-chemical parameters of Rushikulya Estuary, Bayt. Of Bengal. *Indian J, Environ. &Ecoplan* 5(1) 35-40.

- **Manjare S.A., Vhanalakar S.A., Mulley D.V., (2010):**Analysis of water quality using physico-chemical parameters at Tamdolge tank in Kolhapur Maharashtra. *International Journal of Advance Biotechnology and Research*, Volume 1(2), pp 115-119.
- **Michael, R. G. (1968):** Studies on the bottom fauna in a tropical fresh water fish pond *Hydrobiologia*, 31 (1): 203-229.
- **Michail, R.G. (1980):** A historical resume of Indian Limnology. *Hydrobiologia* 72. 15-20.
- **Mortimer, C. H. (1942):** The exchange of dissolved substance between mud and water in lakes part II. *J. Ecol.* 30: 147 – 201.
- **Muhammad Zaheer Kahn (2006):** impact of climate changes and environmental pollution an ecosystem and wild life. *J. Exp. Zool. India* 9 (1) 77-84.
- **Munawar, M. (1974):** Limnological studies of fresh water ponds of Hyderabad, India IV. The biocenose periodicity and species composition of unicellular and colonial phytoplankton in polluted and unpolluted environments; *Hydrobiologia*, 45 (1): 1 – 32.
- **Munawar, Mohiddin. (1970):** Limnological studies of fresh water pond of Hyderabad, India, *Hydrobiologia*, 39 (1): 127-162.
- **Murhekar G.H., (2011):** Determination of Physico-Chemical parameters of Surface Water Samples in and around Akot City. *International Journal of Research in Chemistry and Environment.*, Volume 1, pp (183-187), ISSN 2248-9649.
- **Mustow S. E. (1996):** 'Biological monitoring of rivers in Thailand: use and adaptation of the BMWP score', *Hydrobiologia*, 479 (1) 191-229.

- **Naik., Shrikanta., Purohit K.M., (2001):** Studies on water quality of river Brahmi in Sundergarh district Orissa, India. J. Env. 7 Ecoplan, Volume 5 (2), pp 397-402.
- **Needham, J.G. & Needham, P.R. (1969):** A guide to the study of fresh water biology Holden – Day Inc. Sanfranhisco, 108. Ruttener, F: 1953. Fundamentals of Limnology. *Publ. E.E.J. Unic. Press*, Toronto 242.
- **Negi, R.K. Negi, Tarana& Joshi, P.C. (2008):** Study on physico-chemical parameters of Hinval fresh water stream & Ganga river of Shivpuri in the Garhwal Region. J. Env. Bio. Sci. Vol.22 (2) 203-212.
- **Nesemann, H. Sharma, G. and Sinha, R.K. (2003):** The Bivalvia species of the Ganga river and adjacent stagnant water bodies in Patna, (Bihar, India) with special reference on Unionacea. *Acta Conchyliorum* 7:1-43, Ludwigsburg, Wein.
- **Nesemann, H. Sharma, S. Sharma, G. Khanal, S N. Pradhan, B. Shah, D N. and Tachamo, R.D. (2007):** Aquatic Invertebrates of Ganga river system. (Mollusca, Annelida, Crustacea (in part) *first edition*. ISBN 978-99946-2-674-8.
- **Nkwoji, K. J. Yarub, A. Ajani, G.E. Ariyo, A. A. and Bello, B. O. (2010):** Seasonal variations in water chemistry and benthic macro invertebrates in South Western lagoon, logos, Nigaria. *Journal of American Science*; 6(3) pp 85-92. (ISSN: 1545-1003).
- **Odum, E. P. (1971):** Fundamentals of ecology 3rd Ed. Saunders, Philadelphia.
- **Ogbeibu A.E. (2001):** Distribution Density and Diveristy of Dipterans in a Temporary pond in Okomu Forest Reservoir, Southern Nigeria. J. Aqu. Sci. 16: 43-52.

- **Ogbogu S.S. and Olajide S.A. (2002):** Effect of Sewage Oxidation pond effluent on Macroinvertebrate communities of a Tropical Forest Stream, Nigeria. *Journal of Aquatic Science*, 17(1) 22–27.
- **OgidiakaEfe (2012):** Physico-chemical parameters and Benthic Macro-invertebrates of Ogunpa River at Bodija, Ibadan, Oyo State. *European Journal of Scientific Research*. Vol. 85(1) 89-97.
- **Oomachan, L. and Belsare, D.K. (1980):** Bathymetric distribution of Mollusca in Lower Lake of Bhopal. *Bull. Bot. Soc. Univ Sagar*. 32: 109-113.
- **Olomukoro J. O. and Egborge, A. B. M. (2003):** Hydrobiological studies on Warri, River, Nigeria Part 1: The Composition, distribution and diversity of macrobenthic fauna. *Biose. Res. Commun*. 15: 279-296.
- **Pandey S., (2007):** Biodiversity of benthic macro-invertebrates and their tropic relationship with shore birds and fishes of Yashwant Sagar reservoir, Indore, (M.P.). Ph.D. Thesis, Devi Ahilya Vishwavidyalaya, Indore, (M.P.).
- **Pandey, A.K. Dutta, S. and Sharma, K.C. (2002):** physico-chemical characteristics of Hamir pond of Kishangarh, Ajmer (Rajasthan), Indian *J. Environ and Ecoplan* 6(1):167-169.
- **Pathak S.K., Mudgal L.K., (2005):** Limnology and biodiversity of fish fauna in Virla reservoir, MP India. *J. Comp. Toxicol. Physiol.*, Volume 2(1), pp 86-90.
- **Pathak, S.K. (2004):** Limnology status of virla reservoir, district-west nimar (Khargone) M.P. India, A. Ph.D. thesis, D.A.V.V. Indore.

- **Patil S.G., Harsney, D.K. & Singh D.F. (1984):** Benthic organism as indicators of pollution in lentic & lotic Environment Geobies 11: pp 77-80.
- **Pennak Robert W. (1989):** Fresh-water invertebrates of the United States: Protozoa to Mollusca. 3rd. ed. John Wiley and Sons, New York.
- **Praveen Sharma & Shailendra Sharma (2008):** Rapid field bioassessment of the rivers using benthic macro-invertebrates as indicators. Presented in International Conference in international River festivals, Hoshangabad. (February, 23-25,2008). Conference proceeding, pp 68.
- **Prasum T., Arora M.P., Akolkar P., Tyagi R. and Arora A. (2006):** Occurrence of benthic macro-invertebrate families encountered in river Hindon in Uttar Pradesh (India) J. Exp. Zool. India 9(1) 209-216.
- **Roa, K.S. Dad, N.K. & Pandya, S.S. (1985):** Community structure of benthic macro-invertebrates & their utility as indicators of pollution in river Khan (Indore), India. Proc. Nat. Symp. Pure. & Appl. Limnology 32:114-119.
- **Rao, K. S. and Shrivastava, S. (1989):** Studies on biological monitoring of water quality in chamber a Khan river of Central India Giobios, 16:78-82.
- **Rao, N. G. (1987):** Synecology of the lake Rangasagar in relation to the limnology and eutrophication Ph. D. Thesis Sukhadia Univ. Udaipur.
- **Rao, K. S., Gupta, D. K., Choubey, V. and Sreenivasan, P. (1987):** Benthic studies in Gandhisagar reservoir with special reference to macrobenthos perspectives in Hyderabad. Sci. IV; 34: 173 - 178.

- **Rosenberg D.M. and Resh V.H. (1993):** Freshwater Biomonitoring and Benthic Invertebrate. Chapman and Hall, New York. Chapman & Hall. 488.
- **Ruttner, F. (1963):** Fundamentals of limnology. English translation by D.G. Frey and F.E.J. Fry. Toronto University Press, Canada. 307 pp (First German edition in 1940).
- **Sandwar, B.B. & Prasad. J. (2000):** Physico-chemical characteristics of Ganga river at Makamah, Bihar. In (Trivedi, R.K. Edited) pollution & Biomonitoring of Indian Rivers. ABP. Pub: pp 130-132.
- **Sarkar A., (2012):** Bioindicators of River Yamuna at Agra International Journal of Geology and Environmental Sciences, Volume 2 (1), ISSN: 2277- 2081, pp 16-21.
- **Saxena, A. K., Tiwari, R. K. and Kulshrestha, S. K. (1988):** Population structure of certain Ephemeropterans in the upper and lower lakes of Bhopal. Proc. Nat. Symp. Past, Present and Future of Bhopal Lake, 49 - 54.
- **Singhal P.K. (1991):** Distribution & abundances of the macro benthic fauna of certion semi-Arid fresh water bodies. Ph.D. Thesis Univ. of Jodhpur, India.
- **Sinha, M. P., Sinha, R. and Mahato, P. N. (1991):** Composition and dynamics of freshwater macrobenthic community I oligochaete oikoassay 8: 21-24.
- **Sinha, R. K. and Das, N. K. (1993):** Organic waste and effect of macro – zoobenthos in ganga at Patna (Bihar) Indian J. Fershwater. 5: 33-40.

- **Sinha R. Ojha N.K., Das S.K. & Sinha, M.P. (1997):** Community composition, population dynamics & species diversity of macrobenthic fauna of a tropical fresh water lentic body. In Recent advances in Ecobiological Research Vol. I. (Ed, M.P. Singh) APH Publishing Corporation New Delhi.
- **Sharma S., Barkale S., (2016):** The Species Richness and Abundance of Macro-Invertebrates In bilawali Talab, Indore (M.P.), India. International Journal of Fisheries and Aquatic studies, Volume 4(5), pp 311-315.
- **Sharma S., Barkale S., Patel S. B., (2015):** Biological indices of ecosystem. Limnology and Fisheries Discovery Publishing House Pvt. Ltd., New Delhi; ISBN: 978-93-5056-731-8 (157-177).
- **Sharma S., Dubey S., Chaurasia R., Solanki S., Dave V., (2015):** Diversity of water beetles (coleopteran) in Tropical and Sub tropical region (Khlaghat) of Narmada River, M.P. India. Limnology and Fisheries Discovery Publishing House Pvt. Ltd., New Delhi; ISBN: 978-93-5056-731-8, pp 1-16.
- **Sharma S., Dawar B., Barkale S., (2013):** Biomonitoring a biological approach to water quality management. Elixir Bio. Diver., Volume 66 (2014), pp 20635-20638.
- **Sharma S., Dubey S., Chaurasia R., (2013):** The insect diversity of river Kunda, Khargone district M.P. (India), G.J.B.B., Volume 2, pp 454-455.
- **Sharma S., Dubey S., Chaurasia R., Dave V., (2013):** Macro-invertebrates community diversity in relation to water quality status of Kunda river (M.P.) India; Discovery, Volume 3 (9), pp 40-46.

- **Sharma S., Pandey P., Dave V., (2013):**Role of aquatic beetles for water quality assessment International journal of Recent Scientific Research, Volume 4, Issue 11, pp 1673-1676, ISSN; 0976-3031.
- **Sharma S., Pandey P., Dave V., (2013):**Abundance of benthic macro-invertebrate diversity of littoral zone of Bilawali Talab, Indore M.P. (India). Elixir Bio. Diver, Volume 62, pp 17825-17827.
- **Sharma S., Patel B., Barkale S., Dave V., (2013):**Benthic macro-invertebrates use as a tool of water quality assessment of fresh water resources. The Journal of Ecology. Photon 107(2013)216-225. ISJN: 6853-3275.
- **Sharma S., Zahoor Pir, ImtiyazTali., Mudgal L.K., (2013):** Molluscs can decide the water quality of Rivers. International Journal of current research, Volume 5, pp 405-405.
- **Sharma S., Dubey S. and Chaurasia R. (2013):** Benthic Macro-invertebrate abundance and its Correlations with physico-chemical parameters from Kunda River, Khargone (M.P.), India. International Journal of Advanced Research Vol. 1(2) 8-13.
- **Sharma, S. Mudgal, L.K. Mimrot, K. (2012):** Role of species diversity of aquatic benthic macro invertebrates for assessment of water quality of river Narmada (M.P.), India.
- **Sharma S., Tali I., Pir Z., Siddique A., Mudgal L.K., (2012):** Evaluation of Physico- chemical parameters of Narmada River, M.P., India Researcher, Volume 4(5), pp 13-19. (ISSN: 1553-9865) <http://www.science pub.net / researcher>.
- **Sharma K.K., Chowdhary S., (2011):** Macro-invertebrates assemblages as biological indicators of pollution in a Central Himalayan River, Tawi (J & K) International Journal of Biodiversity and Conservation, Volume 3 (5), pp 167-174.

- **Sharma S. and Sharma P. (2010):** Biomonitoring of aquatic ecosystem with concept and procedures particular reference to aquatic Macro-invertebrates. Jour. of American Science, 6 (12) 1246-1255.
- **Sharma Shailendra et.al (2008):** The effect of environmental conditions on the taxonomic and functional structure of benthic macro invertebrate communities in the Hindu Kush-Himalayan region. Proceedings of the Scientific Conference Rivers in the Hindu Kush-Himalaya – Ecology & Environmental Assessment. Katmandu University Nepal (3rd to 5 th March- 2008).
- **Sharma P., Sharma S., (2008):** Rapid field bioassessment of the rivers using benthic macro-invertebrates as indicators. Presented in International Conference in International River Festivals, Hoshangabad. Conference proceeding, Volume 68.
- **Sharma, Shailendra M.P. Sharma, Vivek Goel, Praveen Sharma and ArunKumar. (2008):** Water quality assessment of Ninglad stream using Benthic macroinvertebrates', Life Sciences Journal (China) (Vol.5) No.3. pp 67-70.
- **Sharma S., Joshi V., Kurde S., Singhavi M., (2007):** Biodiversity of benthic macro-invertebrates & fish species communities of Krishnpura lake, Indore, M.P. J. Aqua. Biol., Volume 22 (1), pp 1-4.
- **Sharma, M.P., Sharma, S., Goel, V. Sharma, P. Kumar, A. (2006):** Water quality assessment of Behta river using benthic macro-invertebrates. Life Sc. Journal, 3(4): 68-74.

- **Sharma S., (2003):** Biodiversity of littoral benthic organism & their tropical relationship with shorebirds & fishes in Krishnpura lake Indore M.P., D.A.V.V.
- **Sharma S., (2002):** Biodiversity of littoral benthic organism and their tropical relationship with shorebirds and fishes in Krishnpura lake. *Ph.D. Thesis*, D.A.V.V. Indore (M.P.).
- **Shah K.A. & Pandit A.K. (2001):** Macro invertebrates associated with macrophytes in various fresh water bodies of Kashmir. *Journal of Research & Development*. I: pp 44-53.
- **Shrivastava, S. Roa, K.S. & Shukla, A.N. (2001):** Benthic macro-invertebrate fauna & feeding relationship of Catfish from tropical Kshipra river (M.P.) India. *Pakistan J. Zool.*, 33(4): 299-306.
- **Srivastava G.J. (1980):** Fishes of Eastern Uttar Pradesh. VishwavidyalyaPrakashan, Varansi pp.207.
- **Shukla, A. & Shrivastava, S. (2004):** Species diversity of macro zoobenthos: A tool for monitoring water pollution of Gandhi Sagar Reservoir. M.P. India. *Biol. Memoir*. 30(1): pp 7-13.
- **Siraj S., Yousuf A. R., Bhat F.A., Parveen M., (2010):** The ecology of macro zoobenthos in Shallubugh Wetland of Kashmir Himalaya, India. *Journal of Ecology and the Natural Environment*, Volume 295, pp 84-91.
- **Sreenivasan, A. (1974):** Limnological features of a tropical Impoundment Bhavanisagar reservoir (Tamil Nadu) India; *Int. Rev. Ger. Hyrdobiol.*, 59 (3): 327 – 342.
- **Strayer D. (2008):** Twenty years of zebra molluscans, lessons from the mollusk that made headlines. *Frontiers in Ecology and the Environment*, 7(3) 135–141.

- **Sultana R. and Kala D.S. (2012):** Water body quality analysis by Benthic Macro-invertebrates. International Journal of Pharmacy and Biological Sciences Vol. 2 269-279.
- **Talwar, P. K. and Jhingran, A. G. (1991):** Inland fishes of India and adjacent countries. Oxford and IBH Publishing Co. Pvt. Ltd., New Delhi.
- **Tonapi G.T. (1980):** Fresh water animals of India-an Ecological approach. Oxford and IBH Publishing Co. New Delhi; 341.
- **Trivedy R K and Goel P K, (1986)**Chemical and Biological Methods for Water Pollution Studies, Environmental Publication, India.
- **Vasisht, H. S. and Bhandal, R. S. (1979):** Seasonal variation of benthic fauna in some North Indian lakes and ponds. Ind. J. Ecol. 62: 33-37.
- **Vasisht, H. S. (1968):** Limnological studies of Sukhna lake Chandigarh (India). Proc. Symp. Recent Adv. Trop. Ecol., (part I): 316 – 325.
- **Unni, K. S. (1985):** Comparative limnology of several reservoirs of central India Int. Rev. Ges. Hydrobiol. 70 (6): 845 – 856.
- **Unni, K. S. (1972):** An ecological study of macrophytic vegetation of Doodhadhari lake, Raipur. Part III, Chemical Factors. Hydrobiol. Vol. 39.
- **Unnisa S.A. & Khalilullah M. (2004):** Impact of industrial pollution on ground and surface water quality in the Katledan industrial area. J. of Indian Association for Environmental Management, 31: 77-80.
- **Voelz NJ.** The effect of urban areas on benthic macroinvertebrates in two Colorado plains rivers. Environmental Monitoring and assessment. **2005**; 101:175-202.

- **Welch E.D., Jacoby J.M. and Christopher W.M.(1998):** Stream Quality, In: River Ecology and Management from the Pacific Coastal Eco-region, Springer-Verlag Publication. New York, 69-94.
- **Welch, P. C. (1948):** Limnological Methods, Blakiston, Philadelphia.
- **Welch,P.S. (1952):** Limnological methods McGraw Hills Book. Company, New York. 381 PP.
- **Welch, P. S. (1952):** Limnology, McGraw Hill Co: 4-538.
- **Welch, P.S. (1998):** Limnology. *McGraw Hill Book Co, New York*, pp 538.
- **Wetzel. R.G. (1970):** Limnological Analysis II ed. Sprihger-Verluag New York.
- **Wetzel. R. G. and Likens, G. E. (1971):** Limnological Analysis IInded. Sprihger – Verluag New York.
- **Wetzel R.J., (1975):** Limnology W.B. Saunders Company Philodelphia, pp 743.
- **William DP, Feltmate BW.** Aquatic insect C.A.B. *International, Wallingford Oxon, UK.1992.*
- **Williams, L.R. & Taylor, C.M. (2003):** Influence of fish predation on assemblage structure of macro-invertebrates in an intermitted stream. Transaction of the American Fisheries. Soc. 132: 120-130.
- **Yadav R.C., Srivastva V.C., (2011):** Physico-chemical properties of the water of River Ganga at Ghazipur. Indian J. of Sci. Res., Volume 2(4), pp 41-44.

- **Zabbey N., Hart A.I., (2005):**Physico-chemical and benthic fauna of Woji Creek in the Lower Niger Delta, Nigeria. Environment and Ecology, Volume 23(2), pp 361-368.